# A Quantitative Analysis of N-phenyl-6,11-dihydrodibenzo[b,e]oxepin-11-carboxamides and Related Derivatives as the Inhibitors of Acyl-CoA Cholesterol Acyl Transferase (ACAT)

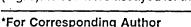
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The ACAT inhibitory activity, -log IC<sub>50</sub> (molar), of N-phenyl-6,11-dihydrodibenzo[b,e]oxepin-11-carboxamides and related derivatives, has been quantitatively correlated with structural and electronic substituent parameters; the sum of molar refraction,  $\Sigma$ MR for the R<sub>1</sub>, the Kier's  $\Delta$  parameter and a hydrogen bond count parameter, HB for 2-R<sub>2</sub> and the sum of  $\Delta$  parameters,  $\Sigma\Delta$ ' for the 3-R<sub>2</sub>, R<sub>3</sub>, X, and Y-Z. The analysis has revealed that for good ACAT inhibitory activity, the substituent R<sub>1</sub> should have a large MR value while 2-R<sub>2</sub> should have a higher  $\Delta$  but a lower HB. The remaining variations at 3-R<sub>2</sub>, R<sub>3</sub>, X, and Y-Z parts should be such that these give a lower  $\Sigma\Delta$ ' values.

HE enzyme, acyl-CoA cholesterol acyl transferase (ACAT, EC 2.3.1.26), is mainly responsible for the intracellular esterification of cholesterol and lipoprotein secretion by the liver<sup>1-5</sup>. It is believed to play an important role in the absorption of dietary cholesterol from the intestine, the metabolism of cholesterol in liver, and the accumulation of cholesteryl esters in arterial lesions<sup>5,6</sup>. Reportedly, ACAT activity is enhanced in the intestinal mucosal cells when cholesterol is ingested<sup>7</sup> and in arterial cells undergoing atherosclerosis8. Inhibition of arterial macrophage ACAT enzyme can directly intervene in the progression of lesions. Therefore, it would be beneficial to reduce plasma cholesterol concentration, to reduce the secretion of very low density lipoproteins (VLDL) into the plasma, and to prevent the formation of foam cells in the arterial walls. In view of this, the ACAT inhibitors provide potential as hypocholesterolemic and antiatherosclerotic agents9,10.

During the course of their search for potent ACAT inhibitors of novel structures, Kumazawa et al.<sup>11</sup> have reported a series of N-phenyl-6,11-dihydrodibenzo [b,e]oxepin-11-carboxamides and related analogues (Fig. 1). These were assayed for their ability to inhibit ACAT



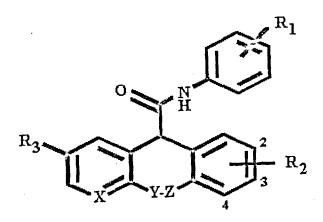


Fig. 1: Structures of N-phenyl-6,11-dihydrodibenzo[b,e] oxepin-11-carboxamides and related derivatives

in vitro and to decrease serum total cholesterol in vivo. The compounds, their in vitro inhibitory actions, as determined by incubation with [14C]oleolyl-CoA and liver microsomes from cholesterol-fed rabbits, are listed in Table 1.

The title compounds (Fig. 1) have substituents at five different sites, viz.,  $R_1$ ,  $R_2$ ,  $R_3$ , X, and Y-Z. Further, their activities, -log IC<sub>50</sub>'s (molar basis), show considerable variation, with compound 27 behaving as the most potent

member of the series. We were, therefore, temped to derive quantitative structure-activity correlations for this series of compounds using multiple regression analysis (MRA) to obtain the best-fit of biological data with certain substituent parameters, described herein. It is intended to throw some light on the physicochemical basis of their biological actions. The various substituent parameters are also given in Table 1.

