Effect of Protective Coating of Aspirin Tablets with Acrylatemethacrylate Copolymers on Tablet Disintegration Times and Dissolution Rates

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Tablets of aspirin (a moisture degradable drug) have been film coated with two analogous Eudragit RL and RS copolymers designated here as A and B which differ only in their cation content in the ratio 2:1 (A:B). A, is therefore more hydrophilic than B. The tablets were film coated with ethanol solutions of these two polymers. Film coating with either A or B significantly reduced the moisture uptake potentials of the tablets but caused an increase in the disintegration times of the tablets and retarded dissolution rates. The mean disintegration times were 0.5±0.1 min (uncoated tablets), 16±2.5 min (tablets coated with A) and 115±3.6 min (tablets coated with B). The corresponding dissolution rates % h⁻¹ were 28.3 for uncoated, 16.6, coated with A and 14.8, coated with B, respectively. Thus, coating with polymer B considerably impaired the disintegration and dissolution properties of the tablets.

Key words: Eudragit copolymer, film coating of tablets, tablet disintegration time and dissolution rates

An important area of application of polymeric film coating of tablets is to protect against moisture degradation. Aspirin for instance is moisture degradable and therefore its tablets require protective coating. Polymer films for this area of application should have a high moisture resistance and should dissolve or swell and disrupt when in contact with aqueous fluids to allow disintegration and dissolution of the tablet, otherwise bioavailability will be compromised. A previous study has indicated that film coating of tablets will invariably lead to increase in disintegration time.

The acrylatemethacrylates are water insoluble but swellable polymers. The presence of cationic (quaternary ammonium) groups in the polymer chemical structure confers the hydrophilic swelling property. Thus, the higher the cationic content the higher the porosity and permeability of resulting films. The polymers have been investigated as binders in tableting, microencapsulation of drug particles for controlled release application.

The present study investigates their applicability in protective coating of aspirin tablets against moisture degradation. Pursuant to this objective the water uptake potentials of the tablets, their disintegration times and dissolution rates were determined.

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Two analogous acrylate-methacrylate copolymers designated here as A and B were received under the trade names Eudragit RL and RS, respectively from Rhom Pharma, Darmstadt, Germany. A and B differ only in their cation content in the ratio 2:1 (A:B) and have an average molecular weight of 1,500,000. Glycerol triacetate (Sigma-Aldrich, Italy) and ethanol (analar grade, BDH) were used as plasticiser and casting solvent respectively in the formulation of the coating fluid. Aspirin tablets BP were used as substrate (core) in the coating procedure.

A sample of the polymer A or B (5 g) was dissolved in ethanol (100 ml) by overnight stirring. In another aspect of the study, the polymers A and B were mixed in different proportions to form the coating fluid with intermediate cation content (Table 1). Glycerol triacetate was added as plasticiser in a concentration of 0.5% w/v to the coating fluid. Aspirin tablets (n=30) were placed in a laboratory-coating pan (Manesty, UK. Model 75096) whilst the tablets were rotating in the pan, the coating fluid was applied in three aliquots of 5 ml each allowing 5 min drying at 60°C between each application and drying finally for 1 h also at 60°C.

The thickness of the tablets (n=10) were measured with a digital micrometer before and after coating. The difference divided by two gave the μm thickness. The tablet thickness before coating was 3240 μm and after coating was 3300 μm (tablets coated with polymer A) and 3296 μm (tablets coated with polymer B). The calculated mean film thickness was 30±8 μm (A) and 28±6 μm (B). The tablet hardness (kg) was determined using a Monsanto hardness tester (Monsanto Chemical USA). Twenty tablets randomly selected were used and mean values are reported.

Table disintegration test (DT) was determined in 0.1N HCL at 37±0.5°C by the British Pharmacopoeia method. Six tablets were used in each determination. Mean values are reported.

