Preclinical Evaluation of Scorzonera sp. Root Extracts and Major Compounds against Acute Hepatotoxicity Induced by Carbon Tetrachloride

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Evaluation of hepatoprotective activities of Scorzonera roots and their major compounds, was aimed in current study. Scorzonera latifolia, S. tomentosa, S. mollis ssp. szovitii, S. parviflora and S. cana var. jacquiniana roots, methanol-water (80:20) extracts together with chlorogenic acid, scorzotomentosin-4'-O-β-glucoside, hydrangenol-8-O-β-glucoside as major compounds isolated from S. latifolia roots were tested for their hepatoprotective activities. Sprague Dawley rats were used for experiment and hepatotoxicity was induced by carbon tetrachloride. Aspartate aminotransferase and alanine aminotransferase levels were measured and all results were confirmed by histopathological examination. Plasma aspartate aminotransferase and alanine aminotransferase levels of examined groups were not significant when compared to carbon tetrachloride-treated groups. However histopathological results have revealed that all tested groups have less damage when compared to carbon tetrachloride group except scorzotomentosin-4'-O-β-glucoside and hydrangenol-8-O-β-glucoside groups. Scorzonera species displayed moderate hepatoprotective activities against carbon tetrachloride induced acute toxicity. Chlorogenic acid, among tested compounds exhibited higher activity than all tested Scorzonera species as well as other isolated compounds. Therefore chlorogenic acid could be suggested as responsible compound.

Key words: Scorzonera sp., hepatoprotective effect, carbon tetrachloride, chlorogenic acid

In Turkish folk medicine, the roots of Scorzonera latifolia and similar species are mainly used as analgesic, antihelmintic, wound healer, and for treatment of women infertility[1,2]. Additionally, the usage of this genus plants against hypertension, kidney diseases, diabetes mellitus, arteriosclerosis, and rheumatism have been recorded[1]. Analgesic activities of the S. latifolia, S. tomentosa, S. mollis ssp. szovitii, S. suberosa ssp. suberosa were proven scientifically by previous studies and triterpenoids were isolated as responsible compounds[3-5]. Antiinflammatory, antioxidant, and wound healing activities have also been reported[6-9]. Phytochemical analyses revealed that S. latifolia roots contained taraxasteryl myristate, taraxasteryl acetate, motioli, 3-β-hydroxy fern-8 en-7-one acetate, urs-12-en-11-one-3-acetyl, 3-β-hydroxy-fern-7-en-6-one-acetate, olean-12-en-11-one-3-acetyl, fern-7-en-3-β-one, leucodin, β-sitosterol[3,5,10], chlorogenic acid, chlorogenic acid methyl ester, 1,5-dicaffeoyl quinic acid, 3,5-dicaffeoyl quinic acid, methylester of 3,5-dicaffeoyl quinic acid, hydrangenol-8-O-β-glucoside, hydrangenol-4'-O-β-glucoside, scorzotomentosin-4'-O-β-glucoside and a new isocoumarine derivative[11] as well as scorzoveratrin 4'-O-β-glucoside, scorzoveratrin, 4,5-dicaffeoylquinic acid, 4,5-dicaffeoylquinic acid methyl ester and cafeic acid[12]. Scorzotomentosin, scorzotomentosin-4'-O-β-glucoside, scorzoptalide, scorzoerzincanin, hydrangenol, hydrangenol-4'-O-β-glucoside, hydramacrophyllolA and B have been isolated from S. tomentosa roots previously[13]. Chlorogenic acid has been determined in roots of Scorzonera sp.,

