

Analytical Methods for Standardization of Ayurvedic *Asavas* and *Aristas*; A Review

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Das et al.: Analytical Methods of Standardization of Asava and Arista

Plants serve as a rich source of bioactive molecules, which are used to treat various diseases in Ayurveda. Medicinal plant materials are formulated into valuable Ayurvedic medicines by application of modern scientific techniques, where standardization plays a pivotal role for authentication. Standardization confirms the identity, quality and purity of drugs. World Health Organization has set up suitable specific standardization parameters to evaluate the crude drugs and their finished products. These include various evaluation techniques such as pharmacognostical, physico-chemical, phytochemical, analytical, biological and biotechnological. Nowadays, application of several modern analytical techniques have become inevitable for evaluating the polyherbal Ayurvedic formulations to ensure quality, safety and efficacy. Various spectroscopic and chromatographic methods applicable for this purpose are ultra-violet spectroscopy, Fourier-transform infrared spectroscopy, nuclear magnetic resonance spectroscopy, thin-layer chromatography, high performance thin layer chromatography, high performance liquid chromatography, gas chromatography, mass spectroscopy and hyphenated techniques such as gas chromatography-mass spectroscopy, liquid chromatography-mass spectroscopy, liquid chromatography-nuclear magnetic resonance spectroscopy. Out of several formulations available in Ayurveda, *asavas* and *aristas* are considered as unique dosage forms due to their indefinite shelf life. Standardization of different *asava-arista* formulations using various analytical techniques are vividly discussed in this review.

Key words: Ayurveda, *asava*, *arista*, standardization, analytical methods

Medicinal plants are known to contain various bioactive phytoconstituents, which are used for prevention and mitigation of various ailments all over the world. About 80 % of the world population depends upon herbal preparation for their primary health care^[1]. India is considered as rich source of a good number of medicinal plants and thus called as medicinal garden of the world. Medicinal plants are best adopted by rural and urban community in India due to their non-toxic nature, less side effects and low price. Nowadays, there has been an increase in demand for these plant-based products in developed countries as well^[2]. Plant, mineral and animal-based natural drugs are the main sources, which contribute the bioactive components for preparation of various Ayurvedic formulations. The ancient books such as *Rigveda*, *Atharvaveda*, *Charaka Samhita*, *Sushruta Samhita*, *Astanga Hridaya* and *Sangraha*, describe various formulations and their uses against different diseases. Serious adverse toxic effects of synthetic drugs have shifted attention of modern

civilization more towards the Ayurvedic formulations for safer remedies. However, there is a lack of quality in herbal preparations due to geographical variation, confusion with different regional names, adulteration and substitution and absence of proper standardization procedures for evaluation of raw materials and finished products. Therefore, at present standardization of crude drugs as well as their formulations by implementation of quality control parameters has become highly essential^[3]. It is observed that development of suitable standard procedures for authentication of complex herbal formulations is not an easy task. The traditional methods of drug evaluation are not sufficient to establish the quality aspects of complex polyherbal

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Ayurvedic preparations. However, World Health Organization has framed certain standardization guidelines for evaluation of the crude drugs and their finished products, which include determination of their macro and microscopical characters, physico-chemical characters, presence of heavy metals, microbial limit, analytical parameters for qualitative and quantitative study of biomarkers, toxicity and biological study, DNA finger printing^[4]. *Asavaristas* are such Ayurvedic self-generated alcoholic formulations, which are prepared by fermentation of an infusion or juice or decoction of drug ingredients with the addition of sugar and *Dhataki pushpa* (*Woodfordia fruticosa*) as a fermenter^[5]. In this review, a discussion on the standardization of various marketed and in-house *asvarista* formulations by different analytical methods is presented.

STANDARDIZATION OF ASAVA AND ARISTA FORMULATION

Asavas and *aristas* are alcoholic preparations, prepared either by soaking the powdered drugs or the decoction of a drug, in a solution of jaggery along with a fermenter for a specified period of time, during which it undergoes fermentation to produce alcohol. These self-generated alcohols facilitate the extraction of active principles present in the drug and also serve as a preservative^[6].

