

Variation in Copy Number of *MTUS1* Gene among Healthy Individuals and Cancer Patients from Gujarat

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Almal and Padh: *MTUS1* Gene Copy Number and Risk to Cancer in Indians

Genome variations in the form of single nucleotide polymorphism, INDELS (insertions or deletions),

duplications, inversions and copy number variations are more widely prevalent than initially predicted. Mitochondrial tumor suppressor gene maps on chromosome 8p, in which exon 4-specific deletion is associated with susceptibility towards breast and various other forms of cancer. In this study, 41 head and neck cancer, 15 breast cancer patients and 280 healthy control individuals were analysed for mitochondrial tumor suppressor gene copy number variation. The frequencies of wt/wt (homozygous wild-type), wt/Del (heterozygous variant) and Del/Del (homozygous deletion variant) mitochondrial tumor suppressor gene genotypes were found to be 73.3, 26.7 and 0% in breast cancer patients, 41.5, 58.5 and 0% in head and neck cancer patients and 43, 57 and 0% in healthy individuals, respectively. A significant association of the deletion variant with breast cancer (odds ratio=0.27, 95% confidence interval=0.08–0.87, P=0.0207) was found in our population. In addition, the allele and genotype frequency varied significantly (P<0.0001) among Indian and German populations, reflecting ethnic diversity. This pilot case-control study highlighted the indicative role of mitochondrial tumor suppressor gene deletion in protection from breast cancer in Indian population. However, the findings need to be investigated in larger patient sample size before any conclusive role of mitochondrial tumor suppressor gene copy number variation on cancer risk.

Key words: Breast cancer, copy number variation, head and neck cancer, health, pharmacogenetics

Copy number variation (CNVs) arise from the genomic rearrangements resulting in the deletion, duplication, insertion and translocations covering about 12% of the human genome^[1,2]. Recent studies have highlighted the role of CNVs in complex disorders including susceptibility to cancer^[3-5]. The association of CNVs in disease susceptibility involving wide range of common human disorders is well compiled and documented by several researchers^[6,7]. In order to detect these changes several approaches have been developed and applied till date^[8]. Among them, array-based comparative genomics approach by Hinds *et al.* in 2006 was used to detect common deletions ranging from 70 bps to 7 Kb^[9]. Of the 100 CNVs identified, the mitochondrial tumor suppressor gene 1 (*MTUS1*) encompassing the deletion of complete coding exon 4 (162 bp) was one of them. The breakpoint of the deleted region was redefined and validated in large case-control samples from German population^[10]. In 2005, Lupski and Stankiewicz documented the role of CNVs in genetic disorders but the extent to which CNVs contribute to disease development remains poorly understood^[11]. However, the exon-specific deletion of *MTUS1* gene is an illustration of a CNV associated with the development of cancer^[12].

MTUS1 gene, also designated as MTS1, GK1 or angiotensin II (AT2) receptor-interacting protein (ATIP1) is located at chromosomal position 8p22 spanning over 112 Kb in size. It is found to be ubiquitously expressed in normal tissues but transiently up-regulated during initiation of cellular quiescence and differentiation process^[4]. The tumor suppression

function was confirmed by mRNA expression studies in the pancreatic cancer tissues and the pancreatic cancer cell lines MIA PaCa-2, whereas the recombinant expression of *MTUS1* in MIA PaCa-2 cells inhibited proliferation^[13].

MTUS1 is found to be associated with disease progression in human cancers, including bladder, colorectal, oesophageal, head and neck squamous cell, hepatocellular, lung, ovarian, pancreatic, prostate and especially breast carcinomas^[13-17]. Furthermore, it is found that *MTUS1/ATIP1* is an early mediator of AT2 receptor activation. Together with AT2, it antagonizes AT1 receptor function, inhibiting epidermal growth factor signalling via autophosphorylation of epidermal growth factor receptor. This leads to altering the activity of growth factor-induced extracellular regulated kinase, phosphorylation of signal transducer and activation of transcription 3 (STAT3) and protein kinase C, involved in apoptosis and proliferation^[18-24].