<table>
<thead>
<tr>
<th>TABLE 1: INTERMEDIATE POLYMER CATION CONTENT OBTAINED BY MIXING A AND B IN DIFFERENT PROPORTIONS</th>
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<tr>
<td>Proportions of the polymers (g)</td>
</tr>
<tr>
<td>A</td>
</tr>
<tr>
<td>10</td>
</tr>
<tr>
<td>7.5</td>
</tr>
<tr>
<td>5</td>
</tr>
<tr>
<td>2.5</td>
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A humidity chamber was created by placing a beaker of supersaturated solution of sodium chloride (500 ml) in a glass chamber giving a relative humidity (RH), 78% at the ambient temperature, 30°C. The weight of the tablets (previously equilibrated in a dessicator for 24 h) was individually determined using a sensitive electronic balance (Mettler Toledo B154, Switzerland) and then placed in a humidity chamber for various time intervals up to a maximum of 24 h. At pre determined time intervals the tablets were removed from the humidity chambers and re-weighed. The percentage (%) increase in weight was taken as the moisture uptake. Different humidity chambers were used for the different sampling intervals such that the tablets were not in contact with the external environment during the storage interval in the chambers.

The stirred beaker method described earlier was followed. In the procedure, aspirin tablet (300 mg) was placed in a basket of mesh size 450 μm which was in turn immersed in dissolution medium i.e. 800 ml water at 37±0.5°C, which was stirred 100 rpm with a Gallenkamp single blade stirrer. At specific time intervals a sample (5 ml) was withdrawn from the leaching fluid. The sample was heated to 100°C to allow for complete hydrolysis to salicylic acid. Freshly prepared ferric chloride solution (5% w/v, 1 ml) was then added. The sample was allowed to stand for 5 min during which a blue colour developed. Absorbances of the coloured samples were read at λ_{max} 540 nm in a UV spectrometer (Unicam, Sp500). The experiment was carried out in triplicate; and the mean absorbances were used to obtain the amount of drug dissolved at the different time intervals.

The data in Table 2 shows that tablet hardness increases slightly after film coating compared with the uncoated tablets. For instance, the mean hardness of the uncoated tablets was 4.7±0.5 kg while that for polymer A coated tablets was 5.6±0.5 kg, and for polymer B coated tablets, 5.6±0.7 kg. The difference in tablet hardness was not statistically significant (p >0.05).

The results of film coating on tablet disintegration time are presented in Table 2. The uncoated tablets disintegrated within half a minute. Coating with polymer A increased the disintegration time to 16 min against the BP limit of 30 min for coated tablets. Coating with polymer B however prolonged the
disintegration time considerably up to 115 min, completely outside BP specification. Thus protective coating with polymer B is not recommended while A is recommendable.

Mixing polymers A and B in different proportions gave a means of varying the cation content of the polymeric coating. The data in Table 2 indicated that as the cation content increased; DT decreased correspondingly. This finding is attributable to the potential of the cationic groups for promoting hydrophilic swelling of the films leading to their eventual rupture during disintegration test.

The amounts (%) of drug dissolving from the tablets at the various time intervals are plotted in fig. 1. The dissolution rates were obtained from the slopes of the linear portions of the plot. These were (% h⁻¹) 28.3 (uncoated tablets) 16.6 (tablets coated with polymer A), and 14.8 (tablets coated with polymer B). The dissolution rates of tablets coated with films of intermediate polymer cation content are presented in Table 2. These results indicate that polymer B markedly retard dissolution rates, attributable to its effect on the tablet disintegration times noted above. This retarded dissolution rate will have serious implication for bioavailability.

The results are presented in fig. 2 which shows that the uncoated tablets absorbed moisture more rapidly than the coated tablets, attributable to the barrier property of the films against moisture transmission. Polymer B, exhibited a slightly stronger barrier property compared to A. The difference can be associated with the higher content of the hydrophilic cationic groups in A. As the cation content increased the extent of moisture uptake will also increase (Table 2). This is attributable to the higher cationic content, hence higher porosity and permeability of resulting films.