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including *S. latifolia*, *S. tomentosa*, *S. mollis* ssp. *szovitsii*, *S. parviflora* and *S. cana* var. *jacquiniana* as one of the major compounds by high-performance liquid chromatography (HPLC)\[^{6,14}\]. Chlorogenic acid has potent antioxidant and hepatoprotective activities. Liver damage and symptoms of liver fibrosis were reduced significantly by chlorogenic acid as well as serum aspartate aminotransferase (AST) and alanine aminotransferase (ALT), alkaline phosphatase (ALP) and total bilirubin levels were lowered\[^{15,16}\]. In current study the roots of *S. latifolia*, *S. tomentosa*, *S. mollis* ssp. *szovitsii*, *S. parviflora* and *S. cana* var. *jacquiniana* were investigated for their potential hepatoprotective activities due to their high content of chlorogenic acid as well as caffeoylquinic acid derivatives. Chlorogenic acid and isocoumarine derivatives; hydrangenol-8-β-glucoside and scorzotomentosin-4’-O-β-glucoside obtained from *S. latifolia* roots as major components were also evaluated for their activities.

**MATERIALS AND METHODS**

*Scorzonera* species were collected from different parts of Turkey. The taxonomic identification of the plants was confirmed in the Department of Biological Sciences, Faculty of Art and Sciences, Gazi University. Voucher specimens are kept in the herbarium of Ankara University, Faculty of Pharmacy (Table 1). Carbon tetrachloride (CCl\(_4\)) was obtained from Merck (Darmstadt, Germany), and olive oil was obtained from Fluka (Steinheim-Germany). CCl\(_4\) dissolved in olive oil (v/v, 1:1) and *Scorzonera* extracts were prepared using water (w/v).

**Extraction of plant material:**

Dried and powdered *S. latifolia* roots (700 g) were extracted with methanol-water (80:20) at room temperature for 24 h×3 times by continuous stirring. Methanol-water extract was filtered and concentrated to dryness under reduced pressure and low temperature (40-50\(^\circ\)) on a rotary evaporator to give crude extract (182.46 g).

**Isolation and identification of the compounds:**

Solvent-solvent partition was performed with petroleum ether, chloroform, ethylacetate, respectively, and with the remain methanolic part four different extract were obtained. Ethylacetate part (11.02 g) was subjected to column chromatography on silica gel column eluting with ethylacetate:methanol:water (100:13.5:10) to yield 92 fraction. Fraction 24-26 and fraction 38-42 gave hydrangenol-8-O-β-glucoside and scorzotomentosin-4’-O-β-glucoside, respectively. Chlorogenic acid was obtained from 89-92 fractions as a dirty white precipitate. Structures of the compounds were established using MS (Waters 2695 Alliance Micromass ZQ, LC/MS), \(^1\)H and \(^13\)C NMR (Varian Mercury 400, 400 MHz High Performance Digital FT-NMR Spectrometer) techniques.

**Experimental animals:**

The study protocol (01-06-2015/41) was approved by the Ethical Committee of, İstanbul Medipol University. Male and female Sprague-Dawley rats (170-240 g) were used in this experiment. The animals were housed in standard cages (48×35×22 cm) at room temperature (22±2\(^\circ\)), with artificial light from 7.00 am to 7.00 pm, and provided with pelleted food and water ad libitum.

**Hepatoprotective activity assay:**

Hepatotoxicity test model induced by CCl\(_4\) was used with slight modification\[^{16,17}\]. Liver toxicity was induced by i.p. (intraperitoneal) administration of CCl\(_4\) (0.8 ml/kg) diluted in olive oil (1:1 v/v) for two days. Animal groups were designed as follow (n=5): control group 1 received isotonic saline solution (ISS) 0.1 ml, group 2 received CCl\(_4\) 0.8 ml/kg i.p. Group 3 received *S. cana* var. *jacquiniana* root extract (100 mg/kg)+CCl\(_4\) (0.8 ml/kg), group 4 received *S. latifolia* root extract (100 mg/kg)+CCl\(_4\) (0.8 ml/kg), group 5 received *S. mollis* ssp. *mollis* root extract (100 mg/kg)+CCl\(_4\) (0.8 ml/kg), group 6 received *S. parviflora* root extract (100 mg/kg)+CCl\(_4\) (0.8 ml/kg), group 7 received *S. tomentosa* (100 mg/kg)+CCl\(_4\) (0.8 ml/kg), group 8 received chlorogenic acid (5 mg/kg)+CCl\(_4\) (0.8 ml/kg), group 9 received hydrangenol-8-O-glucoside (5 mg/kg)+CCl\(_4\) (0.8 ml/kg), group 10 received scorzotomentosin-4’-O-β-glucoside (5 mg/kg)+CCl\(_4\) (0.8 ml/kg) i.p. daily for seven days.