Various methods applied for standardization of herbal drugs are depicted in fig. 1. Due to complexity of most Ayurvedic formulations, use of only conventional methods for standardization are not adequate for their evaluation. The Ayurvedic Pharmacopoeia of India and Pharmacopoeial standards for Ayurvedic formulations mention only the study of physico-chemical parameters and thin-layer chromatography of raw materials and formulations, which are not sufficient for proper standardization in present era^[7,8].

Therefore, modern analytical methods such as high performance thin layer chromatography (HPTLC), high performance liquid chromatography (HPLC), gas chromatography (GC) and hyphenated techniques such as liquid chromatography-mass spectroscopy (LC-MS), liquid chromatography-nuclear magnetic resonance spectroscopy (LC-NMR) and gas chromatography-mass spectroscopy (GC-MS) are applied to ascertain the quality of herbal products. Fingerprints obtained from HPTLC, HPLC are used as important tools for identification of marker compounds in the phytoconstituents and for quality control development of herbal formulations. Development and application of analytical techniques help in rapid

analysis of herbal formulation in industry and assist in maintaining the therapeutic efficacy and safety of Ayurvedic preparations^[9].

Thin layer chromatography (TLC) is a common fingerprinting technique used to identify the phytoconstituents present in the drugs and thus, helps in differentiation of various plant species^[10-12], simultaneously HPTLC is an important modern analytical method, where low or moderate polar compounds can be analysed. Pharmaceutical industries widely use this technique for method development, identification and detection of adulterants and substituents in the Ayurvedic formulations^[13]. Preparative and analytical HPLC methods are used for isolation, purification and quantification of phytoconstituents in the herbal formulation^[14]. Better resolution, sensitivity and rapid analysis are the important parameters considered in HPLC analysis^[15]. The combination of HPLC and MS is currently the most powerful technique for the quality control of Chinese herbal medicine^[16]. GC is used in characterization of volatile compounds due to its powerful separation efficiency and sensitive detection^[17]. Compounds present in essential oils are identified and quantified by GC-MS analysis^[18]. LC-MS is another important analytical technique for determination of quality of the drug^[19]. LC-NMR is used in pharmacokinetics, toxicity studies, drug metabolism and drug discovery process due to its rapidity and sensitivity of detection^[20,21]. LC-NMR technique is also used to detect adulterants in Chinese herbal medicine^[22]. Standardization procedures of various *asavarista* formulations are discussed as follows:

Abhayarista:

A comparative study of *Abhayarista* formulations showed that the main polyphenolic compounds of *Terminalia chebula* (chebulic acid and chebulinic acid) were hydrolysed to their respective monomers and the amount of chebulic acid, gallic acid and ethyl gallate were increased after fermentation when compared to the decoction, which was estimated by HPLC (Jasco PU 1580, detector UV/Vis, Jasco UV 1575) method^[23]. Ethanol content of four marketed *Abhayarista* preparations were determined by GC method (Chemito GC 7610, Carbowax 20 M)^[24]. In another study the ethanol content was determined using redox titration and GC method (Chemito GC 7610, Carbowax 20 M). Gallic acid, the major active constituent of the *Terminalia chebula*, was quantified