The aim of this study was to determine the frequency distribution of the *MTUS1* deletion variant in Indian population. Subsequently in a pilot case-control study we intended to investigate the association with *MTUS1* deletion variant with development of cancer.

MTUS1 deletion was genotyped in 41 patients with

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head and neck cancer (HNC), 15 patients with breast cancer (BC) and 280 healthy control individuals. The healthy control population comprised of the individuals 20-50 y of age (245 males and 35 females) mainly from the Western India. They were recruited at B. V. Patel PERD Centre, Ahmedabad. The case (HNC and BC patients) population comprised of the individuals 18 or older in age, mainly from the Western India. The recruited patients were undergoing treatment at local cancer hospital in Ahmedabad. Newly diagnosed BC and HNC solid tumor patients before medication were selected for the study and significant inclusion and exclusion criteria were taken into consideration. The study was approved by the Institutional Ethics Committee and written informed consent form was obtained prior to blood collection from individuals.

Genomic DNA isolation was carried from 5 ml of blood withdrawn from healthy individuals and cancer patients. Blood (5 ml) was collected in anticoagulant (K₂ EDTA or sodium heparin) precoated collection tubes and genomic DNA was extracted using phenol-chloroform

method^[25]. The RBCs were first ruptured using lysis buffer followed by the lysis of the WBC pellet using digestion buffer. The protein degradation was carried out using proteinase K treatment and the purification of the sample using phenol:chloroform:isoamyl alcohol mixture. The DNA was precipitated using absolute alcohol and subsequently washed with 70% alcohol. The DNA pellet thus obtained was air dried and dissolved in appropriate amount of TE buffer^[25]. *MTUS1* genotyping was performed by polymerase chain reaction (PCR) amplification. Two separate reactions were designed to detect *MTUS1* deletion (figs. 1 and 2) using primer pairs as described earlier^[10]. The confirmation of the PCR products were carried out by sequencing deletion-specific and exon-specific amplicons using Applied Biosystems 3730xl sequencer (Macrogen Inc., Korea).

The difference between the genotype frequencies of HNC and BC cases with control group was calculated by χ^2 test for statistical significance. Hardy-Weinberg equilibrium (HWE) was examined using χ^2 test with