Blood samples were collected by direct cardiac puncture after seven days treatment and the serum was used for the assay of the marker enzymes AST and ALT. The percentage of daily changes in body weight \(\%\)=100\((weight_n-weight_{initial})/weight_{initial}\) where, weight_{initial}; measurement on the first day, weight_{n}; measurement after 2., 3., … 8th d.

**Histopathological examination of the liver:**

The livers of the experimental animals were fixed in 10% neutral buffered-formalin prior to routine processing in paraffin-embedded blocks. Sections
(5 µm thick) were cut and stained using Hematoxylin eosin (HE). Axio V 16 microscope was used to take photographs. Histological damage was measured as one-blind experiment and was expressed using the following score system; 0: absent; +: mild; ++: moderate; +++: severe. Ballooning degeneration, sinusoidal dilatation, vascular congestion and steatosis were used for evaluation of the histopathological results.

**Statistical analyses:**

Results are reported as mean±SEM (standard error of mean) and as percentage (%). Kruskal-Wallis test (post hoc Mann-Whitney U with Bonferroni adjustment and Moses Extreme Reactions Test) and one-way analysis of variance (One-Way ANOVA, post hoc Scheffe) were used for statistical analyses. Probability levels of less than 0.05 (P<0.05) were considered significant.

**RESULTS AND DISCUSSION**

Plasma AST and ALT levels were given in Table 2. Differences in plasma AST levels of CCl₄ group and *Scorzonera* sp. extracts as well as chlorogenic acid, hydrangenol-8-O-β-glucoside and scorzotomentosin-4'-O-β-glucoside were not significant. However plasma ALT levels were measured higher than CCl₄ group after treatment of *S. mollis* ssp. *szowitsii*, *S. tomentosa* and *S. cana* var. *jacquiniana* root extracts.

Chlorogenic acid, hydrangenol-8-O-β-glucoside and scorzotomentosin-4'-O-β-glucoside groups serum ALT levels were reduced when compared to CCl₄ group.

Results of the body weight changes of animals are given in Table 3. Body weight of the animals was measured in the beginning and at the end of the study. While animals in control group (ISS) were gaining weight as measured by 2.64%, in all other groups animals had weight loss in different percentage as given in Table 3.

Histopathological examination results using score system were exhibited in Table 4. Significant differences between control (ISS) group and treatment groups were observed (P<0.05). Ballooning degeneration, sinusoidal dilatation, vascular congestion and steatosis were observed by severe score in CCl₄ treated group. Histopathological results have revealed CCl₄ treated group has high histopathological score as shown in Table 4 and remarkable results were assigned between all other treatment groups except hydrangenol-8-O-β-glucoside and scorzotomentosin-4'-O-β-glucoside treated groups (P>0.05).

In control (ISS) group no degeneration was observed in hepatocytes, sinusoids and vascular cells (fig. 1A). On the other hand CCl₄ treated group were displayed severe damage; ballooning degeneration, vascular congestion, sinusoidal dilatation and steatosis (figs. 1B and 2).

**TABLE 1: VOUCHER SPECIMENS IN THE HERBARIUM**

<table>
<thead>
<tr>
<th>Species</th>
<th>Place</th>
<th>Herbarium number</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>S. latifolia</em> (Fisch. &amp; Mey.) DC.</td>
<td>Kop passage, 2010</td>
<td>AEF 23830</td>
</tr>
<tr>
<td><em>S. mollis</em> Bieb. subsp. <em>szowitsii</em> (DC.) Chamberlain</td>
<td>Kizilcuharnam, Ankara, 2006</td>
<td>AEF 23844</td>
</tr>
<tr>
<td><em>S. parviflora</em> Jacq.</td>
<td>Gölbasi, Ankara, 2012</td>
<td>AEF 25894</td>
</tr>
<tr>
<td><em>S. tomentosa</em> L.</td>
<td>Akdagmadeni, Yozgat, 2013</td>
<td>AEF 23841</td>
</tr>
</tbody>
</table>