Methods of evaluation of asava & arista formulation

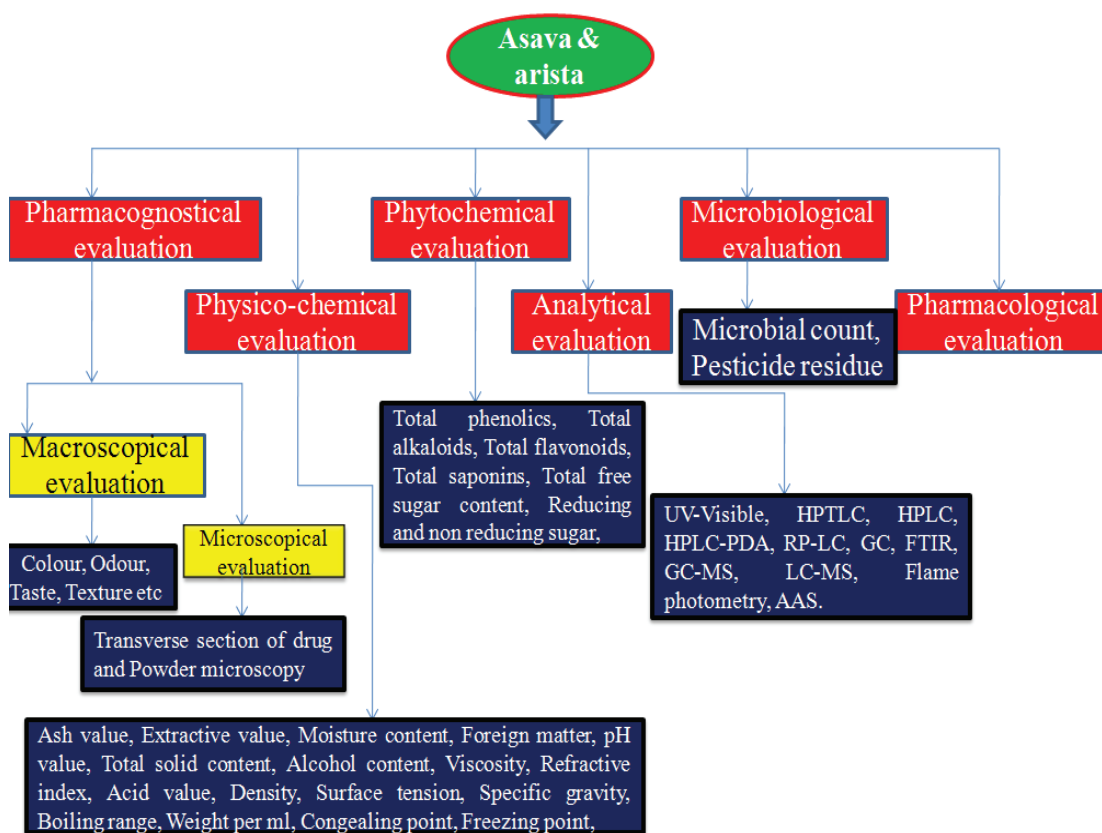


Fig. 1: Methods of evaluation of *asava* and *arista*

by UV (Shimadzu 1800, cyclomixer, Remi) and HPLC (Waters platform ZMD 4000 system, Micromass ZMD mass spectrometer, Waters 2690 HPLC equipped with a Waters 996 diode array detector, Mass Lynx software version 3.1, Waters Corp., Milford, MA, USA) methods^[25].

Amritarista:

Amritarista was prepared using pure and authenticated ingredients according to the Ayurvedic Formulary of India and was analysed by TLC method. It showed a single spot of yellowish-brown colour under UV light with the solvent system, n-butanol:glacial acetic acid:water (4:4:2), while chromatogram was sprayed with alcoholic KOH solution^[26].

In another study, HPTLC (Camag, TOSOH-CCPM system, Switzerland) method was developed in order to standardize the marketed *Amritarishta* formulation using luteolin and apigenin as marker compounds. Two fractions (I and II) of ethanol extract of the above formulation and standard luteolin and apigenin were applied on HPTLC plate using toluene:ethyl acetate:glacial acetic acid (5:4:1) as the mobile phase. It was observed that R_f values of both luteolin (0.64)

and apigenin (0.81) were found comparable in the sample and the reference standard^[27].

Aravindasava:

Aravindasava was prepared by traditional method and analysed by TLC. *Aravindasava* showed three spots having yellowish-grey, grey and violet colour and two brown spots under UV light at 365 nm using the solvent system, n-butanol:glacial acetic acid:water (4:4:2), while sprayed with alcoholic KOH solution^[26].

