

# Assessment of *In Vitro* Antioxidant, Antibacterial, Anticancer, Anti-Elastase and Anti-Collagenase Activity of *Hypnea pannosa* Extracts for Unveiling its Potential in Cosmeceuticals

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Baskar *et al.*: Assessment of Bioactive Properties of *Hypnea pannosa* extracts and its Implication on Cosmeceuticals

The present study compared aqueous and ethanolic solvents to evaluate the *in vitro* anticancer, anti-elastase and anti-collagenase activity of *Hypnea pannosa* seaweed extracts. Additionally, the study evaluated the extract's preliminary phytochemical composition, total phenolic and flavonoid content, antioxidant and antibacterial activity. The analysis revealed the presence of several phytochemicals and the range of total phenolic ( $11.14 \pm 0.17$  mg gallic acid equivalents/g and flavonoid ( $5.83 \pm 0.09$  mg quercetin equivalent/g) content in the ethanolic extract. The ethanolic extract showed the highest antioxidant activity against 2,2-diphenyl-1-picrylhydrazyl (half-maximal inhibitory concentration =  $6.32 \pm 0.04$   $\mu$ g/ml) and the most potent anticancer effect against COLO 829 melanoma cancer cell lines (half-maximal inhibitory concentration =  $95.70$   $\mu$ g/ml). It also exhibited the most potent anti-elastase and anti-collagenase effect against human melanoma cells, with half-maximal inhibitory concentration values of  $48.53$  and  $48.76$   $\mu$ g/ml, respectively. The ethanol extract showed maximum antibacterial activity and the highest inhibition zone of  $14.5$  and  $14$  mm against *Escherichia coli* and *Pseudomonas aeruginosa*, respectively. Fourier transform infrared spectroscopic analysis shows that many polyphenols are present in ethanol extract, which has received importance in cosmeceutical perspectives ( $3925$ – $3893$   $\text{cm}^{-1}$ ). The study hence provides evidence that the ethanol extract of *Hypnea pannosa* has significant potential for antioxidants, anticancer, antibacterial, anti-elastase and anti-collagenase and may lead to further cosmeceutical applications through the isolation and purification of bioactive compounds.

**Key words:** Antioxidant, antibacterial, anticancer, anti-elastase, anti-collagenase, 2,2-diphenyl-1-picrylhydrazyl, phenols, flavonoids, Fourier transform infrared spectroscopy

Skin ageing is a complex process influenced by various factors, both intrinsic and extrinsic. Intrinsic factors are related to the passage of time, while extrinsic factors mainly arise from exposure to the sun<sup>[1]</sup>. When exposed to Ultraviolet (UV) radiation, the skin changes complex pathways, eventually releasing Reactive Oxygen Species (ROS), Matrix Metalloproteinases (MMPs) and elastase<sup>[2]</sup>. ROS is responsible for skin aging by causing oxidative damage to the skin's lipids, proteins and DNA<sup>[3,4]</sup> and consequently induces severe human diseases, including chronic inflammation, atherosclerosis, cancer, cardiovascular disorders and aging<sup>[5]</sup>. Fortunately, recent developments in marine biotechnology have opened up new avenues for research in skin aging,

inflammation and degradation. Marine bioactive compounds can positively impact the treatment of skin disorders<sup>[6]</sup>. Marine algae, also known as seaweed has become an essential component of the cosmeceutical industry, accounting for 40 % of the world's hydrocolloid market<sup>[7]</sup>. Seaweed has become increasingly popular due to its bioactive properties, such as potent antioxidant, antibacterial, antiviral, antifungal, anti-inflammatory and anticancer agent<sup>[8]</sup>.