# **Parameterization**

Amongst the large number of alternatives, those consisdered the most appropriate predictor parameters for explaining the observed inhibitory actions of the title compounds, are the sum of the molar refractions  $^{12,13}$   $\Sigma$ MR for the R<sub>1</sub> substituents, and the Kier's  $\Delta$  parameter  $^{14,15}$  and a hydrogen bond count parameter  $^{16}$ , HB, for the R<sub>2</sub>'s. The MR reports on the bulk of the substituents. However, it is also known that MR parameter measures the electronic effects and may also reflect dipole-dipole interaction  $^{13}$  at the active site. The Kier's  $\Delta$  parameter is given by Eq. (1).

$$\Delta = (\delta^{v} - \delta) / N^{2}$$
 (1)

Where  $\delta^{v}$  and  $\delta$  are the valence and simple deltas, respectively, and are the counts of electrons in  $\sigma$ , $\pi$  or lonepair orbitals. N is the principal quantum number (e.g., 1, 2, 3, ..., etc). Kier and Hall15 have obtained a highly significant linear correlation between  $\Delta$  and the Mulliken's electronegativities for 19 atoms (considering these in different hybridisation states) of the first, second, and third rows of periodic table. Therefore,  $\Delta$  parameter is believed to report upon the electronic and other related effects. Since very few variations occur at 3-R2, R3 X, and Y-Z, the sum of their Kier's  $\Delta$  values,  $\Sigma\Delta$ ' instead of an indicator or any other parameter, was used for deriving the follow up correlations. Lastly, the HB parameter is based on a consideration of the number of atoms in a group that can be involved in the hydrogen-bonding interaction, either as a H-bond donor or H-bond acceptor<sup>16</sup>.

## RESULTS AND DISCUSSION

MRA on the data of Table 1 returned the correlation as given by Eq. (2)

-log IC<sub>50</sub> =  $0.412(\pm 0.056)$ ΣMR +1.267(±0.268)Δ -0.335 (±0.074)HB -1.104(±0.284) ΣΔ'+6.116 n=43, s=0.268, R=0.840, EV=0.676, F<sub>4.38</sub>=22,875 (2).

Where n is the data-size, s, the standard error of estimates, R, the multiple correlation cofficient, EV, the explained variance in the calculated activity data, and F, the F-ratio between the variances of calculated to observed activities. The ± data are the 95% confidence interval associated with the individual regression coefficients. SMR accounts for the bulk at R, in the phenyl parts. No other attempted parameters for this position could replace  $\Sigma$ MR. However, the van der Waals volume, V,, also a measure of size, was found to be perfectly correlated with  $\Sigma$  MR  $(r=0.994; s=0.026; F_{1.41} = 3223.7)$  and its use in place of MR gave a comparable EV and R but a higher ± data then the ones in Eq. (2). The variables  $\Delta$  and HB stand to account respectively for electronegativity based property and Hbonding (both donor and acceptor) property of the 2-substituents. This position seems to be of critical and decisive importance and demands two separate predictors to distinguish between the types of R2's at this position, specially the halogens. The R2 substituents having a sulphur atom or one bigger than it, were arbitrarily selected for  $\Delta$ calculations (Eq. 1) while those being smaller in size but more electronegative than sulphur were retained for the HB count parameter. For example, for R<sub>2</sub>=2-F and 2-Cl (compound numbers 20 and 21, the substituents being smaller and more electronegative than sulphur) the HB parameter is important while for  $R_2 = 2$ -Br and 2-I (compound numbers 22 and 24, which are bigger and less electronegative than sulphur) the  $\Delta$  parameter is very necessary. Further, the value of Kier's parameter are the function of the principal quantum number, N (Eq. 1). therefore, these values of  $\Delta$  are also dependent on the type of halogen. Besides the halogen at 2-R2, the other substituents such as  $R_2 = 2 - CF_3$ , 2-OMe and 2-CN (compound numbers 25, 26 and 28) and R<sub>2</sub> = 2-COOMe, 2-CONMe,, 2-CH,OH and 2-NO, (compound numbers 29-32) are considered for HB parameter, according to the method suggested by other workers<sup>16</sup>. The electronic σ. field F, resonance R, or the hydrophobicity  $\pi$ , were all tried but to no avail. Finally, for the 3-R2, R3, X and the Y-Z positions also,  $\Sigma\Delta'$  was found convincing.

Though Eq. (2) is sound statistically as is evinced from a low, s, a reasonably good R and EV as well as a F-value that is significant at 99% level  $[F_{4,38}(0.01)=3.86]$ , there is still need for its betterment. Using this equation, the calculated activities (omitted here) for compounds 7, 14, 16 and 28, were found to deviate much from the observed ones. Also, these compounds are not active (see Table 1).