Arjunarista:

Phenolic compounds such as ellagic acid, gallic acid, ethyl gallate, quercetin and kaempferol were identified using a reversed-phase HPLC (Shimadzu HPLC, Japan, LC-10AT pump, Shimadzu SPD-M10 A, version 6.10, Rheodyne 7725 I manual injector, CA, USA) method and presence of these marker compounds were compared between the drug decoction and finished formulation. Other constituents present in the formulation did not interfere with these marker compounds as these were present at very low concentration. The comparative study of two chromatograms (decoction and formulation) showed

that the amounts of gallic acid and ellagic acid were increased during fermentation, which could be due to the hydrolysis of ellagitannins and gallitannins^[28]. TLC of *Terminalia arjuna* showed the presence of ellagic acid^[29]. In another study, the amount of gallic acid was estimated by colorimetric analysis^[30]. The amount of gallic acid and ellagic acid in *Arjunarista*-T (prepared by traditional method), *Arjunarista*-M (prepared by modern method) and its marketed formulation were determined using HPTLC (Camag, Desaga, Ziegel Wiesen, Germany, AS 30 Win sample applicator, Switzerland) method^[31].

Ashokarista:

The bark extract of *Ashoka* was standardised using TLC using catechin as the standard marker^[32]. TLC studies of the marketed brands (Baidhyanath, Dabur, Zandu) and in-house preparation showed the presence of kaempferol, which was closer to the standard kaempferol^[33]. Different types of phenolic compounds such as gallic acid, protocatechuic acid and rutin present in *Ashokarista* were identified by a liquid chromatograph coupled with photodiode array detector^[34]. Total phenolics, total alkaloids, total flavonoids and total saponins in marketed and in-house *Ashokarista* preparation were determined on a UV spectrophotometer (Shimadzu 1800, Cyclomixer, Remi). Total phenolic, flavonoid and alkaloid compounds were found to be more in the marketed formulation than in the in-house preparation. Whereas total saponin compounds were found to be very less in the marketed preparation as compared to the in-house preparation. These differences in the amount of chemical constituents could be due to variations in geographical regions of raw materials and different methods of processing^[35]. TLC of *Ashokarista* showed three spots with yellowish-grey, grey and violet colour in the solvent system n-butanol:glacial acetic acid:water (4:4:2) and one brown colour fluorescent spot under UV light at 365 nm, which became red after spraying with alcoholic KOH^[26].

Ashwagandharista:

Ashwagandharista was standardised using a HPLC (Jasco PU 1580, detector UV/Vis, Jasco UV 1575) method with standard withaferin-A and withanolide-A^[36]. TLC of *Ashwagandha* root powder showed one blackish-brown spot in the solvent system benzene:ethyl acetate (9:1)^[37]. *Ashwagandharista* was standardised by FTIR (Jasco FTIR 410) and HPTLC

(Camag HPTLC, Linomat V applicator, TLC scanner 3, Reprostar 3 with 12 bit CCD camera, WinCATS-4 software, Switzerland) methods for the estimation of biomarker withanolide-A. Heavy metals such as lead (Pb), mercury (Hg), cadmium (Cd), arsenic (As) were detected by atomic absorption spectroscopy. FTIR studies of the formulation revealed the presence of functional groups similar to *Ashwagandha* powder and HPTLC study showed the presence of biomarker withanolide-A^[38]. *Ashwagandharista* manufactured by three different companies was evaluated using chromatographic methods. The presence of withanolide-D was detected by HPTLC (Camag HPTLC, TLC scanner 3, WinCATS-4 software, Switzerland) in ASA-DAB (*Ashwagandharista* manufacture by Dabur) and ASA-BDN (*Ashwagandharista* manufacture by Baidyanath) but it was not clearly visible in ASA-AVP (*Ashwagandharista* manufacture by Arya Vaidya Pharmacy) due to overlapping of the bands^[39]. TLC of *Ashwagandharista* showed only three spots under UV light with the solvent system n-butanol:glacial acetic acid:water (4:4:2)^[26].