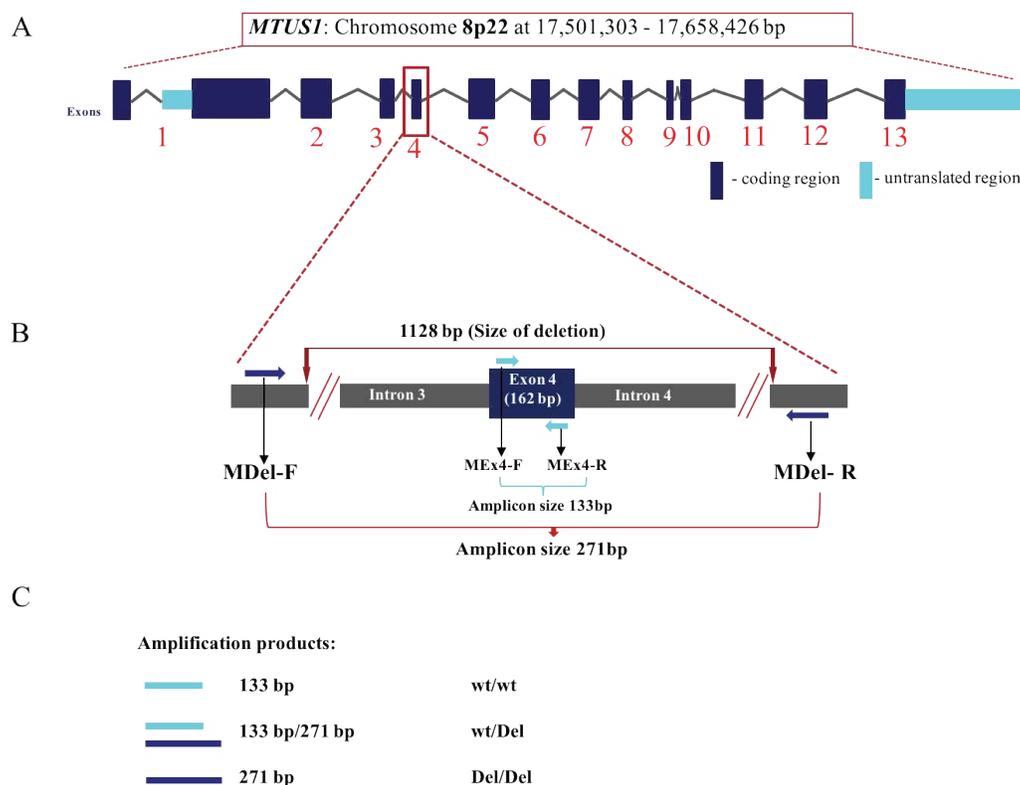


Fig. 1: Schematic representation of the amplification strategy for the detection of *MTUS1* exon specific deletion

A: Schematic representation of *MTUS1* gene encompassing 13 coding exons. **B:** Representation of the *MTUS1* gene deletion comprising the entire exon 4 (162 bp). Detection of the deletion is done by two separate reactions. The PCR primers MDEL-F and MDEL-R are deletion-specific primers located in the intron 3 and intron 4, whereas MEX4-F and MEX4-R exon-specific primers. **C:** *MTUS1* amplification products. MDEL-F and MDEL-R primers generate 271 bp amplicon in the presence of deletion. MEX4-F and MEX4-R primers generate 133 bp amplicon size in the absence of deletion. The homozygous wild-type (wt/wt) genotype will be represented by 133 bp amplicon alone, homozygous deletion (EDL/DEL) will be represented by 271 bp amplicon alone and heterozygous deletion (wt/DEL) will be represented by both 133 bp and 271 bp amplicons (adapted from Frank *et al.* 2007)

one degree of freedom. Genotype-specific odds ratios (ORs), 95% confidence intervals (CI) and P-values were calculated by unconditional logistic regression using IBM SPSS 16.0 software.

The distribution of *MTUS1* allele and genotypes was assessed in 280 healthy Indian control individuals. The frequency for wt allele was 0.71 and that of Del allele was 0.29. The genotype frequency was found to be 49.2% for wt/wt (homozygous wild-type) genotype, 57.1% for wt/Del (heterozygous variant) genotype while no healthy individual carried Del/Del (homozygous deletion variant) genotype. However, it was observed that the genotype frequencies studied in the control individual were not in the agreement with the HWE. Deviation from HWE could have been because of a number of reasons, but sequencing and confirming the results ruled out the technical error resulting in such conclusion. However, the reason for lack of HWE is still not clear.