<table>
<thead>
<tr>
<th>Parameters evaluated</th>
<th>Uncoated tablets</th>
<th>Cation content (moles, mol⁻¹ polymer chain) in the polymer coating:</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>33(B)</td>
</tr>
<tr>
<td>Hardness (kg)</td>
<td>4.7±0.5</td>
<td>5.6±0.5</td>
</tr>
<tr>
<td>DT (min)</td>
<td>0.5±0.1</td>
<td>115±3.6</td>
</tr>
<tr>
<td>Dissolution, % h⁻¹</td>
<td>28.3</td>
<td>14.8</td>
</tr>
<tr>
<td>Moisture content (%)</td>
<td>0.34</td>
<td>0.15</td>
</tr>
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</table>

![Fig. 1: Drug release profiles from aspirin tablets](image1)

Drug release profiles from uncoated (●) and coated aspirin tablets with Polymer A (●) and Polymer B (●).

![Fig. 2: Effect of relative humidity (78%) on moisture uptake](image2)

Effect of relative humidity (78%) on moisture uptake of uncoated aspirin tablets (●) and coated with Polymer A (●) and Polymer B (●).
In conclusion, film coating with the acrylate-methacrylate effectively protects the tablets against moisture uptake. However, polymer B impaired the disintegration and dissolution properties of the tablets. Hence, it is not applicable in the protective coating of the tablets. Whereas polymer A will be suitable for the protective coating of the aspirin tablets against moisture uptake. Polymer B is not considered suitable because of its effect on the disintegration and dissolution profiles of tablets.

REFERENCES


RP-HPLC and HPTLC Methods for the Estimation of Nebivolol hydrochloride in Tablet Dosage form

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Two simple, specific, accurate and precise methods, namely, reverse phase high performance liquid chromatography and high performance thin layer chromatography were developed for estimation of nebivolol hydrochloride in tablet dosage form. For the H PLC method, Lichrospher 100 C-18, 5 µm column consisting of 200×4.6 mm i.d. in isocratic mode, with mobile phase containing 50 mM KH2PO4 buffer (pH 3.0±0.1):acetonitrile (45:55 v/v) was used. The flow rate was 1.0 ml/min and effluent was monitored at 282 nm. The retention time was found to be 3.76±0.02 min. For the high performance thin layer chromatographic method a Camag system comprising of Linnomat V automatic sample applicator, Hamilton syringe, Camag TLC Scanner-3, Camag Win CAT software with stationary phase precoated silica gel 60F254 and mobile phase consisting of ethyl acetate:toluene:methanol:ammonium hydroxide (1:6:2:0.1 v/v/v/v) were used. The detection of spot was carried out at 282 nm. The R, value was found to be 0.33±0.02. The methods were validated in terms of linearity, accuracy and precision. The linearity curves were found to be linear over 10-150 µg/ml for high performance thin layer chromatography and 100-600 ng/spot for high performance thin layer chromatography. The limit of detection and limit of quantification for high performance thin layer chromatography were found to be 2.0 and 10 µg/ml, respectively, and for the high performance thin layer chromatography, 30 and 100 ng/spot, respectively. The proposed methods were successfully used for estimation of nebivolol hydrochloride in tablet formulation.

Key words: Nebivolol, H PLC, H PTLC, validation, analysis

Nebivolol is chemically, α,α'-[imino bis (methylene)]bis[6-fluoro-3,4-dihydro-2H-1-benzopyran-2-methanol]1, which is a selective β1-receptor antagonist without partial agonist activity2. Liquid chromatography-mass spectroscopic (LC-MS) methods3-5 for analysis of nebivolol in biological fluids, are reported in the literature. The present investigation describes two precise, accurate and specific, reverse phase high performance liquid chromatography (RP-HPLC) and high performance thin layer chromatography (HPTLC) methods for the estimation of nebivolol hydrochloride in tablet formulation.

All the reagent used were of HPLC and analytical