Ayaskrti arista:

Ayaskrti arista formulation of three different batches, manufactured by Arya Vaidya Sala, were collected for the analytical study. Gallic acid was quantitatively estimated in ethyl acetate extract of the formulation by a HPTLC (Camag HPTLC, Switzerland) method using the solvent system toluene:ethyl acetate:acetic acid:water (3:3:0.8:0.2)^[40].

Balarista:

Total phenolics, alkaloids, flavonoids and saponins content in marketed and in-house preparations of *Balarista* were determined on a UV spectrophotometer (Shimadzu 1800 and Cyclomixer, Remi). Total phenolic content was found to be less while the total flavonoid content was more in the in-house preparations, whereas the total alkaloid content in the in-house preparation was very less as compared to marketed formulation. Total saponin content was found to be almost double in the in-house preparation compared to that of the marketed formulations. The difference in the values of phytoconstituents might have developed due to variation in geographical localization of raw materials and different methods of processing^[35]. In a separate study, toxic alcohol residue like methanol was found to be absent, which was confirmed by GC-MS (PerkinElmer Clarus 500 with Mass selective detector)

analysis. The presence of aflatoxins and heavy metals were also found to be absent in the tested formulation^[41]. TLC showed three spots under UV light at 365 nm with solvent system n-butanol:glacial acetic acid:water (4:4:2)^[26].

Brahmiarista:

Marketed formulation of *Brahmiarista* was analysed by HPTLC (Camag Linomat V, TLC scanner 3, WinCATS software, version 1.3.0 Camag, Switzerland) using bacoside-A as the marker compound. The formulation did not show any peak corresponding to bacoside-A when it was run in toluene:ethyl acetate:methanol:glacial acetic acid and sprayed with 10 % sulphuric acid in alcohol, which confirmed the absence of *Bacopa monnieri* as the main ingredient, rather it could contain some substitutes^[42].

Chandanasava:

The effect of time on fermentation and storage of *Chandanasava* was studied and evaluated using TLC. *Chandanasava* was prepared in an earthen pot according to the Ayurvedic Pharmacopoeial method and at 15 d interval, the pot was opened and the content was analysed. The formulation, which was stored in glass bottle after fermentation was subjected to TLC study. TLC showed no difference between the formulation obtained after 30 d of fermentation in an earthen pot and the product stored in glass bottle for three months^[43]. HPTLC (Camag HPTLC, TLC scanner 3, WinCATS 4 software, Switzerland) chromatogram of *Chandanasava* showed a spot at the same location for all batches to confirm the batch-batch consistency^[44]. Marketed *Chandanasava* formulation was subjected to GC-MS analysis (PerkinElmer Clarus 500 with mass selective detector), which showed the absence of toxic ingredients such as methanol, aflatoxins and heavy metals^[41]. TLC showed three spots under UV light at 365 nm with solvent system n-butanol:glacial acetic acid:water (4:4:2), while sprayed with alcoholic KOH^[26].

Drakshasava:

Drakshasava was prepared and subjected to quantitative determination of phytoconstituents using UV spectroscopy (Shimadzu UV-1800 UV/Vis scanning spectrophotometer). Quantitative determination of total phenolics and tannins showed that these contents were 6.34 and 1.18 µ/ml, respectively. Formulation did not show presence of alkaloids^[45].

Dasamularista:

In a comparative study, *Dasamularista* was prepared using identical size, shape and capacity of earthen pots, stainless steel vessel and porcelain jar by filling up to 1/2, 2/3rd and 3/4th capacity of the container. TLC using solvent system butanol, acetic acid and water (63:17:10) showed five spots in different preparations. From different analytical and physico-chemical study results, it was found that pot filled up to 3/4th capacity was most suitable for fermenting *Dasamularishta*^[46]. Presence of total phenolics, alkaloids, flavonoids and saponins in the marketed *Dasamularishta* and in the in-house preparation was analysed on a UV spectrophotometer (Shimadzu 1800 and Cyclomixer, Remi). Percent total phenolic content was found to be more in the in-house preparation than the marketed formulation, whereas the total flavonoid, alkaloid and saponin content was less in the in-house preparation than the marketed formulation. Variations in geographical sources of raw materials and different methods adopted for their processing might have influenced these differences^[35]. TLC showed two spots under UV light with the solvent system n-butanol:glacial acetic acid:water (4:4:2) when sprayed with alcoholic KOH^[26].

