

High Efficient Extraction of Tryptophan Using Deep Eutectic Solvent-based Aqueous Biphasic Systems

YANHONG CHAO, H. DING¹, J. PANG², YAN JIN, X. LI³, H. CHANG³, W. JIANG³, GUAGYING CHEN⁴ AND W. ZHU^{3*}

School of Pharmacy, Jiangsu University, Zhenjiang 212013, ¹Affiliated People's Hospital of Jiangsu University, Zhenjiang 212002, ²Henan Engineering Research Center of Industrial Circulating Water Treatment, College of Chemistry and Chemical Engineering, Henan University, Kaifeng 475004, ³School of Chemistry and Chemical Engineering, Jiangsu University, Zhenjiang 212013, ⁴Key Laboratory of Tropical Medicinal Plant Chemistry of Education, Hainan Normal University, Haikou 571158, P. R. China

Chao *et al.*: Extraction of Tryptophan with Eutectic Solvent-based Aqueous Biphasic Systems

Deep eutectic solvent-based aqueous biphasic systems were established as greener replacements of organic solvents for the extraction of amino acid in this work. Choline chloride/polyethylene glycol 2000-based deep eutectic solvent was synthesized and characterized by element analysis, thermogravimetric analysis, and Fourier-transform infrared spectroscopy. The phase diagrams of deep eutectic solvent+salt ($\text{Na}_3\text{C}_6\text{H}_5\text{O}_7/\text{Na}_2\text{CO}_3/\text{NaH}_2\text{PO}_4/\text{K}_2\text{HPO}_4$)+ H_2O aqueous biphasic systems were determined at 298 K and the aqueous biphasic systems were applied to extract the tryptophan for the first time. The extraction conditions of amino acid concentration, deep eutectic solvent amount, salt dosage, and the extraction temperature were investigated and optimized. Under the optimum conditions, the extraction efficiency of tryptophan can reach up to 86.72-93.88 % using deep eutectic solvent-based aqueous biphasic systems with the salt of Na_2CO_3 , $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$, NaH_2PO_4 or K_2HPO_4 . UV/Vis and fluorescence spectra confirmed that the conformation of tryptophan did not change after extraction. Deep eutectic solvent-based aqueous biphasic systems have great potential application in highly efficient extraction of biologically relevant amino acids and other substances.

Key words: Aqueous biphasic systems, extraction, deep eutectic solvent, tryptophan

In current liquid-liquid separation systems, water-immiscible volatile organic solvents are widely used with their good extraction performance. But in the perspective of environmental issues of these hazardous solvents, there is a growing interest in finding greener replacements for liquid-liquid separation in industry and academic research. In recent years, aqueous biphasic systems (ABSs) aiming at avoiding the use of organic solvents as extractive phases have developed and been shown to be effective for separating organics^[1,2], metals^[3], carbon nanotubes^[4,5], and biological compounds^[6,7]. ABSs are composed of two aqueous-rich phases formed by polymer/polymer, polymer/salt, or salt/salt combinations. The basis of separation in ABSs is a direct result of the equilibration and selective distribution of target (bio) molecules between the two distinct aqueous phases.

Lately, ionic liquids (ILs, by definition salts which have melting points less than 100°) emerged as promising and alternative green replacements for

ordinary organic solvents^[8,9], due to their chemical and thermal stabilities, negligible volatility, general non-flammability, and great dissolving ability for a wide variety of compounds^[10,11]. Since 2003, Gutowski *et al.*^[12] found imidazolium-based ILs can also form ABSs with inorganic salt of K_3PO_4 . ABSs-based on ILs have received great attention in the field of liquid-liquid separations with their novel extractive ability and the feature of being environment friendly^[13,14]. However, the synthesis of ILs is complex and expensive, which limit the large-scale industrial applications and development of ILs. As a new class of ILs or ILs analogue, deep eutectic solvents (DES) have similar physical properties and phase behavior^[15] to traditional

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms

*Address for correspondence

E-mail: zhuws@ujs.edu.cn

ILs^[16,17]. DES can be synthesized easily using a certain molar ratio of two or three non-expensive components, which are capable of self-association through hydrogen bond interactions^[18,19]. Also, the excellent sustainable and biodegradable properties of DES highlight their advantages as promising non-toxic green solvents^[15,20]. DES have shown excellent performance in adsorption^[21,22], organics extraction^[23,24] and bioactive compounds separation^[25,26]. So, as a candidate ILs, DES are promising green solvent to form ABSs with potential great value in the field of extraction and separation.

Amino acids are building blocks of proteins and can be employed as elements or intermediates in antibodies, chelating agents and polypeptides^[27-30]. These wide applications make amino acids play a vital role in daily routines and manufacture^[31]. Separation and purification of amino acids are important and have crucial research value in human life. Traditional methods such as solid-liquid adsorption^[32-34], extraction with organic solvents^[35-37], ion-exchange^[38], centrifugation^[39,40] and chromatography^[39,41] are expensive and time consuming, and are not easily scalable^[42]. Moreover, the operational environment is not mild in many cases, which may result in amino acid inactivation. So finding new methods for easy and low-cost extraction of amino acids along with bioactivity retention has become an instant issue for biotechnology. The use of environment friendly and economical DES-based ABSs would be a good strategy.

In this paper, choline chloride (ChCl)/polyethylene glycol-based DES was synthesized and characterized. A series of ABSs composed of the synthesized DES and salt ($\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$, Na_2CO_3 , NaH_2PO_4 , or K_2HPO_4) solutions were established and applied for the extraction of tryptophan for the first time. Some of the significant factors effecting the extraction, such as the species and amount of salts, the concentration of tryptophan, the dosage of DES, and the temperature were optimized. The chemical nature of tryptophan before and after extraction was investigated using UV/Vis and fluorescence spectrometer.