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Red seaweed is being considered by the cosmetic industry as a source of bio-sustainable components for skin care due to its high concentration of biologically active compounds like agar and carrageenan, which can be used as effective skin care agents<sup>[9]</sup>. *Hypnea* has recently grown significantly among red algae due to its biological properties, such as anticancer, antiaging and antioxidant activities<sup>[10]</sup>. Algal extracts have also been utilized in dietary supplements, cosmetics, and alternative medicines recommended for skin-related diseases<sup>[11]</sup>. Plant extract such as *Hypnea pannosa* (*H. pannosa*) has discovered their ability to scavenge free radicals and exhibit anti-collagenase and anti-elastase activities. The extract from *Hypnea* positively impacts the skin as it aids in repairing and calming any damage caused by exposure to UV rays or sunburn<sup>[12]</sup>. Studies have shown that marine algae have properties that protect against cancer cells. Different types of algae have been found to have cytotoxic and antitumor activities<sup>[13,14]</sup>. UV B radiation is also the main factor in initiating and developing melanoma and non-melanoma skin cancers<sup>[15]</sup>. Polyphenolic compounds extracted from different algal species have many biological activities, including their role in inhibiting melanogenesis<sup>[16-18]</sup>, elastase and collagenase enzymes<sup>[19]</sup> are essential in cosmeceutical product development. Collagenase is crucial in remodelling the Extracellular Matrix (ECM), especially in breaking down collagen. Elastase, on the other hand is a serine proteinase that is responsible for breaking down elastin in the ECM. The depletion of collagen and elastin in the skin can lead to undesired wrinkles and aging skin, as they are essential for maintaining the structural integrity and elasticity of the skin<sup>[20]</sup>. Currently, researchers are focusing on discovering phytochemicals, such as phenols and flavonoids, from marine algae, which have potent antioxidant effects. Seaweed has been identified as an essential source of these phytochemicals. The present study examined and compared the total phenolic and flavonoid content of the aqueous and ethanolic *H. pannosa* extract while determining its *in vitro* antioxidant, antibacterial, anticancer, anti-collagenase and anti-elastase properties. The study also examined the presence of functional groups of compounds by the FT-IR spectroscopic method. The results suggest that crude *H. pannosa* extracts possessing high bioactivity could be

used as natural anti-aging ingredients, which can be incorporated into cosmetic products.

## MATERIALS AND METHODS

### Plant material:

The red seaweed, *H. pannosa* was collected along the coast of Mandapam and washed thoroughly to remove all traces of sand and epiphytic organisms. The dried seaweed was then ground to a fine powder using a pulverizer equipped with a 3 mm perforated grill (Centertech, Madurai) and stored in the refrigerator until use.

### Extraction:

*H. pannosa* was subjected to extraction using two solvents viz., ethanol and water, following the procedure outlined by Cho *et al.*<sup>[21]</sup>. Initially, 1 g of a powdered sample was added to 10 ml of ethanol and agitated using a magnetic stirrer at 720 revolutions per minute (rpm) at ambient temperature for 24 h. The sample was centrifuged at 2090 rpm for 10 min and collected. The extraction procedure was repeated twice to obtain the residue, which was subsequently combined with the prior extract. *H. pannosa* was extracted using water as a solvent in a similar manner. The mixture was then filtered through Whatman filter paper (125 mm) and concentrated using a rotary evaporator. The concentrate was then dried at 37° and stored refrigerated until use.

### Preliminary phytochemical screening:

All freshly prepared crude extracts of seaweed were evaluated qualitatively for the identification of different classes of active phytochemical constituents, such as terpenoids<sup>[22]</sup>, tannin<sup>[23]</sup>, saponin<sup>[24]</sup> and alkaloids<sup>[25]</sup> using standard techniques. The presence or absence of these compounds in the crude extracts tested was revealed during these general reactions.

### Test for terpenoids (Salkowski test):

5 ml of each extract was added with 2 ml chloroform and 3 ml of concentrated sulphuric acid (H<sub>2</sub>SO<sub>4</sub>). The formation of a reddish-brown colour at the interface indicates the presence of terpenoids in the sample extracts<sup>[22]</sup>.

### Test for tannins:

3 ml of extract was mixed with a few drops of

1 % Hydrochloric acid (HCl) and observed for precipitation. A red precipitate showed positive results for tannins in the crude extract<sup>[23]</sup>.

#### Test for saponin (frothing test):

About 3 ml of each extract was treated with 3 ml of distilled water and the sample was shaken vigorously for a stable, persistent froth for about 1 min. The frothing was added with 3 drops of olive oil and shaken vigorously, then observed for emulsion formation, indicating the presence of saponins in the extracts<sup>[24]</sup>.

#### Test for alkaloids:

By adding 1 ml of Dragendorff's reagent to 2 ml of extracts, an orange-red precipitate was formed, indicating the presence of alkaloids<sup>[25]</sup>.