Draksharista:

HPTLC (Desaga, Ziegel Wiesen, AS 30 Win, Desaga TLC scanner CD 60, ProQuant software 1.06, Germany) method was developed for comparative quantification of quercetin and rutin in *Draksharista* formulations prepared using traditional, non-traditional method and also in marketed formulation. Mobile phase such as toluene:ethyl acetate:methanol:formic acid (6:3:0.2:0.4) and ethyl acetate:n-butanol:formic acid:water (10:6:2:2) were used for estimation of quercetin and rutin, respectively. Comparatively, formulation prepared by traditional method found to contain more quercetin and rutin than the formulations prepared by other methods^[47].

Pillai *et al.* performed HPTLC study (Camag HPTLC, Linomat V, WinCATS software version 1.4.6) profile of marketed *Draksharista* formulation and the raw materials used in the preparation. The alcohol extract of two marketed formulations and all raw materials were subjected to HPTLC analysis using gallic acid, catechin and resveratrol as marker compounds with solvent system toluene:ethyl acetate:formic acid (6:4:0.8), toluene:ethyl acetate:formic acid (5:6:1) and chloroform:ethyl acetate:formic acid

(5:4:1), respectively. Results revealed that marketed *Draksharista* formulation contained all the components as mentioned in Ayurvedic Formulary of India and also contained the marker compounds^[48].

Validated HPTLC (Camag HPTLC, Linomat V, WinCATS software version 1.4.6) method was employed for the estimation of gallic acid, catechin and resveratrol in three batches of in-house *Draksharista* formulation and two marketed formulation (M1 and M2) using the same extract and solvent system as reported previously by Pillai *et al.* Gallic acid was found to be more in the in-house formulation (batch-3) as compared to marketed formulation. Catechin was found to be more in the in-house formulation (batch-1), while it is very less in the marketed formulation (M2). Resveratrol was also found to be more in the in-house formulation (batch-1)^[49].

Gallic acid and catechin were also quantitatively estimated by HPTLC (Desaga, Ziegel Wiesen, AS 30 Win, Desaga TLC scanner CD 60, ProQuant software version 1.06, Desaga, Germany) in *Draksharista* formulation prepared by traditional and modern methods; and also in the marketed formulation. It was observed that the amount of gallic acid and catechin were found to be more in the formulation prepared by traditional method^[50].

Jirakadyarista:

Two major compounds, apigenin-7-O-[galacturonide (1->4)-O-glucoside] and luteolin-4'-O-[glucoside-7-O-galacturonide] of *Jirakadyarista* were detected by RP-HPLC (Shimadzu Co., Japan, LC-20AT pump, UV detector, Shimadzu SPD-20 A, Rheodyne 7725 I, CA, USA). During fermentation process 7-O-glucosides of luteolin and apigenin underwent hydrolysis to increase the amount of luteolin and apigenin in the preparation. It was also observed that monomeric phenolic compounds and 5-hydroxymethyl furfural were introduced into the formulation through the jaggery and other plant materials^[51].

Kanakasava:

Three brands of *Kanakasava* were procured from market and evaluated for ethanol content using specific gravity and GC methods (Chemito GC7610, Carbowax 20M). Results showed that ethanol content measured by both methods were comparable to each other and the values were found to be within the limit^[52]. Gallic acid and ethyl gallate were quantitatively estimated in the ethanol fraction of *Kanakasava* by HPTLC (Camag

HPTLC, Linomat-V, TLC scanner 3, WinCATS-IV) using toluene:ethyl acetate:formic acid:methanol (3.5:3.5:0.8:0.5) as mobile phase^[53].