MATERIALS AND METHODS

All reagents used were of analytical grade (AR) and purchased from Sinopharm Chemical Reagent Co., Ltd., Shanghai, China without further purification but polyethylene glycol 2000 (PEG, chemically pure) was dried under vacuum before use. Double-distilled water was used throughout the experiments. L-tryptophan

with a purity >99.0 % w/w was employed to investigate the extraction capacity.

Synthesis and characterization of DES:

DES was synthesized by stirring two eutectic mixture at 110° till a homogeneous, colorless liquid was formed. The investigated DES was based on ChCl and PEG with a certain mass ratio (PEG = 50, 60, 70, 80, 90, 100 %) according the process reported by Li *et al.*^[43]. The structure of the synthetic DES was confirmed using elemental analyses, thermogravimetric analysis (TGA) and Fourier-transform infrared spectroscopy (FT-IR).

Elemental analysis was carried out on an Element analyzer (Flash1112A) by weighting 2.3 mg samples in a tinfoil. TGA was done on STA-449C Jupiter (Netzsch Corporation, Germany). The sample was tested from room temperature to 800° with a heating rate of 10°/min under air atmosphere with airflow of 60 l/min. FT-IR spectra of samples were measured on a Nicolet Avatar-370 spectrometer at room temperature using the standard KBr disk method.

Phase diagram:

The phase diagrams were recorded by turbidimetric method under room temperature^[44]. DES solution (1.5 ml, 50 %, w/w) was added into a 10 ml centrifuge tube. Saturated solution of $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$ (or other salts) was added dropwise and shaken until the appearance of a cloudy solution. This was followed by dropwise addition of water to make the mixture clear again. The above process was repeated to obtain sufficient data to construct a liquid-liquid equilibrium phase diagram.

Extraction of tryptophan:

A certain amount of DES, salt ($\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$, Na_2CO_3 , NaH_2PO_4 , or K_2HPO_4) and tryptophan solution were added into graduated tubes, respectively. Afterwards, the biphasic solutions were left to equilibrate for 30 min (a time period established by shaking vigorously) to achieve a complete tryptophan partitioning between the two phases. The concentration of amino acid, in both phases, was determined on a UV/Vis spectrophotometer at a wavelength of 278 nm. A calibration curve obtained in the range of 0-0.1 g/l was $A = 25.52 C + 0.027$ ($R^2=0.9993$), where C (g/l) is the concentration of amino acid and A is the UV absorbance. Partitioning of tryptophan between the two phases were characterized by the partition coefficient (K), phase volume ratio (R), and extraction efficiency (E %), which can be calculated by

the following Eqns. 1, 2 and 3, $K = C_t/C_b$; $R = V_t/V_b$; $E \% = C_t V_t / (C_t V_t + C_b V_b) \times 100$, where C_t and C_b (g/l) are the concentrations of tryptophan in the DES-rich top phase and salt-rich bottom phase, respectively. V_t and V_b (ml) stand for the volume of the top and bottom phase, respectively.

RESULTS AND DISCUSSION

A series of DESs were synthesized with different mass ratios of ChCl and PEG, and the melting points were detected using a melder (fig. 1). DES had a minimum melting point at 60 % of PEG mass fraction (ChCl:PEG = 2:3, w/w), and this DES with the mass ratio of 2:3 (ChCl:PEG) was chosen for the later investigation^[45]. Elemental analysis of the prepared DES is listed in Table 1. The analysis value was approximated with the calculated value. It indicated that there was no mass defect in the process of the reaction.

The TGA, DSC and DTG curves for DES, PEG and ChCl are shown in fig. 2. The decomposition of PEG and ChCl were 259.7 and 316.7°, respectively. After DES is formed, the weight loss was divided into two steps, which were just assigned to its two reactants. According to the DSC curves of DES, the first step was an endothermic process attributed to the thermolysis of PEG, while the second step was an exothermic process attributed to the thermolysis of ChCl.

FT-IR spectra of the ChCl, PEG and DES are shown in fig. 3. The bands between 3000 and 2700 cm^{-1}

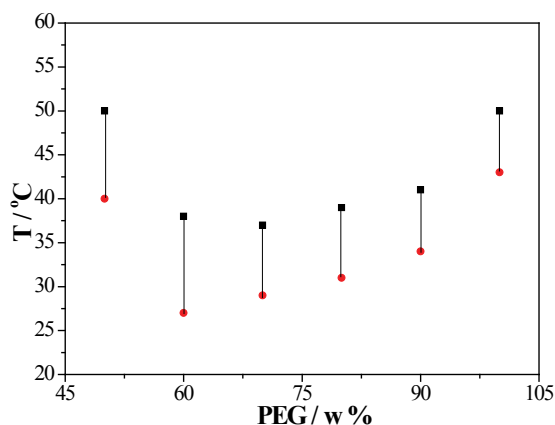


Fig. 1: Melting points of DES with different PEG mass fraction (●) Incipient melting point, (■) final melting point