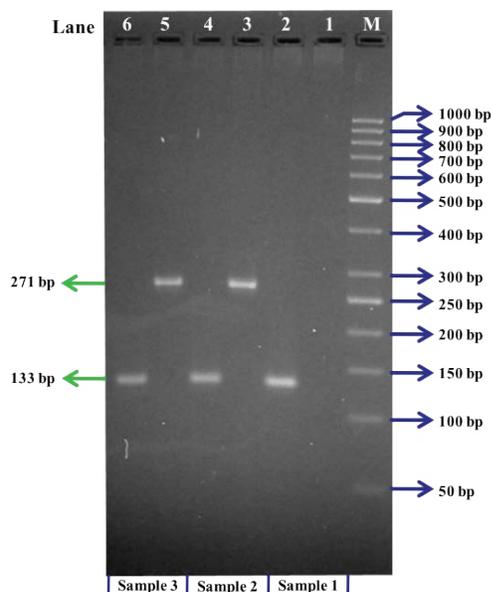


Fig. 2: Agarose gel (2%) of the PCR products for *MTUS1* gene. Lane M shows the marker O' Gene ruler 50 bp ladder. Lane 1, 3, 5 represents *MTUS1* exon 4 specific deletion reaction while lane 2, 4, 6 represents reaction specific for *MTUS1* gene presence. Lane 1, 2 (sample 1)-homozygous wild-type genotype (PCR product: 133 bp), lane 3, 4 (sample 2) and lane 5, 6 (sample 3)-heterozygous genotype (PCR products: 133 bp and 271 bp)

Furthermore, a comparison of the allele and genotype frequency among Indian and German population highlighted a significant ($P < 0.0001$) difference, reflecting the ethnic diversity between individuals of different population (Table 1).

The *MTUS1* genotype frequency in BC patients and HNC patients was found to be 73.3 and 41.5% for wt/wt (homozygous wild-type) genotype, 26.7 and 58.5% for wt/Del (heterozygous variant) genotype while no BC or HNC patient carried Del/Del (homozygous deletion variant) genotype. A significant association of the deletion variant with a decreased risk to breast cancer was observed (OR=0.27, 95% CI=0.08–0.87, $P=0.0207$). However, a non-significant association was observed between *MTUS1* deletion variant and risk to HNC (OR=1.06, 95% CI=0.85–1.33, $P=0.8671$) (Table 2) in Indian population. Since, this study is limited by smaller sample size, the results need to be validated using larger population.

CNV has become an integral part of genetic variation contributing to the susceptibility towards various diseases^[26]. CNV in terms of exon-specific deletion in the tumor suppressor gene (*MTUS1*) is well studied in German population^[10]. As reported, this phenomenon of exon-specific deletion provokes an increased protein activity leading to enhanced tumor suppressor activity. Further analysis revealed that the deleted exon was rich in polyproline-motifs, which are usually involved in interactions with the SH3 or WW functional domains implying that the wild type and deleted variants of *MTUS1* may interact with distinct intracellular partners and exhibit different cellular functions, such as tumor suppression^[27-29]. In conjunction with the previous reports of *MTUS1* gene deletion variant with a decreased risk for both familial and high-risk familial BC in German population we examined the frequency of the deletion variant in Indian population. The frequency of the deletion variant was quite different when compared to the German population (Table 1). Further the 1128 bp deletion was genotyped in healthy Indian population and patients with either HNC or BC.

TABLE 1: FREQUENCY DISTRIBUTION OF *MTUS1* DELETION VARIANTS IN INDIAN AND GERMAN POPULATION^[10] IN HEALTHY INDIVIDUALS

Genotype	Indian population n (%) (data from this study)	German population n (%) (data ^[10])
wt/wt	120 (42.9)	668 (91.3)
wt/Del	160 (57.1)	63 (8.6)
Del/Del	0 (0)	1 (0.1)
DEL/DEL + wt/DEL	160 (57.1)	64 (8.7)

Data obtained from this study represented the Indian population data. wt/wt is the homozygous wild-type genotype; wt/Del is the heterozygous genotype; Del/Del is the homozygous deletion genotype

TABLE 2: GENOTYPE FREQUENCIES OF *MTUS1* DELETION VARIANTS IN PATIENTS AND CONTROL SUBJECTS IN INDIAN AND GERMAN POPULATION^[10]