#### Total phenolic content:

The total phenolic content determined in seaweed extract was carried out according to the method of Slinkard *et al.*<sup>[26]</sup>. 0.5 ml of sample extract was mixed with 0.5 ml of Folin-Ciocalteu (5 ml of Folin-Ciocalteu reagent diluted with 50 ml of distilled water). After 5 min, 2.5 ml of Sodium carbonate ( $\text{Na}_2\text{CO}_3$ ) (2 %) was added. After 40 min, the absorbance was measured at 725 nm using a double-beam spectrophotometer (Model UV-1800, Shimadzu, Kyoto, Japan). The total phenolic content was determined using a calibration curve derived from gallic acid and the results were expressed as mg of Gallic Acid Equivalents (GAE)/g of crude extract.

#### Total flavonoid content:

The determination of flavonoid content in seaweed extracts was analysed according to Kumar *et al.*<sup>[27]</sup>. 4 ml of distilled water, 0.3 ml of 10 % aluminum chloride ( $\text{AlCl}_3$ ) and 0.3 ml of 5 %  $\text{NaNO}_2$  were added to 0.5 ml of crude extract. After 6 min, 2.5 ml of distilled water and 2 ml of 1 N NaOH were added to the mixture. At 506 nm, the absorbance was then determined. Total flavonoid concentration in the extracts was calculated using a calibration curve using quercetin and the results were expressed as mg Quercetin (QE)/g of crude extract.

#### *In vitro* antioxidant activity using 2,2-Diphenyl-1-Picrylhydrazyl (DPPH):

The antioxidant properties were determined by DPPH scavenging potential, in which ascorbic

acid was used as a positive control, according to Blois *et al.*<sup>[28]</sup>. A 2 ml aliquot of 0.16 mM DPPH solution in methanol was mixed with aliquots of extract (2 ml) and then incubated in the dark at room temperature for 30 min. The same procedure was followed for positive standard containing ascorbic acid. At 517 nm, the absorbance of the samples and standards was measured.

$$\text{DPPH radical scavenging activity (\%)} = \frac{\text{Abs}_{\text{control}} - \text{Abs}_{\text{sample}}}{\text{Abs}_{\text{control}}} \times 100$$

#### *In vitro* antibacterial activity:

The disc diffusion method performed the *in vitro* antibacterial activity of the *H. pannosa* extracts. The test bacteria were inoculated in peptone water and incubated for 3-4 h at 35°. Mueller Hinton agar was prepared and poured into sterile petri plates. 0.1 ml of bacterial culture was inoculated on the Mueller Hinton agar plates' surface and spread using L-rod. The inoculated plates were allowed to dry for 5 min. The disk loaded with a sample concentration of 1000 µg/ml was placed on the surface of inoculated petri plates using a sterile technique. The plate was incubated at 37° for 18-24 h. The plate was then examined for the inhibitory zone and the zone of inhibition was measured in mm<sup>[29]</sup>.

#### *In vitro* anticancer activity:

The 3-(4, 5-dimethylthiazolyl-2)-2, 5-diphenyltetrazolium bromide (MTT) cytotoxic assay was performed on a COLO-829 malignant melanoma (human) cancer cell line to evaluate the anticancer activity of *H. pannosa* extracts. Before being treated with the *H. pannosa* extract, the cancer cells were cultured in Rosewell Park Memorial Institute media for 24 h. 100 µl of the extracts were diluted with Dimethyl Sulfoxide (DMSO) and added at three different concentrations (100 µg/ml, 200 µg/ml and 300 µg/ml), along with a negative control of DMSO. After treatment, the cells were incubated for 24 and 48 h before determining the effect of the different concentrations of the seaweed extract on the cancer cell line<sup>[30,31]</sup>.

#### *In vitro* anti-elastase activity:

*In vitro* anti-elastase activity protocol by Senior *et al.*<sup>[32]</sup> and Sallenave *et al.*<sup>[33]</sup> was adopted with the required modifications in the present assay. SK-MEL (15 000 cells/well) were seeded on 12 well plates and allowed to acclimate to the culture

conditions, such as 37° and 5 % CO<sub>2</sub> environment, in the incubator for 24 h. The test samples were prepared in Dulbecco's Modified Eagle Medium (DMEM) (100 mg/ml) and filtered and sterilized using a 0.2 µm Millipore syringe filter. The samples were diluted in DMEM media and added to the wells containing cultured cells at final concentrations of 6.25, 12.5, 25, 50, 100 and 200 µg/ml respectively. Untreated wells were kept in control. All the experiments were done in triplicate and average values were taken to minimize errors. After treatment with the test samples, the plates were further incubated for 24 h. After the incubation period, the media from the wells were aspirated and discarded. The anti-elastase activities of the extracts were finally expressed as half-maximal Inhibitory Concentration (IC<sub>50</sub>) value.