TABLE 1: ELEMENTAL ANALYSIS OF C, H, N OF THE PREPARED DES

DES	C %	H %	N %
Calculated value	49.47	9.28	1.75
Analysis found	49.01	9.38	1.62

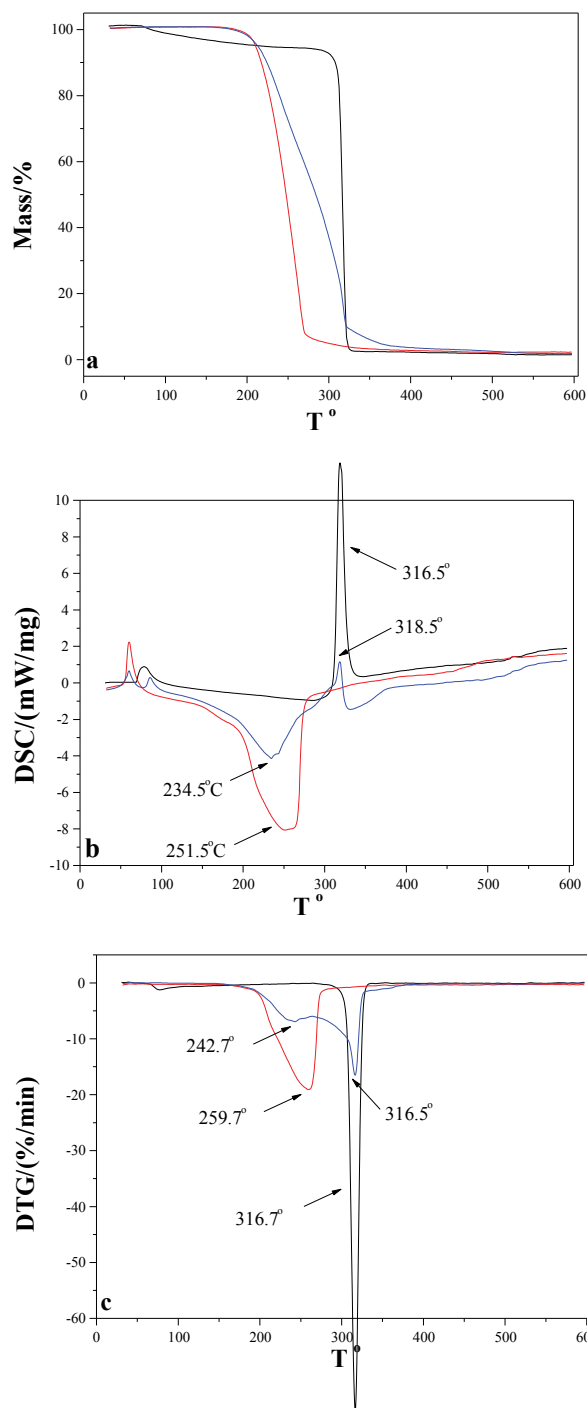


Fig. 2: (a) TGA, (b) DSC and (c) DTG curves for DES, PEG and ChCl

(—) DES; (—) PEG; (—) ChCl

corresponded to the stretching vibration of C-H in ChCl and DES. Compared with ChCl, the redshift of the peak of C-O (from 960 to 950 cm^{-1}) in DES was clearly seen, indicated that large amount of the stable hydrogen bonds were formed^[24]. The main characteristic peaks of ChCl and PEG could be observed in the FT-IR spectrum of DES, which identified that functional groups of reactant were stable when the reaction was processing.

Liquid-liquid equilibrium data were significant for the design of aqueous biphasic extraction process. In this study, the equilibrium compositions of the ABSs, DES (1)+salt ($\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$, Na_2CO_3 , NaH_2PO_4 , or K_2HPO_4) (2)+ H_2O (3), determined at 298 K are shown in fig. 4. These binodal curves can provide the information about the concentration fractions of DES and salt required to form an ABS. The region above the curve represented the biphasic system, while the system was a homogeneous phase below the curve. In addition, larger biphasic region indicated stronger ability for the salts to undergo ABS^[46]. It can be seen from fig. 4 that the phase-forming ability of salts followed the order: $\text{Na}_2\text{CO}_3 > \text{Na}_3\text{C}_6\text{H}_5\text{O}_7 > \text{K}_2\text{HPO}_4 > \text{NaH}_2\text{PO}_4$.

The extraction efficiency of tryptophan was investigated in a series of ABSs employing 0.28 g/ml of DES and 0.14 g/ml of different salts of $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$, Na_2CO_3 ,