Population	Genotype	Cases n (%)	Control n (%)	OR (95% CI), P value	
Indian population	Breast cancer	wt/wt	11 (73.3)	120 (42.9)	0.27 (0.08-0.87), 0.0207
		wt/Del	4 (26.7)	160 (57.1)	
		Del/Del	0 (0)	0 (0)	
	Head and neck cancer	wt/wt	17 (41.5)	120 (42.9)	1.06 (0.85-1.33), 0.8671
		wt/Del	24 (58.5)	160 (57.1)	
		Del/Del	0 (0)	0 (0)	
German population*	Familial BC	wt/wt	562 (94.8)	668 (91.3)	0.58 (0.37-0.90), 0.01
		wt/Del	30 (5.1)	63 (8.6)	
		Del/Del	1 (0.1)	1 (0.1)	
	High-risk familial BC	wt/wt	355 (96.2)	668 (91.3)	0.41 (0.23-0.74), 0.003
		wt/Del	14 (3.8)	63 (8.6)	
		Del/Del	0 (0)	1 (0.1)	

Data obtained from this study represented the Indian population data. *The German population data was obtained from Frank *et al.* 2007. BC, breast cancer; wt/wt, homozygous wild-type genotype; wt/Del, heterozygous genotype; Del/Del, homozygous deletion genotype; OR, odds ratio (Del/Del + wt/Del) versus (wt/wt); CI, confidence interval; P, significance values

The association of the *MTUS1* deletion variant was found to be significant ($P=0.0207$) in case of BC while for HNC non-significant association was highlighted in Indian population. However, no conclusive inference can be drawn with the small sample size and hence, validation in a large sample size of cancer patients is warranted.

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Authors report no conflict of interests.

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REFERENCES

- Redon R, Ishikawa S, Fitch KR, Feuk L, Perry GH, Andrews TD, *et al.* Global variation in copy number in the human genome. *Nature* 2006;444:444-54.
- Perry GH, Ben-Dor A, Tsalenko A, Sampas N, Rodriguez-Revenga L, Tran CW, *et al.* The fine-scale and complex architecture of human copy-number variation. *Am J Hum Genet* 2008;82:685-95.
- Sebat J, Lakshmi B, Troge J, Alexander J, Young J, Lundin P, *et al.* Large-scale copy number polymorphism in the human genome. *Science* 2004;305:525-8.
- Sharp AJ, Locke DP, McGrath SD, Cheng Z, Bailey JA, Vallente RU, *et al.* Segmental duplications and copy-number variation in the human genome. *Am J Hum Genet* 2005;77:78-88.
- Feuk L, Carson AR, Scherer SW. Structural variation in the human genome. *Nat Rev Genet* 2006;7:85-97.
- Stankiewicz P, Lupski JR. Structural variation in the human genome and its role in disease. *Annu Rev Med* 2010;61:437-55.
- Zhang F, Gu W, Hurles ME, Lupski JR. Copy number variation in human health, disease, and evolution. *Annu Rev Genomics Hum Genet* 2009;10:451-81.
- Dhawan D, Padh H. Pharmacogenetics: technologies to detect copy number variations. *Curr Opin Mol Ther* 2009;11:670-80.
- Hinds DA, Kloek AP, Jen M, Chen X, Frazer KA. Common deletions and SNPs are in linkage disequilibrium in the human genome. *Nat Genet* 2006;38:82-5.
- Frank B, Bermejo JL, Hemminki K, Sutter C, Wappenschmidt B, Meindl A, *et al.* Copy number variant in the candidate tumor suppressor gene *MTUS1* and familial breast cancer risk. *Carcinogenesis* 2007;28:1442-5.
- Lupski JR, Stankiewicz P. Genomic disorders: molecular mechanisms for rearrangements and conveyed phenotypes. *PLoS Genet* 2005;1:e49.
- Shlien A, Malkin D. Copy number variations and cancer. *Genome Med* 2009;1:62.
- Seibold S, Rudroff C, Weber M, Galle J, Wanner C, Marx M, *et al.* Identification of a new tumor suppressor gene located at chromosome 8p21.3-22. *FASEB J* 2003;17:1180-2.
- Pineau P, Nagai H, Prigent S, Wei Y, Gyapay G, Weissenbach J, *et al.* Identification of three distinct regions of allelic deletions on the short arm of chromosome 8 in hepatocellular carcinoma. *Oncogene* 1999;18:3127-34.
- Pils D, Horak P, Gleiss A, Sax C, Fabjani G, Moebus VJ, *et al.* Five genes from chromosomal band 8p22 are significantly down-regulated in ovarian carcinoma: N33 and EFA6R have a potential impact on overall survival. *Cancer* 2005;104:2417-29.
- Chaib H, MacDonald JW, Vessella RL, Washburn JG, Quinn JE, Odman A, *et al.* Haploinsufficiency and reduced expression of genes localized to the 8p chromosomal region in human prostate tumors. *Genes Chromosomes Cancer* 2003;37:306-13.
- Yokota T, Yoshimoto M, Akiyama F, Sakamoto G, Kasumi F,