Table 1: Predictor Variables and Cholesterol Acyltransferase Inhibition Actions of N-Phenyl-6, 11-dihydrodibenzo[b,e]oxepin-11-carboxamides and Related Derivatives (Fig. 1 for structures)

S. No.	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>	Х	Y-Z	ΣMRª	Δ	НВ	ΣΔ'	-log IC <sub>5</sub>	<sub>o</sub> (M) Calcd°
1	2,4-F <sub>2</sub>	2-Br	Н	СН	CH <sub>2</sub> O	0.29	0.38	0	0.25	6.24	6.54
2	2,6-Cl <sub>2</sub>	2-Me	Н	СН	CH₂O	1.31	0	0	0.25	6.72	6.45
3	2,6-Cl <sub>2</sub>	2-Br	Н	СН	CH₂O	1.31	0.38	0	0.25	7.13	6.95
4	2,6-Cl <sub>2</sub>	3-Br	Н	СН	CH₂O	1.31	0	0	0.63	6.03	5.97
5	2,6-Cl <sub>2</sub>	2-¹Bu	Н	СН	CH <sub>2</sub> O	1.31	0	0	0.25	6.38	6.45
6	2,6-Br <sub>2</sub>	2-Me	Н	СН	CH₂O	1.88	0	0	0.25	6.72	6.68
7 .	2,4,6-OMe <sub>3</sub>	2-Me	Н	СН	CH₂O	2.36	0	0	0.25	6.24 <sup>d</sup>	
8	2,6-Me <sub>2</sub>	2-Me	Н	СН	CH <sub>2</sub> O	1.23	0	0	0.25	6.46	6.42
9	2,6-Me <sub>2</sub>	2- <sup>i</sup> Pr	Н	СН	CH <sub>2</sub> O	1.23	0	0	0.25	6.44	6.42
10	2,6-Me <sub>2</sub>	2-¹Bu	Н	СН	CH₂O	1.23	0	0	0.25	6.30	6 42
11	2,4,6-Me <sub>3</sub>	2-Me	Н	СН	CH₂O	1.70	0	0	0.25	6.96	6.61
12	2,6-Et <sub>2</sub>	2-Me	Н	СН	CH <sub>2</sub> O	2.16	0	0	0.25	6.57	6.80
13	2,6-Et <sub>2</sub>	2- <sup>i</sup> Pr	Н	СН	CH₂O	2.16	0	0	0.25	6.30	6.80
14	2-Cl,6-Me	2-Br	Н	СН	CH₂O	2.16	0.38	0	0.25	6.60 <sup>d</sup>	
15	2-iPr	2 <b>-</b> Me	Н	СН	CH₂O	1.27	0	0	0.25	6.59	6.43
16	2,6-Pr <sub>2</sub>	2-Me	Н	СН	CH₂O	1.70	0	0	0.25	6.14 <sup>d</sup>	_
17	2,6- <sup>i</sup> Pr <sub>2</sub>	н	Н	СН	CH <sub>2</sub> O	3.10	0	0	0.25	7.03	7.17
18	2,6- <sup>1</sup> Pr <sub>2</sub>	2-Me	Н	СН	CH₂O	3.10	0	0	0.25	7.30	7.17
19	2,6- <sup>i</sup> Pr <sub>2</sub>	2-Et	Н	CH	CH <sub>2</sub> O	3.10	0	0	0.25	6.89	7.17
20	2,6- <sup>i</sup> Pr <sub>2</sub>	2-F	Н	СН	CH₂O	3.10	0	1	0.25	6.68	6.83
21	2,6- <sup>i</sup> Pr <sub>2</sub>	2-Cl	Н	CH	CH₂O	3.10	0	1	0.25	6.82	6.83
22	2,6- <sup>i</sup> Pr <sub>2</sub>	2-Br	Н	CH	CH₂O	3.10	0.38	0	0.25	7.64	7.68
23	2,6- <sup>1</sup> Pr <sub>2</sub>	3-Br	Н	СН	CH₂O	3.10	0	0	0.63	6.74	6.70
24	2,6- <sup>i</sup> Pr <sub>2</sub>	2-1	Н	СН	CH₂O	3.10	0.24	0	0.26	7.46	7.50
25	2,6- <sup>i</sup> Pr <sub>2</sub>	2-CF <sub>3</sub>	Н	СН	CH₂O	3.10	0	1	0.25	6.80	6.83
26	2,6- <sup>i</sup> Pr <sub>2</sub>	2-OMe	Н	СН	CH₂O	3.10	0	1	0.25	6.85	6.83
27	2,6- <sup>i</sup> Pr <sub>2</sub>	2-SMe	Н	СН	CH <sub>2</sub> O	3.10	0.44	0	0.25	8.15	7.91
28	2,6- <sup>i</sup> Pr <sub>2</sub>	2-CN	Н	СН	CH₂O	3.10	0	1	0.25	6.31 <sup>d</sup>	
29	2,6- <sup>i</sup> Pr <sub>2</sub>	2-COOMe	Н	СН	CH₂O	3.10	0	2	0.25	6.74	6.48
30	2,6- <sup>i</sup> Pr <sub>2</sub>	2-CONMe2	Н	СН	CH₂O	3.10	0	2	0.25	6.40	6.48
31	2,6-Pr <sub>2</sub>	2-CH <sub>2</sub> OH	Н	СН	CH₂O	3.10	0	2	0.25	6.24	6.48
32	2,6- <sup>i</sup> Pr <sub>2</sub>	2-NO <sub>2</sub>	Н	СН	CH₂O	3.10	0	2	0.25	6.62	6.48