The percentage of elastase inhibition activity was calculated using the following equation;

Elastase inhibition (%) =  $(1 - \text{enzyme activity in the presence of the inhibitor} / \text{enzyme activity in absence of the inhibitor}) \times 100$

#### ***In vitro* anti-collagenase activity:**

*In vitro* anti-collagenase activity protocol by Park *et al.*<sup>[34]</sup> and Shanura *et al.*<sup>[35]</sup>, was adopted with the required modifications in the present assay. 1 mg of azo dye-impregnated collagen (A4341, Sigma) was measured in test tubes and homogenized in 800 µl of 0.1 M Tris-HCl (pH 7.0). It was then mixed respectively with 400 µl of the sample cell lysates (control-6.25, 12.5, 25, 50, 100 and 200 µg/ml). Collagenase (C0130, Sigma) prepared at a concentration of 200 units ml<sup>-1</sup> was rapidly incorporated into the reaction mixture. All the tubes were then incubated at 43° for 1 h. The contents were centrifuged at 3000 rpm for 10 min and the absorbance of the supernatant was measured at 550 nm. The anti-collagenase activities of the extracts were finally expressed as an IC<sub>50</sub> value.

The percentage of collagenase inhibition activity was calculated using the following equation;

Inhibitory rate of collagenase (%) =  $[(\text{absorbance of control} - \text{absorbance of sample}) / \text{absorbance of control}] \times 100$ .

#### **Fourier Transform-Infrared spectroscopy (FT-IR):**

FT-IR spectroscopy was carried out by using FT-IR spectrophotometer (ATR-FT-IR, Model P-4600, Thermo Scientific, USA). The dried seaweed

extracts (aqueous and ethanol) were mixed with Potassium Bromide powder (KBr) in a ratio of 1:9 and ground well. After assembling the die, the powder was added to the collar and given a hydraulic press of about 100 kg/m<sup>3</sup> to form the pellet. The pellet was taken out from the collar after disassembling the die. The spectrum was then recorded after placing the pellet on the sample holder and the signal was obtained in 32 scans at a resolution of 4 cm<sup>-1</sup> from 600-4000 cm<sup>-1</sup>.

#### **Statistical analysis:**

All data were recorded, processed and entered using Statistical Package for Social Sciences (SPSS) statistics version 20 (Chicago, IL, USA). The data obtained were expressed in average ± Standard Deviation (SD).

### **RESULTS AND DISCUSSION**

The extraction yield depended solely on the solvent polarity, pH, extraction time and sample composition<sup>[36]</sup>. The most critical stages in isolating bioactive compounds from plant material are mainly the extraction methods, significantly optimizing the most suitable solvent for the extraction from the plant material<sup>[37]</sup>. The yield of aqueous and ethanol solvent extracts of *H. pannosa* was reported as 10.26 ± 0.03 % and 8.26 ± 0.02 % respectively (Table 1). The variation in extraction yield among different solvents may be attributed to the polarities of different compounds present in the seaweeds<sup>[38,39]</sup>.

The presence or absence of phytoconstituents depends on the extraction solvent and the seaweed's physiological characteristics. The essential bioactive compounds in seaweed can be examined using standard methods involving various solvents and ambient conditions<sup>[40]</sup>. This study used ethanol and water extracts for phytochemical screening, confirming the presence of active chemical components such as terpenoids, saponin, tannin, alkaloids, phenolics, and flavonoids (Table 2). Eight out of ten tests were positive, while two were negative. However, Rafiquzzaman *et al.*<sup>[41]</sup>, identified some phytochemicals in the red algae *Hypnea musciformis* (*H. musciformis*) collected in Bangladesh. The study results indicate that ethanol extract contains more phytochemicals than aqueous extract, which has implications for cosmeceutical properties.



