Synthesis and Pharmacological Evaluation of Cyclodextrin Conjugate Prodrug of Mefenamic acid

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In the present investigation mefenamic acid prodrug of β -cyclodextrins was synthesized. The primary hydroxy group of β -cyclodextrins was used to block the acid group. The synthesis involved a series of protection and deprotection reaction. The ester was evaluated for stability in simulated gastric and intestinal fluid. The hydrolysis of cyclodextrin conjugate in colon is confirmed by the hydrolysis kinetics studies in rat faecal material. The ester was also evaluated for ulcerogenicity. Results of these studies established the primary aim of masking the ulcerogenic potential of free drug, by using 12-fold dose of the normal dose of mefenamic acid and equivalent doses of the ester.

Cyclodextrins (CDs) belongs to family of cyclicoligosaccharides; the most common being α -, β - and γ -CDs consisting of 6, 7 and 8 glucopyranosyl units, respectively, linked by a (1->4) glucosidic bonds¹. CDs are obtained by enzymatic degradation of starch, glucosyl transferase a type of amylase of bacterial origin obtained usually from Bacillus macerans, Bacillus megaterium, Klebsiella pneumoniae M5, and Bacillus sterothermophillus. CDs are moderately soluble in water, methanol and readily soluble in strongly polar aprotic solvents². After oral administration, CDs are not hydrolyzed during their transit through the stomach, hydrolysis occurs only in colon by colonic microflora. The oral administration of CDs does not result in toxicity^{3,4}. Thus, CDs were thought to be one of the most suitable promoieties to reduce the ulcerogenic tendencies of mefenamic acid since they eliminate the exposure of free drug in stomach and small intestine but release the drug in colon.

Mefenamic acid is a nonsteroidal antiinflammatory and analgesic compound belonging to the family of N-aryl anthranilic acid. It is also described as 2-[(2,3-Dimethylphenyl) amino] benzoic acid. The main side effects include GIT disturbance, peptic ulceration and gastric bleeding. Mefenamic acid is widely used in rheumatic disorders such as ankylosing spondilytis, osteoarthritis and rheumatoid arthritis.

The major factor, which limits the use of mefenamic acid, is

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its gastric effect due to local irritation of gastric mucosa by free -COOH group of the drug⁵⁻⁷. Hence efforts have been made to mask the free -COOH group by the primary hydroxyl group of CD and releasing the drug in colon and preventing the exposure of free drug to the stomach.

MATERIALS AND METHODS

All melting points were determined by open capillary method and are uncorrected. TLC ascertained the purity of the compounds on precoated silica gel-60 F₂₅₄ plates. Solvent used was butanol:ethanol:water:acetic acid (3:2:3:0.1). Thus different spots for reference and test substance were detected using iodine vapours or by charring the plates using 5% methanolic sulfuric acid. The final compound was re-crystallized from water, after extracting the impurities with ethyl acetate. The IR spectrum of the synthesized compound was recorded on a FT-IR spectrophotometer, in potassium bromide (anhydrous IR Grade) pellets. The NMR spectra of the synthesized compounds were determined on a FT-NMR spectrophotometer. The $\lambda_{\mbox{\tiny max}}$ of the synthesized compounds was determined on UV/Vis double beam spectrophotometer by scanning the compounds between 400-200 nm in various solvents. Mefenamic acid was obtained as a gift sample from P&B Laborotries, Mumbai. CDs were obtained as gift sample from S. A. Pharmachem, Mumbai. All the other chemicals used were of synthetic grade.

Mefenamic acid ester of β CD (SA-001):

Synthesis of mefenamic acid ester of β -CD involved 5 steps. First step involved tritylation of one of the primary

hydroxyl groups of β-CD (0.0044 mol, 5 g). This reaction involved reacting the β-CD with trityl chloride (a) (0.00484 mol, 1.28 g) in pyridine (30 ml) by stirring it for 24 h⁸. The residue obtained was refluxed with n-hexane to remove trityl alcohol, the by-product. The solid was dried under vacuum to give tritylated β-CD (5.44 g). Next step involved acetylation of tritylated β-CD (0.00366 mol, 5 g) using acetyl chloride (b) (0.072 mol, 5.18 ml) and triethyl amine (c) (0.072 mol, 10.04 ml) in ethylene dichloride (75-80 ml). The temperature was maintained at 0° throughout the reaction. The reaction mixture was filtered and dried under reduced pressure to give acetylated monotritylated β-CD (6.0 g)⁹.