Kharjoorasava:

Kharjoorasava was prepared by a traditional method using *Dhataki pushpa* (*Woodfordia fruticosa*) and *Hapusha* (*Juniperus communis*) as fermenter, which was evaluated using TLC. Results revealed close resemblance between the two formulations due to presence of similar secondary metabolites. During fermentation process the bioactive compounds were transformed in the formulation medium due to very slow progress in chemical reaction^[54].

Kumaryasava:

Alcohol content of *Kumaryasava* was determined using both specific gravity as well as GC (Chemito GC7610 Carbowax 20M) method. The result showed gradual reduction in ethanol content on storage of *Kumaryasava* in different containers, which may be due to vaporization on opening of the container. Therefore, *asava* and *arista* preparations have to be consumed within a shorter period of time or the formulation can be prepared in smaller volume. Total phenolic content was found to be 0.1 %^[55]. According to Dash *et al.*^[56] the limit of total phenolic compound should not be less than 0.06 % w/v.

In another study, *Kumaryasava* was subjected to UV (Shimadzu Pharma spec UV-1700) and FTIR (Jasco FTIR 410) spectroscopic analysis. Aloin was isolated from different fraction of *Kumariasava* and used as marker for standardization by HPTLC (Camag, Linomat IV, Camag scanner 3, Switzerland) and HPLC (TOSOH-CCPM) instrumental methods. *Kumaryasava* was fractionated with petroleum ether, benzene, chloroform and ethyl acetate. Chloroform fraction showed five spots, which confirmed presence of five components, out of which three components were separated by preparative TLC. Fraction III, IV and V were found to be flavonol, isoflavones and anthraquinone, respectively. UV and IR spectra of fraction V produced the characteristic peaks indicating the presence of quinones. UV, IR and HPTLC fingerprints of fractions III-V could be used for routine standardization of *Kumaryasava*. Fraction V was subjected to HPLC analysis, which gave two peaks. Aloin, an anthraquinone glycoside, whose presence was confirmed by modified Borntrager test, TLC and HPLC analysis in fraction V. Both fraction

V and standard marker aloin were also well compared by TLC, HPTLC and HPLC analysis, confirming the presence of aloin in *Kumaryasava* preparation. This could be a simple, accurate and routine method for analysis of *Kumaryasava*^[57].

Kutajarista:

Kutajarista was standardized by HPTLC (Camag, Linomat IV, model 3 scanner, Cats integration software, version 4.03) employing a solvent system of ethyl acetate, n-hexane and triethyl amine (70:24:6), HPLC (Water HPLC system equipped with model 510-HPLC pump, 410 RI detector, Milford, MA, USA) with methanol and water (95:5) and HPLC-MS (Waters HPLC-MS system equipped with model 2525 pump, ZQ detector) with acetonitrile and water (95:5) using conessine as the biomarker. HPTLC chromatogram of alkaloidal fraction of *Kutajarista* showed three well resolved spots. The spot having R_f value 0.40 matched with standard conessine after spraying with Dragendorff's reagent. The presence of conessine was confirmed in alkaloidal fraction by HPLC-MS as well. The peak of standard conessine appeared at the retention time of 12.5 min also appeared in extract and exhibit similar mass fragmentation. HPLC analysis of alkaloidal fraction of formulation gave a single major peak with retention time of 4.17 min, which matched with HPLC analysis of standard conessine^[58]. Microbial presence of *Kutajarista* at initial stage of fermentation was studied by culture independent 16SrRNA gene clone library approach. Gallic acid was recovered on d 0, ellagic acid and gallic acid on d 8 and gallolyl derivatives and ellagic acids on d 30 of fermentation, which were determined by HPLC-MS analysis (Waters HPLC-MS system equipped with model 2525 pump and ZQ detector)^[59]. TLC showed two spots under UV with solvent system n-butanol:glacial acetic acid:water (4:4:2) when sprayed with alcoholic KOH^[26].