S. No.	•	R <sub>2</sub>	R <sub>3</sub>	Х	Y-Z	ΣMRª	Δ	НВ	ΣΔ'	-log lC₅ Obsd⁵	₀(M) Calcd°
33	2,6-'Pr <sub>2</sub>	2,3-Me <sub>2</sub>	Н	СН	CH₂O	3.10	0	0	0.25	7.19	7.18
34	2,6-'Pr <sub>2</sub>	2-Me,4-COOM	leH	СН	CH₂O	3.10	0	0	0.25	7.57	7.18
35	2,6-Pr <sub>2</sub>	2-Me,4-Br	Н	СН	CH <sub>2</sub> O	3.10	0	0	0.25	7.00	7.18
36	2,6- <sup>1</sup> Pr <sub>2</sub>	Н	Br	СН	CH₂O	3.10	0	0	0.63	6.72	6.70
37	2,6-Pr <sub>2</sub>	2-Me	Br	СН	CH <sub>2</sub> O	3.10	0	0	0.63	6.92	6.70
38	2,6-'Pr <sub>2</sub>	2-Br	Br	СН	CH <sub>2</sub> O	3.10	0.38	0	0.63	7.24	7.20
39	2,6- <sup>i</sup> Pr <sub>2</sub>	Н	Н .	N	CH₂O	3.10	0	0	0.75	6.19	6.54
40	2,6-'Pr <sub>2</sub>	2-Br	Н	N	CH <sub>2</sub> O	3.10	0.38	0	0.75	7.11	7.04
41	2,6-Pr <sub>2</sub>	Н	Н	СН	CH <sub>2</sub> CH <sub>2</sub>	3.10	0	0	0.25	7.40	7.18
42	2,6-'Pr <sub>2</sub>	2-Me	Н	СН	CH <sub>2</sub> CH <sub>2</sub>	3.10	0	0	0.25	7.26	7.18
43	2,6- <sup>1</sup> Pr <sub>2</sub>	2-Br	Н	СН	CH <sub>2</sub> CH <sub>2</sub>	3.10	0.38	0	0.25	7.41	7.68

<sup>\*</sup>The MR values are scaled to 0.1; the IC<sub>50</sub> data are from ref.11; calculated using Eq. (3), and the outlier compounds, not included in deriving Eq. (3).

However, no specific reason is immediately becoming apparent for their 'outlier' behaviour. Thus the MRA, ignoring these four data points, provided a much improved correlation shown in Eq. (3).

-log  $IC_{50} = 0.407(\pm 0.044)\Sigma MR + 1.343(\pm 0.222)$   $\Delta$ -0.348(±0.059)HB-1.270(±0.226) $\Sigma \Delta$ '+6.234 n=39, s=0.211, R=0.903, EV=0.793, F<sub>4.34</sub>=37.474 (3)

As can be seen from Eq. (3), the R, EV and F-values  $[F_{4,34} (0.01)=3.93]$  have been sharply raised while the ± data have been lowered. The EV value now accounts for 79% variance in the calculated log  $IC_{50}$  values. Furthermore, the predictor variables of this equation are mutually orthogonal as shown by the intercorrelation matrix given in Table 2. This equation reveals that for a compound to be an effective inhibitor of ACAT, both the  $\Sigma$ MR (for R, substitutions) and the  $\Delta$  value (for 2-R, substitution) should be large positive, the later should preferably be >0.5, while both the HB and  $\Sigma\Delta$  should be smaller. This is in agreement with the observation of Kumazawa et al.11 that at 2- and 6-R<sub>1</sub>, potency is higher for bulky group (hence, a more MR value) while substitution at R<sub>3</sub> (hence, a larger ΣΔ') leads to reduced potency. They had further observed that 3-bromo analogues were less potent than 2-bromo analogues (thus, demanding a lower  $\Sigma\Delta$  at 3-R<sub>2</sub>). For the 2-halo derivatives,