Acetylated monotritylated β-CD (0.0036 mol, 6 g) was dissolved in N, N-dimethyl formamide (DMF, 200 ml) in round bottom flask. The cation exchange resin *T-63 (MP) (d) (3 g) was added slowly into the reaction flask. The reaction mixture was stirred at room temperature and TLC ascertained reaction completion. The reaction mixture was concentrated under reduced pressure to avoid charring of the carbohydrate material. The residue was refluxed with n-hexane in order to remove trityl alcohol liberated during reaction. After complete removal of trityl chloride traces of solvents were removed under vacuum. The residue obtained was subjected to esterification reaction. [*T-63 (MP) Tulsion Thermax is a catalyst grade strong acid cation exchange resin]

Selectively detritylated B-CD (0.002 mol, 4 g) was dissolved in DMF (50 ml) in a round flask, 1,3-dicyclohexyl carbodiimide (DCC) (e) (0.0028 mol, 0.576 g) was dissolved in DMF. Both the solutions were ice cooled to 0°. The DCC solution and mefenamic acid free acid (f) (0.0028 mol, 0.675 g) were added to CD solution¹⁰ at 0°. The reaction mixture was stirred at 0° for 2 h and then at room temperature for 12 h. The reaction mixture was then filtered to separate the precipitate of N, N-dicyclohexyl urea, the by-product. The filtrate was concentrated under reduced pressure, to give acetylated mefenamic acid ester of β-CD (3.52 g). The ester was deacetylated by dissolving the ester (0.0018 mol, 3 g) in methanol at room temperature and reacting with ethylenediamine (g) (0.0018 mol, 10.8 ml) and triethylamine (c) (0.0018 mol, 18.18 ml)¹¹. The reaction was stirred for 5 h and large amount of acetone (400 ml) was added to precipitate the deacetylated ester. Molecular formula of β-CD ester of mefenamic acid (SA-001) is $C_{57}H_{82}NO_{36}$. Melting point of SA-001 is 245-248°. TLC; $R_{\rm c}$ 0.58; butanol:ethanol:water:acetic acid (3:2:3:0.1) and λ_{max} in distilled water: 284.0 nm and 310.0 nm, in 0.05 M HCl buffer (pH 1.2): 0.05 M phosphate buffer (pH 7.4): 340 nm. The chemical shift was in the ppm relative to TMS

designated as 0.00. NMR (DMSO- d_6) revealed the presence of signals in ppm at δ =2.2 (6H, s, CH₃), 4.48-4.47 (6H, m, CD 6-OH) and 7.6-6.62 (aromatic protons) for SA-001. Schematic representation for synthesis of SA-001 is shown in Scheme 1.

Hydrolysis kinetics:

Hydrolysis of β-CD ester was studied in simulated gastric fluid and simulated intestinal fluid (0.05 M HCl buffer, pH 1.2 and 0.05 M phosphate buffer, pH 7.4). The ester showed negligible release in both the hydrolysis media. The hydrolysis of the ester was studied in rat fecal material (pH 7.4) to confirm the colonic hydrolysis of the esters 12. The release of drug in rat faecal material was almost complete. The results of hydrolysis in rat faecal content are quoted in Table 1.

Release studies in 0.05 M HCl buffer, pH 1.2 and 0.05 M phosphate buffer, pH 7.4:

Same procedure was followed for the synthesized ester, ß-CD ester was dissolved in HCl buffer and in phosphate buffer so that the final concentration of the ester is equivalent to 1 µg/ml of mefenamic acid. To each of the flasks of dissolution apparatus (Veego Scientific DA 6D model) 900 ml of buffer was added when the temperature reached 37±1°C the ester solution was added to the flask

Scheme 1: Synthesis of β cyclodextrin ester of mefenamic acid SA-001

The reagents for each step are trityl chloride (a), acetyl chloride (b) and triethylamine (c), T-63 MP resin, (d), Mefenamic acid free acid and, (e), 1,3-dicyclohexyl carbodiimide, (g) ethylenediamine and triethylamine. n is 7 for SA-001.

TABLE 1: HYDROLYSIS KINETICS OF SA-001 IN RAT FAECAL MATTER

Compound	ound % Drug released with various time intervals in minutes									
	15	30	60	90	120	150	180	210	240	24 h
SA-001	0	5.48	10.7	20.72	32.44	54.2	63.0	75.5	79.2	85.0

The hydrolysis of SA-001 in rat faecal contents was studied with time (n = 6)

and stirred at 100 rpm. The ester showed negligible release in both the hydrolysis media.

Release study in rat faecal matter, pH 7.4:

The ester was dissolved in phosphate buffer (pH 7.4) so that final concentration of solution was 250 μ g/ml. Fresh fecal material of rats was weighed (about 1 g) and placed in different sets of test tubes. To each test tube, 1 ml of the ester solution was added and diluted to 5 ml with phosphate buffer (50 μ g/ml). The sets of test tubes were incubated at 37° for different intervals of time. For analysis the free drug was extracted in 5 ml CHCl₃ and directly estimated on double beam UV-spectrophotometer (Jasco, V-530 model, Japan). The concentration of free drug was determined using K and b values obtained from calibration curve.