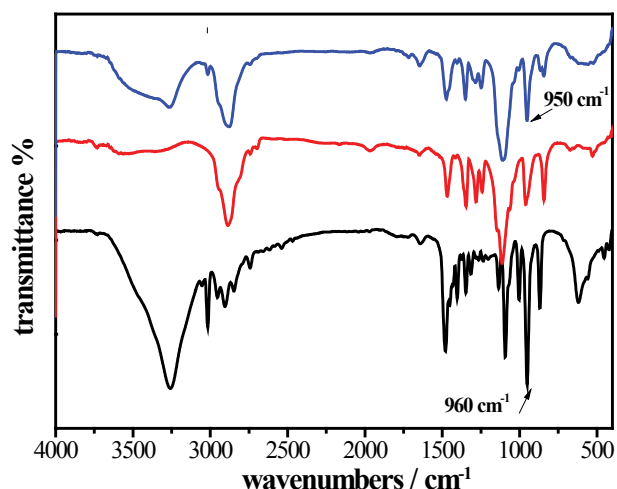


Fig. 3: FT-IR spectra of DES, PEG and ChCl
(—) DES; (—) PEG; (—) ChCl

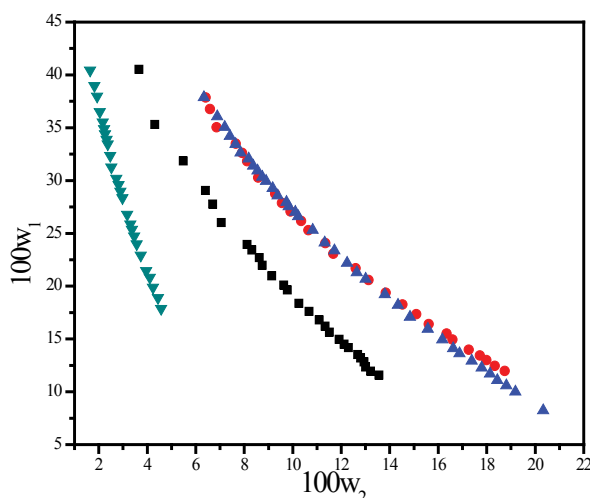


Fig. 4: Phase diagrams of DES-based ABSs with different salts
Phase diagrams of DES-based ABSs with different salts (●) K_2HPO_4 , (▲) NaH_2PO_4 , (■) $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$, and (▼) Na_2CO_3

NaH_2PO_4 , or K_2HPO_4 . As shown in fig. 5a, it was obvious that DES- Na_2CO_3 ABS showed the highest extraction efficiency of tryptophan, and the extraction efficiency in DES-based ABSs with different salt species followed the order of $\text{Na}_2\text{CO}_3 > \text{Na}_3\text{C}_6\text{H}_5\text{O}_7 > \text{K}_2\text{HPO}_4 > \text{NaH}_2\text{PO}_4$, which were in agreement with the phase-forming order mentioned previously.

Effect of tryptophan concentration on the extraction efficiency in series DES-based ABSs with different salts was investigated and shown in fig. 5b. Although there have been some swings along the way, the extraction efficiency of tryptophan decreased slowly with the increase of the concentration of tryptophan in overall trend for these DES-based ABSs. The reason was proposed that both the formation of ABS and the extraction of the target are depending on the competition for the water molecule^[47]. The massive addition of tryptophan will weaken the water affinity to the DES and cause the extraction of tryptophan decline in DES-rich top phase in the ABSs. The optimum concentration of tryptophan was 0.05 g/l.

The amount of DES was a significant factor that influences the extraction of tryptophan for ABSs and the result is shown in Table 2. With the increase of DES, the extraction efficiency of tryptophan in DES-based ABSs with Na_2CO_3 , NaH_2PO_4 , K_2HPO_4 and $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$ firstly continually increased, then reached the maximum values, and then decreased except DES- $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$ system. When the DES amount was more than 1.3 g (0.46 g/ml), there was not enough water to dissolve the DES, and the bi-phase system can't be formed. This result indicated that DES could promote the extraction of tryptophan at a suitable range, and the optimum amount of DES is 0.28, 0.46, 0.40 and 0.40 g/ml for the four ABSs of DES- Na_2CO_3 , DES- $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$, DES- NaH_2PO_4 and DES- K_2HPO_4 , respectively.

The effect of the addition of salts on the miscibility of a given system is very complex. As the salts dissolve in solvent, the ions are surrounded by a layer of water molecules. In the ABS, different compositions possess diverse ions, and different types of intermolecular interactions exist in the system^[48]. Then the extraction target is added in the disordered system increasing another rival to compete the quantificational water molecules. As shown in Table 3, with the increase of the dosage of salt, the extraction efficiency of tryptophan changed regularly and reached the maximum values of 85.86, 86.2, 88.41 and 91.55 % at their optimum dosage conditions for DES-based ABSs with Na_2CO_3 ,

$\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$, NaH_2PO_4 and K_2HPO_4 , respectively. What needs to be mentioned is that the aqueous biophase can't be formed when the salt amount is outside the extreme scope.

The extraction efficiencies of tryptophan in the four ABSs at temperatures ranging from 288-328 K were measured, and the results are shown in fig. 5c. As the temperature increased from 288 to 328 K, the extraction

efficiencies decreased to various extents. The DES- Na_2CO_3 system was more sensitive than the other three ABSs, the decrement value could reach approximately 26 % while the $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$ and NaH_2PO_4 systems reach about 15 %, the K_2HPO_4 system reach about 8 %. The possible reason might be that the water solubility of Na_2CO_3 is low and sensitive to temperature, which cause more tryptophan distributed into the salt-

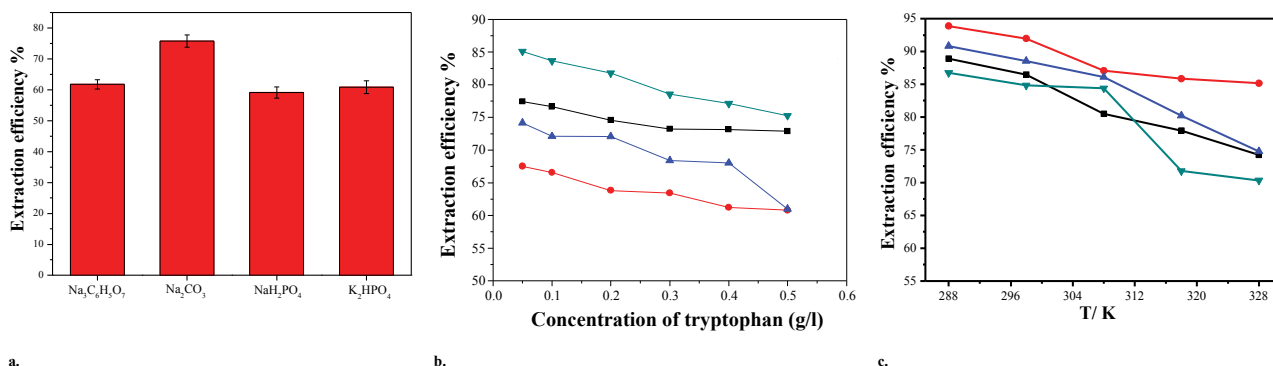


Fig. 5: Effect of salt species, tryptophan concentration and temperature on extraction efficiency