**TABLE 2: EFFECTS OF SCORZONERA SP. EXTRACTS AND COMPOUNDS ON SERUM LEVELS OF AST AND ALT**

<table>
<thead>
<tr>
<th>Groups</th>
<th>ALT</th>
<th>AST</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Serum (U/l)</td>
<td>Serum (U/l)</td>
</tr>
<tr>
<td>Control (ISS)</td>
<td>40.45</td>
<td>96.80</td>
</tr>
<tr>
<td>CCl₄</td>
<td>51.15</td>
<td>110.25</td>
</tr>
<tr>
<td><em>S. cana</em> var. <em>jacquiniana</em></td>
<td>66.70b</td>
<td>114.10</td>
</tr>
<tr>
<td><em>S. latifolia</em></td>
<td>52.20</td>
<td>97.00</td>
</tr>
<tr>
<td><em>S. mollis</em> ssp. <em>szowitsii</em></td>
<td>56.30b</td>
<td>103.90</td>
</tr>
<tr>
<td><em>S. parviflora</em></td>
<td>50.60</td>
<td>102.00</td>
</tr>
<tr>
<td><em>S. tomentosa</em></td>
<td>62.20</td>
<td>108.00</td>
</tr>
<tr>
<td>Chlorogenic acid</td>
<td>58.70</td>
<td>329.20</td>
</tr>
<tr>
<td>Hydrangenol-8-O-glucoside</td>
<td>39.35b</td>
<td>111.20</td>
</tr>
<tr>
<td>Scorzotomentosin-4'-O-β-glucoside</td>
<td>33.25b</td>
<td>124.25</td>
</tr>
<tr>
<td>P value</td>
<td>0.034</td>
<td>0.457</td>
</tr>
</tbody>
</table>

Post-hoc Moses extreme reactions test results; *P*<0.05 in relation to control (ISS); *P*<0.05 in relation to CCl₄.
Ballooning degeneration, vascular congestion, sinusoidal dilatation and steatosis were also occasionally observed in groups after treatment by hydrangenol-8-O-glucoside and scorzotomentosin-4'-O-β-glucoside. Steatosis areas were observed generally around central vein. Sinusoidal dilatation was also observed mainly around central vein (fig. 3).

Chlorogenic acid treatment and S. latifolia, S. parviflora, S. mollis ssp. szowitsii, S. cana var. jacquiniana, S. tomentosa root extracts induce recovery on cellular degeneration (figs. 4 and 5). Notable changes including less ballooning degeneration, sinusoidal dilatation, vascular congestion and steatosis were observed compared to CCl₄ treated groups. Fig. 5A displayed
S. latifolia, S. parviflora root extract treated groups liver. Vascular congestion and sinusoidal dilatation were detected rarely when compared as CCl₄ treated animal livers. Nevertheless more ballooning degeneration, sinusoidal dilatation, vascular congestion and steatosis were seen compared to control (ISS) treated groups.

Fig. 2: Histopathological examination of liver (CCl₄ group)
A: Sinusoidal dilatation. B: ballooning degeneration. C: steatosis

Fig. 3: Histopathological examination of liver
(A) Central vein and steatosis, (B) ballooning degeneration and steatosis, (C) sinusoidal dilatation. (a) Group scorzotomentosin-4'-O-β-glucoside and (b) group hydrangenol-8-O-glucoside
S. mollis ssp. szowitsii and S. cana var. jacquiniana treated groups histopathological results have revealed that ballooning degeneration was seen, however vascular congestion, sinusoidal dilatation and steatosis were scarcely as shown in figs. 5B and C. Fibrosis or fibrotic areas were also detected rarely in S. mollis ssp. szowitsii treated group animal livers (fig. 5B). Fig. 5D exhibited histopathological results of liver treated by S. tomentosa root extract. Ballooning degeneration, vascular congestion and sinusoidal dilatation were frequently seen. On the other hand steatosis was observed rarely (fig. 5D).