The role of different types of containers and methods of preparation in *Kutajarista* was studied. *Kutajarista* was prepared by two methods, traditionally using *Dhatakopuspa* as fermenter and non-traditionally using yeast as fermenter, in different containers made up of mud, wood, stainless steel and plastic. Fermentation was started on d 2 in formulation containing yeast as fermenter and on d 5 in formulation containing *Dhatakopuspa* as fermenter. Qualitative phytochemical test of formulation showed the presence of steroids, triterpenoids, proteins, tannins and alkaloids.

Quantitative determination by UV (Shimadzu UV-1800 UV/Vis scanning spectrophotometer) showed the presence of alkaloids and tannins in highest percentage in stainless steel and plastic container for both samples prepared by traditional and non-traditional methods^[60].

Lohasava:

Lohasava was prepared according to the Ayurvedic Formulary of India and analysed by TLC. TLC study showed two spots; violet and grey under UV light using solvent system n-butanol:glacial acetic acid:water (4:4:2) when sprayed with alcoholic KOH^[26]. Ethanol content of *Lohasava* was determined by GC (PerkinElmer GC, Clarus 500). Analysis of heavy metal was carried out on a PerkinElmer Optical Emission Spectrometer, Optima 2100DV. Pb and Hg were found to be absent, whereas As and Cd were present within specified limit of World Health Organisation (WHO). Gallic acid was quantitatively estimated in ethyl acetate extract of *Lohasava* using toluene:ethyl acetate:formic acid (10:7:1) as mobile phase by HPTLC (Camag Linomat IV, Camag scanner 3, software WinCATS 1.4.2)^[61].

Mustakarista:

Four different brands of marketed *Mustakarista* formulations were evaluated by GC (Chemito GC7610 Carbowax 20M) method for the quantification of alcohol content. Specific gravity and results showed gradual reduction of self-generated ethanol content on storage, which may be due to evaporation on repeated opening of the container. GC method provided accurate and precise result as compared to the specific gravity method. Variation in alcohol content was observed with different containers used in the manufacturing process. Total phenolic content was found to be 0.09 and 0.08 % for two marketed formulations of *Mustakarista*^[55].

Saraswataristam:

FTIR spectrum of *Saraswataristam* formulation showed number of peaks at 400-4000 cm^{-1} . The broad peak was observed at 3446 and 3442 cm^{-1} due to presence of OH and functional group of other ingredients in the formulation. Similar peak was observed in *Centella* powder, which was the main ingredient of the formulation. HPTLC (Camag HPTLC, Linomat V applicator, TLC scanner 3, Reprostar 3 with 12 bit CCD camera, WinCATS software, Switzerland) analysis of formulation showed a peak of asiaticoside^[62].

Vidangarista:

HPTLC (Camag, Linomat V, TLC scanner 3, WinCATS software 1.4.4.6337, Switzerland) method was developed for the quantification of biomarkers gallic acid and conessine in the *Vidangarista* formulation^[63]. It is a polyherbal formulation mentioned in Ayurvedic Formulary of India and used as an anthelmintic.

In India Ayurvedic drug industry is growing at a rapid pace and more herbal products are released into the market. The safety and efficacy of these formulations need to be ensured through application of proper standardization protocols. Traditional methods of standardization are found to be insufficient to validate these formulations, hence, modern advanced techniques play vital role. Indian Ayurvedic formulations could be well accepted by all developed nations world-wide if these are manufactured using standard procedures and standardized using sophisticated modern analytical techniques. Fingerprint profile obtained by various chromatographic techniques play an important role for the standardization of Ayurvedic formulations. It is essential to develop advanced hyphenated techniques to serve as rapid and specific tools for herbal drug standardization. The combination of qualitative fingerprint and quantitative multicomponent analysis act as a novel and rational method in the quality control of Ayurvedic formulations. From this review it could be concluded that great scope existed to develop analytical methods to authenticate majority of *asava-arista* formulations. The standardization protocols using hyphenated techniques such as GC-MS, LC-MS, LC-NMR, could be developed and employed for evaluation of multidrug *asava-arista* formulations where miniscule amount of marker compounds are available. Modern analytical methods of standardization are yet to be developed for most of the other Ayurvedic formulations.

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