Effect of (a) salt species, $C_{\text{DES}}: 0.28 \text{ g/ml}$, $C_{\text{salt}}: 0.14 \text{ g/ml}$, $C_{\text{tryptophan}}: 1.6 \text{ g/l}$, 298 K, (b) tryptophan concentration, ($\text{---}\nabla\text{---}$) Na_2CO_3 , ($\text{---}\blacksquare\text{---}$) $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$, ($\text{---}\blacktriangle\text{---}$) NaH_2PO_4 , ($\text{---}\bullet\text{---}$) K_2HPO_4 and (c) temperature, ($\text{---}\bullet\text{---}$) K_2HPO_4 (ABS-1), ($\text{---}\blacktriangle\text{---}$) NaH_2PO_4 (ABS-2), ($\text{---}\blacksquare\text{---}$) $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$ (ABS-3), ($\text{---}\nabla\text{---}$) Na_2CO_3 on the extraction efficiency of tryptophan

TABLE 2: EFFECT OF DES AMOUNT ON THE EXTRACTION EFFICIENCY OF TRYPTOPHAN

DES (g/ml)	Salt (g/ml)	K	R	E %	DES (g/ml)	Salt (g/ml)	K	R	E %
Na_2CO_3					$\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$				
0.16	0.14	5.40	0.46	71.16	0.28	0.14	2.66	1.26	77.02
0.20	0.14	4.56	0.56	71.72	0.32	0.14	2.68	1.30	77.70
0.24	0.14	5.93	0.71	80.81	0.36	0.14	2.71	1.33	78.28
0.28	0.14	6.33	0.92	85.35	0.40	0.14	2.73	1.48	80.16
0.32	0.14	3.22	0.82	72.62	0.46	0.14	2.87	1.61	82.21
NaH_2PO_4					K_2HPO_4				
0.28	0.14	1.74	1.63	73.92	0.28	0.14	2.48	0.86	68.08
0.32	0.14	1.87	1.71	76.18	0.32	0.14	3.58	1.17	80.73
0.36	0.14	2.02	1.89	79.24	0.36	0.14	4.04	1.23	83.25
0.40	0.14	2.33	2.26	84.04	0.40	0.14	5.09	1.25	86.42
0.46	0.14	1.31	2.64	77.57	0.46	0.14	3.97	1.42	84.93

TABLE 3: EFFECT OF SALT MASS ON THE EXTRACTION EFFICIENCY OF TRYPTOPHAN

DES (g/ml)	Salt (g/ml)	K	R	E %	DES (g/ml)	Salt (g/ml)	K	R	E %
Na_2CO_3					$\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$				
0.28	0.05	3.87	0.99	79.30	0.46	0.10	1.81	1.90	77.47
0.28	0.08	4.27	0.97	80.55	0.46	0.14	2.90	1.64	82.63
0.28	0.11	5.81	0.95	84.66	0.46	0.18	3.92	1.57	86.20
0.28	0.14	6.53	0.93	85.86	0.46	0.22	4.07	1.21	83.12
0.28	0.17	8.57	0.51	81.38	0.46	0.26	4.85	1.01	83.05
NaH_2PO_4									
0.40	0.10	2.18	3.50	88.41	0.40	0.10	2.22	1.74	79.42
0.40	0.14	2.32	2.28	84.10	0.40	0.14	4.94	1.28	86.34
0.40	0.18	3.09	0.93	74.18	0.40	0.18	5.66	1.15	86.69
0.40	0.22	1.99	0.93	64.85	0.40	0.22	9.11	0.92	89.34
0.40	0.26	2.50	0.65	61.76	0.40	0.26	12.53	0.86	91.55

rich phase with the temperature enhanced^[49]. The increased temperature intensify the phenomenon of Brownian motion, and the molecules in the ABS are active to transfer between the two phase leading to homogeneous distribution^[50]. ABS-1 composition was 0.40 g/ml DES+0.26 g/ml K_2HPO_4 ; ABS-2 was 0.4 g/ml DES+0.10 g/ml NaH_2PO_4 ; ABS-3 was 0.46 g/ml DES+0.18 g/ml $Na_3C_6H_5O_7$ and ABS-4 was 0.28 g/ml DES+0.14 g/ml Na_2CO_3 ; $C_{\text{tryptophan}}$: 0.05 g/l; 288-328 K.

In order to investigate the tryptophan confirmation before and after extraction, UV/Vis and fluorescence spectra of tryptophan were studied^[51]. Fig. 6a illustrates the UV/Vis spectra of tryptophan in water before extraction and in DES-rich phase of ABSs after extraction. It is clear that the curves appeared similar shape, the maximum absorption peak before and after extraction is at the same position of 278 nm. Fig. 6b shows the fluorescence emission spectra for tryptophan in the presence of DES at $\lambda_{\text{ex}}=273$ nm, the characteristic peak of tryptophan in water and in DES-rich solution both appeared at 358 nm. While the excitation spectra at $\lambda_{\text{em}}=358$ nm followed the similar situation. The results

indicated that there were no chemical interactions between the tryptophan and DES in the extraction process. The DES-based ABSs can provide a gentle environment for the extraction of tryptophan.