Acute liver toxicity model induced by CCl₄ was used to evaluate hepatoprotective activities of S. latifolia, S. tomentosa, S. mollis ssp. szowitsii, S. parviflora and S. cana var. jacquiniana root extract as well as chlorogenic acid, hydrangenol-8-O-β-glucoside and scorzotomentosin-4'-O-β-glucoside obtained from S. latifolia roots as main components in current study.

CCl₄-induced hepatotoxicity test model has been extensively used in animal studies to evaluate drugs can be used for treatment of liver diseases[16,19]. CCl₄ caused to severe damages in liver such as steatosis, inflammation, apoptosis and cell necrosis[19,20]. Giving CCl₄ to animals as a single dose mediated to necrosis and steatosis in a short time. Hepatocytes functional defects have been occurred by injuring plasma membrane, endoplasmic reticulum (ER), mitochondria and Golgi apparatus[20,21] resulted in elevation of serum AST and ALT levels indicating hepatocellular damage[19]. Movement of fat from the cell is blocked by disruption of mechanism for coupling triglycerides to the appropriate apoprotein to form the lipoprotein carrier molecule is caused steatosis[21]. The acute hepatotoxic effect of CCl₄ is due to its metabolite trichloromethyl radical (CCl₃•) which occurred by metabolization of CCl₄ via cytochrome P-450 system. CCl₃• radical is converted to trichromethyl peroxy radical CCl₃COO• in the presence of oxygen, is more active than the first one. These free radicals are react with different substances including proteins, nucleic acids, lipids and lead to dysfunction. Alkylation of macromolecules including cellular proteins by simultaneous attack on polyunsaturated fatty acids starts the process of lipid peoxidation leading to liver cell necrosis[16,20]. Therefore free radical scavengers or generation inhibitors could be useful in CCl₄-induced liver injury[20]. CCl₄ also caused activation of many catabolic enzymes which disrupt cytoskeletal construction and cell death via apoptosis or necrosis by increasing the levels of Ca²⁺ in cells[20].

Chlorogenic acid an ester of caffeic acid and quinic acid (3-O-caffeylquinic acid (CGA)) possess significant antioxidant activity as well as antibacterial, anticarcinogenic, antiinflammatory, antihypertension activities[22,23]. Chlorogenic acid is one of the most abundant hydroxycinnamic acid derivative in human diet and is widely distributed in medicinal plants, a number of fruits and vegetables as well as daily beverages like coffee, tea, wine, and tobacco[22,24]. Related to its hepatoprotective activity many studies have been reported besides its other biological activities. Lipid peroxidation induced by CCl₄ has been supressed by chlorogenic acid. 200 µmol/kg administration of chlorogenic acid resulted in significant protection against the liver damage[23]. According to the Shi et al.[14] chlorogenic acid treatment inhibited development of hepatic fibrosis in pericentral region. Small degree of bridging fibrosis was observed. Lower severity score for liver fibrosis was determined in chlorogenic acid (60 mg/kg) treatment group when compared to CCl₄ group. It has been reported that chlorogenic acid has protective activity against liver fibrosis due to its inhibitory activity on hepatic stellate cells (HSCs) activation and production of transforming growth factor (TGF-β1) and vascular endothelial growth factor (VEGF) as well as ER stress. Mitochondrial pathway of apoptosis in liver fibrosis could be regulated by chlorogenic acid is suggested[14]. Serum AST, ALT and TB levels

![Fig. 4: Histopathological examination of chlorogenic acid-treated animal liver](image)