- Nakamura Y, *et al.* Localization of a tumor suppressor gene associated with the progression of human breast carcinoma within a 1-cM interval of 8p22-p23.1. *Cancer* 1999;85:447-52.
18. Kerangueven F, Noguchi T, Coulier F, Allione F, Wargniez V, Simony-Lafontaine J, *et al.* Genome-wide search for loss of heterozygosity shows extensive genetic diversity of human breast carcinomas. *Cancer Res* 1997;57:5469-74.
 19. Nouet S, Amzallag N, Li JM, Louis S, Seitz I, Cui TX, *et al.* Trans-inactivation of receptor tyrosine kinases by novel angiotensin II AT2 receptor-interacting protein, ATIP. *J Biol Chem* 2004;279:28989-97.
 20. Nouet S, Nahmias C. Signal transduction from the angiotensin II AT2 receptor. *Trends Endocrinol Metab* 2000;11:1-6.
 21. Di Benedetto M, Bièche I, Deshayes F, Vacher S, Nouet S, Collura V, *et al.* Structural organization and expression of human *MTUS1*, a candidate 8p22 tumor suppressor gene encoding a family of angiotensin II AT2 receptor-interacting proteins, ATIP. *Gene* 2006;380:127-36.
 22. Deshayes F, Nahmias, C. Angiotensin receptors: a new role in cancer? *Trends Endocrinol Metab* 2005;16:293-9.
 23. Wruck CJ, Funke-Kaiser H, Pufe T, Kusserow H, Menk M, Schefe JH, *et al.* Regulation of transport of the angiotensin AT2 receptor by a novel membrane-associated Golgi protein. *Arterioscler Thromb Vasc Biol* 2005;25:57-64.
 24. Greco S, Muscella A, Elia MG, Salvatore P, Storelli C, Marsigliante S. Activation of angiotensin II type I receptor promotes protein kinase C translocation and cell proliferation in human cultured breast epithelial cells. *J Endocrinol* 2002;174:205-14.
 25. Sambrook J, Fritsch EF, Maniatis T. *Molecular cloning: A laboratory manual*. New York: Cold Spring Harbor Laboratory Press; 1989.
 26. Almal SH, Padh H. Implications of gene copy-number variation in health and diseases. *J Hum Genet* 2012;57:6-13.
 27. Di Benedetto M, Pineau P, Nouet S, Berhouet S, Seitz I, Louis S. *et al.* Mutation analysis of the 8p22 candidate tumor suppressor gene *ATIP/MTUS1* in hepatocellular carcinoma. *Mol Cell Endocrinol* 2006;252:207-15.
 28. Macias MJ, Wiesner S, Sudol M. WW and SH3 domains, two different scaffolds to recognize proline-rich ligands. *FEBS Lett* 2002;513:30-7.
 29. Tchatchou S, Burwinkel B. Chromosome copy number variation and breast cancer risk. *Cytogenet Genome Res* 2008;123:183-7.
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