DES was successfully synthesized with proper mass ratios of ChCl and PEG, and DES-salt ($Na_3C_6H_5O_7$, Na_2CO_3 , NaH_2PO_4 , or K_2HPO_4) ABSs were established and applied for tryptophan extraction. DES- Na_2CO_3 ABS showed the strongest phase-forming ability. After the optimization of extraction conditions, the extraction efficiency of tryptophan can reach 93.88 % (DES- K_2HPO_4 ABS), 90.83 % (DES- NaH_2PO_4 ABS), 88.88 % (DES- $Na_3C_6H_5O_7$ ABS) and 86.72 % (DES- Na_2CO_3 ABS), respectively, under the optimum conditions. Finally, UV/Vis and fluorescence spectra confirmed the structure of tryptophan had no change after extraction. DES-based ABSs can provide a mild environment and have potential applications in the high efficient extraction and purification of biological substances.

Financial support sponsorship:

Financial support for this investigation was given by the National Nature Science Foundation of China (Nos., 21506083, 21506080 and 21878133), Postdoctoral Science Foundation of China (2017M611726, 1601254C), and Scientific Research Foundation of Jiangsu University for the Senior Personnel (No., 15JDG176).

Conflicts of interest:

There are no conflicts of interest.

REFERENCES

- Li Y, Wu Y, Chen K, Wu B, Ji L, Zhu J. Partition behavior of spiramycin in an aqueous two-phase system based on polyethylene glycol and sulfates. *Sep Sci Technol* 2018;53(3):496-502.
- Pang J, Han C, Chao Y, Jing L, Ji H, Zhu W, *et al.* Partitioning Behavior of Tetracycline in Hydrophilic Ionic Liquids Two-Phase Systems. *Sep Sci Technol* 2015;50(13):1993-8.
- Wu D, Hu L, Wang Y, Wei Q, Yan L, Yan T, *et al.* EDTA modified beta-cyclodextrin/chitosan for rapid removal of Pb(II) and acid red from aqueous solution. *J Colloid Interface Sci* 2018;523:56-64.
- Fagan JA, Khripin CY, Batista CAS, Simpson JR, Haroz EH, Walker ARH, *et al.* Isolation of Specific Small-Diameter Single-Wall Carbon Nanotube Species via Aqueous Two-Phase Extraction. *Adv Mater* 2014;26(18):2800-4.
- Subbaiyan NK, Cambre S, Parra-Vasquez AN, Haroz EH, Doorn SK, Duque JG. Role of Surfactants and Salt in Aqueous Two-Phase Separation of Carbon Nanotubes toward Simple Chirality Isolation. *ACS Nano* 2014;8(2):1619-28.

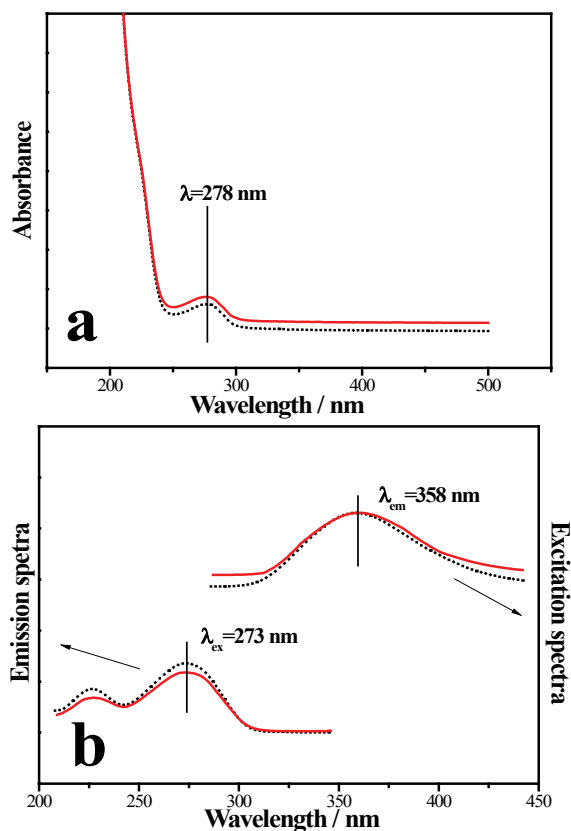


Fig. 6: UV/Vis and fluorescence spectra tryptophan before and after extraction

UV/Vis spectrum (a) and fluorescence spectrum (b) for tryptophan before and after extraction, (.....) tryptophan in pure water, (—) tryptophan in DES-rich phase