Table 2: The Intercorrelation Matrix Among the Predictor Variable of Eq. (3)

	ΣMR	Δ	НВ	ΣΔ'
$\Sigma$ MR	1.000	0.004	0.318	0.181
Δ		1.000	0.236	0.106
НВ			1.000	0.220
ΣΔ'	<del></del>			1.000

Values shown above are the correlation coefficients between predictor variables.

potency increase in the order F, CI, I, Br, while for the alkyl substituents, methyl analogues are more active than the ethyl or isopropyl analogues. Compounds 22, 24, 27 and 34, which all fulfil these criteria, are in fact, reported to more active than many other of Table 1. Using Eq. (3) the calculated -log  $IC_{50}$  values listed in Table 1 are, in most cases, close to the observed ones. Therefore, this equation may be applied in computing the theoretical activity of a potential ACAT inhibitor before its actual synthesis and bioassay. These inferences may, therefore, form the basis for a more rational substituent-selection in future drugdesign strategy.

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## REFERENCES

- Cianflone, K.M., Yasruel, Z., Rodriguez, M.A., Vas, D. and Sniderman, A.D., J. Lipid Res., 1990, 31, 2045.
- 2. Dashti, N., J. Biol. Chem. 1992, 267, 7160.
- 3. Carr, T.P., Parks J.S. and Rudel, L.L., Arterioscler. Thromb., 1992, 12, 1274.
- 4. Tanaka, M., Jingami, H., Otani, H., Cho, M., Ueda, Y., Arai, H., Nagano, Y., Doi, T., Yokode, M. and Kita, T., J. Biol. Chem., 1993, 268, 12713.
- Spector, A.A., Mathur, S. N. and Kaduce, T.L., Prog. Lipid. Res. 1979, 18, 31.
- 5b. Suckling, K.E. and Stange, E.F., **J. Lipid Res.**, 1985, 26, 647.
- 6a. Field, F.J., Cooper, A.D. and Erickson, S.K., Gastroenterology, 1982, 83, 873.
- 6b. Norum, K.R., Helgerud, P., Peterson, L.B., Groot, P.H.E. and de Jonge, H.R., **Biochim. Biophys. Acta,** 1983, 751, 153.

- 7. Bell, F.P. in: Pharmacological Control of Hyperlipidaemia, J.R. Prous Science, Barcelona, Spain, 1986, 409.
- 8. Sliskovic, D.R. and White, A.D., Trends Pharmacol. Sci., 1991, 12, 194.
- 9. Trivedi, B.K., Homes, A., Stoeber, T.L., Blankley, C.J., Roark, W.H., Pickard, J.A., Shaw, M.K., Essenburg, A.D., Stanfield, R.L., and Krause, B.R., J. Med. Chem., 1993, 36, 3300.
- Trivedi, B.K., Purchase, T.S., Holmes, A., Augelli-Szafran, C.E., Essenburg, A.D., Hamelehle, K.L., Stanfield, R.L., Bousley, R.F. and Krause, B.R., J. Med. Chem., 1994, 37, 1652.
- 11. Kumazawa, T., Yanase, M., Harakawa, H., Obase, H., Shirakura, S., Ohishi, E., Oda, S., Kubo, K. and Yamada, K., J. Med. Chem., 1994, 37, 804.
- Hansch, C. and Leo, A.J. in: Substituent Constants for correlation analysis in chemistry and biology, Jonn Wiley, New York, 1979, 48.
- Dunn, W.J., III Eur. J. Med. Chem. Chim. Ther., 1977, 12, 109.
- Kier, L.B. and Hall, L. H. in: Molecular connectivity in chemistry and drug research, Academic Press, New York, 1976, 35.
- 15. Kier, L.B. and Hall, L.H., J. Pharm. Sci., 1981, 70, 583.
- 16. Yang, G., Lien, E.J. and Guo, Z., Quant. Stru-Act. Relat., 1986, 5, 12.