Chlorogenic acid treatment induces recovery on cellular degeneration. Notable changes including less ballooning degeneration, sinusoidal dilatation, vascular congestion and steatosis were observed compared to CCl₄-treated groups.
were lowered significantly by administration of chlorogenic acid at 300 or 500 mg/kg dose while ALT levels were comparable with CCl₄ treated group. Any mortality has not been observed up to 5000 mg/kg[15]. Wu et al. reported that[25] chlorogenic acid administration at 75 mg/kg and 150 mg/kg dose for 3 w
provided significant recovery in liver fibrosis induced by bile duct ligation. Serum alanine transaminase, AST, ALP, total bilirubin, direct bilirubin and total bile acid levels were decreased. Furthermore collagen I, collagen III, TGF-β1 and VEGF mRNA increasing induced by BDL treatment were also suppressed by chlorogenic acid[25]. Free radicals generated by CCl₄ metabolism via cytochrome P450 2E1 induce liver cell apoptosis and necrosis as well as upregulation of TNF-α, IL-10 and TGF-β in necrotic hepatocytes. Especially TGF-β caused progression of liver injury to chronic liver disease by activating local leucocytes and promoting the circulation of leucocytes to the necrotic area. Chlorogenic acid down regulates TGF-β1 protein expression and decreased NFkB expression. Furthermore pretreatment of chlorogenic acid protected the PC12 cells from β-amyloid (Aβ) which are reported to reduced viability of PC12 cells by reducing the level of intracellular Ca²⁺ and apoptosis related of proteins. Chlorogenic acid was reported to potentiate the antiapoptotic and antifenrogenic effects of silymarin and this combination increased the expression of Bcl-2 down regulated by CCl₄[22]. In current study chlorogenic acid was also investigated for its hepatoprotective activity in acute hepatotoxicity test model induced by CCl₄ together with hydrangenol-8-O-β-glucoside and scorzotomentosin-4′-O-β-glucoside. Scorzonera species were evaluated for their possible hepatoprotective activities due to their chlorogenic acid contents in present study. Biochemical results exhibited that plasma AST levels were not significant between CCl₄ group and Scorzonera sp. extracts as well as hydrangenol-8-O-β-glucoside and scorzotomentosin-4′-O-β-glucoside. On the other hand high levels of ALT were measured in S. mollis ssp. szowitsii, S. tomentosa var. jacquiniana root extract treated groups while lower levels of ALT were detected in hydrangenol-8-O-β-glucoside and scorzotomentosin-4′-O-β-glucoside than CCl₄ group. Histopathological examinations clearly displayed all Scorzonera species tested and chlorogenic acid had less cellular damage than CCl₄ treated groups. S. latifolia, S. parviflora, S. mollis ssp. szowitsii, S. tomentosa, S. cana var. jacquiniana root extracts and chlorogenic acid treatment induce recovery on cellular degeneration. Chlorogenic acid treatment at 5 mg/kg dose did not lowered AST and ALT levels significantly. However histopathological examinations have revealed that recovery on cellular damage has been improved and less sinusoidal dilatation, vascular congestion, balloning degeneration as well as steatosis were detected. All Scorzonera species used in current study were investigated for their chlorogenic acid contents by HPLC analyses qualitatively and quantitatively previously. S. latifolia roots contain higher chlorogenic acid (1246.78±3.20 µg/g) and this was followed by S. tomentosa (734.72±1.04 µg/g), S. parviflora (509.96±6.64 µg/g), S. cana var. jacquiniana (331.028±2.83 µg/g) and S. mollis ssp. szowitsii (159.25±0.24 µg/g)[6-7]. According to the current study Scorzonera root extracts displayed hepatoprotective activities probably due to their chlorogenic acid contents. Derivatives of chlorogenic acid 3,4 and 3,5 dicaffeoylquinic acids were also reported to have protective activity in CCl₄ induced hepatotoxicity[26] which have also been isolated from some Scorzonera species previously[11,12].

In conclusion, hepatoprotective activities of the tested Scorzonera species could be attributed to their chlorogenic acid content and its derivatives. Usage of Scorzonera species could be useful for hepatic diseases, furthermore chlorogenic acid is promising agent in treatment of hepatic diseases which needs further studies.

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Conflicts of interest:

There are no conflicts of interest.

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REFERENCES

5. Bahadır Acikara Ö, Saltan Çitoğlu G, Dall’Acqua S, Özbek H,