6. do Nascimento MB, Castro SdS, Veloso CM, Ilheu Fontan RdC, Soares do Nascimento DJ, Ramos Gandolfi OR, *et al.* Equilibrium data and thermodynamic studies of alpha-amylase partition in aqueous two-phase systems. *Fluid Phase Equilib* 2018;463:69-79.
7. Pang JY, Sha XF, Chao YH, Chen GY, Han CR, Zhu WS, *et al.* Green aqueous biphasic systems containing deep eutectic solvents and sodium salts for the extraction of protein. *RSC Adv* 2017;7(78):49361-7.
8. Jiang W, Zheng D, Xun S, Qin Y, Lu Q, Zhu W, *et al.* Polyoxometalate-based ionic liquid supported on graphite carbon induced solvent-free ultra-deep oxidative desulfurization of model fuels. *Fuel* 2017;190:1-9.
9. Xun S, Zhu W, Chang Y, Li H, Zhang M, Jiang W, *et al.* Synthesis of supported SiW 12 O 40 -based ionic liquid catalyst induced solvent-free oxidative deep-desulfurization of fuels. *Chem Eng J* 2016;288:608-17.
10. Jiang W, Zhu W, Chang Y, Chao Y, Yin S, Liu H, *et al.* Ionic liquid extraction and catalytic oxidative desulfurization of fuels using dialkylpiperidinium tetrachloroferrates catalysts. *Chem Eng J* 2014;250:48-54.
11. Jiang W, Zhu W, Li H, Wang X, Yin S, Chang Y, *et al.* Temperature-responsive ionic liquid extraction and separation of the aromatic sulfur compounds. *Fuel* 2015;140:590-6.
12. Gutowski KE, Broker GA, Willauer HD, Huddleston JG, Swatoski RP, Holbrey JD, *et al.* Controlling the aqueous miscibility of ionic liquids: aqueous biphasic systems of water-miscible ionic liquids and water-structuring salts for recycle, metathesis, and separations. *J Am Chem Soc* 2003;125:6632-3.
13. Yao T, Zang H, Yao S, Dai X, Song H. Measurement and correlation of phase equilibria in aqueous two-phase systems containing functionalized magnetic ionic liquids and potassium phosphate at different temperatures. *J Mol Liq* 2018;263:72-80.
14. Shukla SK, Pandey S, Pandey S. Applications of ionic liquids in biphasic separation: Aqueous biphasic systems and liquid-liquid equilibria. *J Chromatogr A* 2018;1559:44-61.
15. Saravana PS, Ho TC, Chae SJ, Cho YJ, Park JS, Lee HJ, *et al.* Deep eutectic solvent-based extraction and fabrication of chitin films from crustacean waste. *Carbohydr Polym* 2018;195:622-30.
16. Haghbakhsh R, Raeissi S. Densities and volumetric properties of (choline chloride plus urea) deep eutectic solvent and methanol mixtures in the temperature range of 293.15-323.15 K. *J Chem Thermodyn* 2018;124:10-20.
17. Yilmaz E, Soylik M. A novel and simple deep eutectic solvent based liquid phase microextraction method for rhodamine B in cosmetic products and water samples prior to its spectrophotometric determination. *Spectrochim Acta A Mol Biomol Spectrosc* 2018;202:81-6.
18. Pena-Pereira F, Namiesnik J. Ionic Liquids and Deep Eutectic Mixtures: Sustainable Solvents for Extraction Processes. *ChemSusChem* 2014;7(7):1784-800.
19. Waite SL, Li H, Page AJ. NO₂ Solvation Structure in Choline Chloride Deep Eutectic Solvents-The Role of the Hydrogen Bond Donor. *J Phys Chem B* 2018;122(15):4336-44.
20. Gano ZS, Mjalli FS, Al-Wahaibi T, Al-Wahaibi Y, AlNashef IM. Solubility of Thiophene and Dibenzothiophene in Anhydrous FeCl₃- and ZnCl₂-Based Deep Eutectic Solvents. *Ind Eng Chem Res* 2014;53(16):6815-23.
21. Lopez-Salas N, Jardim EO, Silvestre-Albero A, Gutierrez MC, Ferrer ML, Rodriguez-Reinoso F, *et al.* Use of Eutectic Mixtures for Preparation of Monolithic Carbons with CO₂-Adsorption and Gas-Separation Capabilities. *Langmuir* 2014;30(41):12220-8.
22. Chao YH, Zhu WS, Wu XY, Hou FF, Xun SH, Wu PW, *et al.* Application of graphene-like layered molybdenum disulfide and its excellent adsorption behavior for doxycycline antibiotic. *Chem Eng J* 2014;243:60-7.
23. Xu H, Zhang DD, Wu FM, Wei XF, Zhang J. Deep desulfurization of fuels with cobalt chloride-choline chloride/polyethylene glycol metal deep eutectic solvents. *Fuel* 2018;225:104-10.
24. Zhu WS, Wang C, Li HP, Wu PW, Xun SH, Jiang W, *et al.* One-pot extraction combined with metal-free photochemical aerobic oxidative desulfurization in deep eutectic solvent. *Green Chem* 2015;17(4):2464-72.
25. Zounr RA, Tuzen M, Deligonul N, Khuhawar MY. A highly selective and sensitive ultrasonic assisted dispersive liquid phase microextraction based on deep eutectic solvent for determination of cadmium in food and water samples prior to electrothermal atomic absorption spectrometry. *Food Chem* 2018;253:277-83.
26. Makos P, Fernandes A, Przyjazny A, Boczkaj G. Sample preparation procedure using extraction and derivatization of carboxylic acids from aqueous samples by means of deep eutectic solvents for gas chromatographic-mass spectrometric analysis. *J Chromatogr A* 2018;1555:10-9.
27. Ali SS, Morsy R, Elzawawy NA, Fareed MF, Bedaiwy MY. Synthesized zinc peroxide nanoparticles (ZnO₂-NPs): a novel antimicrobial, anti-elastase, anti-keratinase, and anti-inflammatory approach toward polymicrobial burn wounds. *Int J Nanomedicine* 2017;12:6059-73.
28. Ruan L, Shen Y, Lu Z, Shang D, Zhao Z, Lu Y, *et al.* Roles of partitioning-defective protein 6 (Par6) and its complexes in the proliferation, migration and invasion of cancer cells. *Clin Exp Pharmacol Physiol* 2017;44(9):909-13.
29. Wang N, Zhu F, Chen K. 1-deoxynojirimycin: Sources, extraction, analysis and biological functions. *Nat Prod Commun* 2017;12(9):1521-6.
30. Li CP, Li Z, Wang AL, Yin JM, Wang J, Li HX, *et al.* Aqueous two phase extraction process of tryptophan based on functionalized ionic liquids. *RSC Adv* 2013;3(18):6356-61.
31. Wan JY, Fan Y, Yu QT, Ge YZ, Yan CP, Alolga RN, *et al.* Integrated evaluation of malonyl ginsenosides, amino acids and polysaccharides in fresh and processed ginseng. *J Pharm Biomed Anal* 2014;107:89-97.
32. Hou J, Sun E, Zhang ZH, Wang J, Yang L, Cui L, *et al.* Improved oral absorption and anti-lung cancer activity of paclitaxel-loaded mixed micelles. *Drug Deliv* 2017;24(1):261.
33. Li H, Zhu W, Zhu S, Xia J, Chang Y, Jiang W, *et al.* The selectivity for sulfur removal from oils: An insight from conceptual density functional theory. *AIChE J* 2016;62(6):2087-100.
34. Zhang M, Wei Y, Li R, Zhu W, Li H, Zhang Q, *et al.* Magnetic POM-based mesoporous silica for fast oxidation of aromatic sulfur compounds. *Fuel* 2017;209:545-51.
35. Sun X, Deng Q, Liang Z, Liu Z, Geng H, Zhao L, *et al.* Cigarette smoke extract induces epithelial-mesenchymal transition of human bladder cancer T24 cells through activation of ERK1/2 pathway. *Biomed Pharmacother* 2016;86:457-65.
36. Li F, Raza A, Wang YW, Xu XQ, Chen GH. Optimization of Surfactant-Mediated, Ultrasonic-assisted Extraction of Antioxidant Polyphenols from Rattan Tea (*Ampelopsis*