Antiinflammatory activity:

The animal study protocols have met with the Institutional Animal Ethics Committee's (IAEC) approval. Carrageenaninduced rat hind paw oedema method was used for determining antiinflammatory activity¹³. Sprague Dawley rats of either sex (150-200 g) were taken in groups of six animals each. The synthesized compound was suspended in 1% solution of carboxy methyl cellulose (CMC) in distilled water. For control 1% solution of CMC in distilled water was given orally. Mefenamic acid 20.0 mg/kg was used as reference drug. Thirty minutes after the drug administration, 0.1 ml of 1% w/v carrageenan solution was injected in the plantar region of the left hind paw of the animals. The inflammation was determined using a plethysmograph 3 h after injecting the phlogistic agents and compared with that of the control. The data was analyzed using student's "t" test and the level of significance was defined at p<0.05. The results are summarized in Table 2.

Analgesic activity:

The analgesic activity was evaluated by acetic acid induced writhing method¹⁴. Albino mice of either sex with a weight range between 20 and 25 g were used. Acetic acid (0.6%) was used as irritant and 0.25 ml of this solution was injected intraperitoneally. An aliquot of groups of 6 animals were used for control and treated mice. Test animals were administered the drug or the

TABLE 2: ANTIINFLAMMATORY, ANALGESIC AND ULCEROGENIC ACTIVITY OF SA-001

Compound	Antiinflam activ (% inhib of oede	ity oition	Analgesic activity (%)	Ulcer index
	6h	24h		
Control	NIL	NIL	NIL	NIL
Mefenamic acid	21.49	08.08	70.2±3.06	45.0
SA-001	44.65	52.03	17.8±5.85	04.0

SA-001, Mefenamic acid (20.0 mg/kg or equivalent) and vehicle were administrated orally to Wistar rats and corresponding parameters were measured after specific time intervals after inducing either inflammation or hyperalgesia with carrageenan

standard at various pretreatment times prior to administration of acetic acid. The mice were placed individually into glass beakers and five minutes were allowed to elapse. The mice were then observed for a period of ten minutes and the number of writhes was recorded for each animal. For scoring purposes, a writhe is indicated by stretching of the abdomen with simultaneous stretching of at least one hind limb. The time period with the greatest percent of inhibition is considered the peak time. Compounds with less than 70% inhibition are considered to have minimal activity. The results are given in Table 2.

Ulcerogenicity:

Ulcerogenicity was determined by the method reported by Rainsford *et al.*¹⁵ According to this method sensitivity of gastric mucosa to ulcerogenic -COOH group is increased by exposing the animals to a short period of physical stress (i.e., cold or restraint)¹⁵. Wistar rats of either sex (150-200 g) were selected. They were kept for fasting for 24 h. Following oral administration of drug that is 10 times the normal dose in case of test and vehicle dose in case of control, the animals were stressed by exposure to cold (-15° for 1.5 h). The animals were sacrificed 3 h after drug administration and the number and severity of gastric mucosal lesions were determined. The results are summarized in Table 2.

RESULTS AND DISCUSSION

The studies have revealed that the CD ester of mefenamic acid has retained its pharmacological activity in

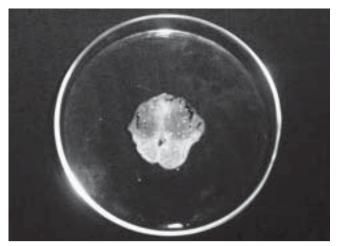




Fig. 1: Photographs of ulcers produced by mefenamic acid and SA-001

Top panel shows the ulcerogenic effect of mefenamic acid treatment while the bottom panel depicts that after treatment with SA-001

terms of % inhibition of oedema and in acetic acid induced writhing method and it was compared with that of its parent compound. After 1 h all the animals were observed for the writhing. Since the prodrug would not hydrolyze in the first 1-2 h as the prodrug probably be in the stomach or duodenum in this time interval hence all the results are statistically insignificant. For the ester the maximum activity was obtained after 6 h showing that absorption of drug does not occur in stomach. Ulcerogenicity studies showed that ester is not ulcerogenic (Table 2). The photographic presentation of ulcers is depicted in fig. 1. In vitro hydrolysis studies have shown that the ester is quite stable in simulated gastric and intestinal fluid where as it is hydrolyzed almost completely in rat faecal contents representing the colon. Although, this is the method of confirming the hydrolysis of ester in colon, it gives an idea about the

behavior of ester in gastrointestinal tract. This can be further confirmed by improved analysis, one approach to it is to isolate the specific strain of microorganism responsible for hydrolysis of CD and study the hydrolysis of conjugates in presence of them. The present study clearly indicates that conjugation of \(\beta \)-CD with mefenamic acid is a good method of masking the -COOH group and thus reducing the ulcerogenicity, a major drawback. The synthesized compound need to be further studied before being considered as potentially useful prodrugs.

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