- grossedentata*) Using Response Surface Methodology. *Pharmacogn Mag* 2017;13(51):446-53.
37. Yang L, Qu H, Mao G, Zhao T, Li F, Zhu B, *et al.* Optimization of subcritical water extraction of polysaccharides from *Grifola frondosa* using response surface methodology. *Pharmacogn Mag* 2013;9(34):120.
 38. Liu H, Liu D, Chen C, He Y, Zhu F, Zhang P, *et al.* Preparation and Evaluation of Carbinoxamine Maleate Sustained Release Suspensions with Ion Exchange Resin as the Carriers. *Lat Am J Pharm* 2017;36(4):797-809.
 39. You Q, Yin X, Zhao Y, Jiang Z. Extraction and purification of polysaccharides from pine medicinal mushroom, *Tricholoma matsutake* (higher Basidiomycetes) fruit bodies. *Int J Med Mushrooms* 2014;16(2):149-60.
 40. Feng W, Zhao T, Zhou Y, Li F, Zou Y, Bai S, *et al.* Optimization of enzyme-assisted extraction and characterization of collagen from Chinese sturgeon (*Acipenser sturio* Linnaeus) skin. *Pharmacogn Mag* 2013;9(1):32-7.
 41. Tang X, Yan L, Jing G, Ge H, Yang H, Na L. Optimization of extraction process and investigation of antioxidant effect of polysaccharides from the root of *Limonium sinense* Kuntze. *Pharmacogn Mag* 2011;7(27):186-92.
 42. Yan JK, Wang YY. Random-centroid optimization of ultrasound-assisted extraction of polysaccharides from *Cordyceps sinensis* mycelium. *Curr Top Nutraceutical Res* 2015;13(4):167-71.
 43. Li CP, Li D, Zou SS, Li Z, Yin JM, Wang AL, *et al.* Extraction desulfurization process of fuels with ammonium-based deep eutectic solvents. *Green Chem* 2013;15(10):2793-9.
 44. Pei Y, Li Z, Liu L, Wang J. Partitioning behavior of amino acids in aqueous two-phase systems formed by imidazolium ionic liquid and dipotassium hydrogen phosphate. *J Chromatogr A* 2012;1231:2-7.
 45. Abbott AP, Cullis PM, Gibson MJ, Harris RC, Raven E. Extraction of glycerol from biodiesel into a eutectic based ionic liquid. *Green Chem* 2007;9(8):868-72.
 46. Abdolrahimi S, Nasernejad B, Pazuki G. Influence of process variables on extraction of Cefalexin in a novel biocompatible ionic liquid based-aqueous two phase system. *Phys Chem Chem Phys* 2015;17(1):655-69.
 47. Trindade JR, Visak ZP, Blesic M, Marrucho IM, Coutinho JAP, Lopes JNC, *et al.* Salting-out effects in aqueous ionic liquid solutions: Cloud-point temperature shifts. *J Phys Chem B* 2007;111(18):4737-41.
 48. Ventura SPM, Neves CMSS, Freire MG, Marrucho IM, Oliveira J, Coutinho JAP. Evaluation of Anion Influence on the Formation and Extraction Capacity of Ionic-Liquid-Based Aqueous Biphasic Systems. *J Phys Chem B* 2009;113(27):9304-10.
 49. Cao Y, Xing H, Yang Q, Li Z, Chen T, Bao Z, *et al.* Biphasic Systems That Consist of Hydrophilic Ionic Liquid, Water, and Ethyl Acetate: The Effects of Interactions on the Phase Behavior. *Ind Eng Chem Res* 2014;53(26):10784-90.
 50. Chen J, Wang Y, Zeng Q, Ding X, Huang Y. Partition of proteins with extraction in aqueous two-phase system by hydroxyl ammonium-based ionic liquid. *Anal Methods* 2014;6(12):4067-76.
 51. Du Z, Yu YL, Wang JH. Extraction of proteins from biological fluids by use of an ionic liquid/aqueous two-phase system. *Chemistry* 2007;13(7):2130-7.