**TABLE 1: ANTIOXIDANT PROPERTIES OF *H. pannosa* SOLVENT EXTRACTS**

Solvent extraction	Extraction yield (%)	Total phenolic content (mg GAE/g of extract)	Total flavonoids content (mg QE/g of extract)	DPPH (IC <sub>50</sub> mg/ml)
Aqueous extract	10.26±0.03 %	8.03±0.28	3.20±0.01	9.45±0.03
Ethanol extract	8.26±0.02 %	11.14±0.17	5.83±0.09	6.32±0.04

Note: All the data were provided in mean±SD of three replicates (n=3)

**TABLE 2: PRELIMINARY PHYTOCHEMICAL SCREENING OF *H. pannosa* EXTRACTS**

Name of the extracts	Terpenoids	Saponin	Tannin	Alkaloids	Phenols	Flavonoids
Aqueous	-	+	-	+	+	+
Ethanol	+	+	+	+	+	+

Note: (-): Absence of the components and (+): Presence of the components

Polyphenolics are secondary metabolites primarily found in plants, with widespread deposition in the cell wall. These compounds possess several biological and redox properties responsible for their antioxidant capacity. The total phenolic content of the extracts was estimated using the Folin-Ciocalteu reagent method. It was found that the aqueous and ethanolic extracts had 8.03±0.28 and 11.14±0.17 mg GAE/g extract, respectively (Table 1). The study results indicate that the ethanolic extract contained more total polyphenols, identified as the primary antioxidant compounds. The solubility of phenolics increases with the polarity of the solvent, which increases the phenolic content<sup>[42]</sup>. The total phenolic content can vary based on the polarity of the solvent used for extraction, so ethanol is more effective than methanol in extracting the total phenolic content due to its higher polarity<sup>[43]</sup>. Earlier reports substantiate our present observation that solvent extracts of *Hypnea* sp. and other red seaweeds are good resources for polyphenolic compounds<sup>[44,45]</sup>. Thus, the study proved that ethanol extracts increase total phenolic content and provide good antioxidant properties.

Flavonoids are polyphenolic compounds that act as antioxidants in the body, preventing damage caused by oxidation. They have high redox potential, chelate metals, and donate hydrogen<sup>[46]</sup>. Their antioxidant capacity comes from the conjugation between flavonoid rings<sup>[47]</sup>. The total flavonoid content showed a significant difference among aqueous extract (3.20±0.01 mg QE/g) compared to ethanol extract (5.83±0.09 mg QE/g) as illustrated in Table 1. The ethanol extraction showed the highest total flavonoid content in *Gracilaria* sp.<sup>[48]</sup>. The variation in flavonoids may be due to

changes in physiochemical characteristics such as salinity. Thus, the study proved that ethanolic extracts contribute to increased flavonoid content and positively correlate with phenolic content.

DPPH is a free radical that remains stable. Antioxidants can eliminate it by donating an electron or hydrogen atom to the DPPH radical, which transforms into a stable molecule, causing a colour change from purple to yellow<sup>[49]</sup>. In the present study, the DPPH assays were represented as IC<sub>50</sub> value and the ethanolic extract exhibited rich scavenging effects on DPPH (6.32±0.04 mg/ml) when compared to aqueous extract with an IC<sub>50</sub> value of 9.45±0.03 mg/ml respectively (Table 1). The differences in antioxidant activity among the aqueous and ethanolic seaweed extract might be due to the changes in their chemical compositions, resulting in a significant difference in antioxidant activity. The stronger radical scavenging activity is linked to the ethanolic extract's phenolic content.

Similarly, Siriwardhana *et al.*<sup>[50]</sup> and Liu *et al.*<sup>[51]</sup>, reported a high correlation between DPPH radical-scavenging activities and total polyphenolics. Components, such as polysaccharides, pigments, proteins or peptides in seaweed, also impart antioxidant activity. The study found that the use of ethanolic solvent extraction resulted in the highest level of antioxidant activity with a similar correlation to the levels of phenolic compounds.

The present study evaluated the antibacterial properties of *H. pannosa* extracts using the disc diffusion method (Table 3 and fig. 1A-fig. 1E). The results showed that the extracts had significant antibacterial activity against the tested bacteria (*Staphylococcus aureus*, *Escherichia coli*, *Bacillus subtilis*, *Bacillus cereus* and *Pseudomonas*

*aeruginosa*), indicating that the seaweed powder used in the extraction contained antimicrobial principles. The zone of inhibition was calculated for both aqueous and ethanolic extracts, with the ethanol extract exhibiting a more significant inhibition zone than the aqueous extract. The ethanolic extract was especially effective against *Staphylococcus aureus*, *Escherichia coli* and *Pseudomonas aeruginosa*, with inhibition zones of 8 mm, 14.5 mm and 14 mm respectively. The aqueous extract was effective against

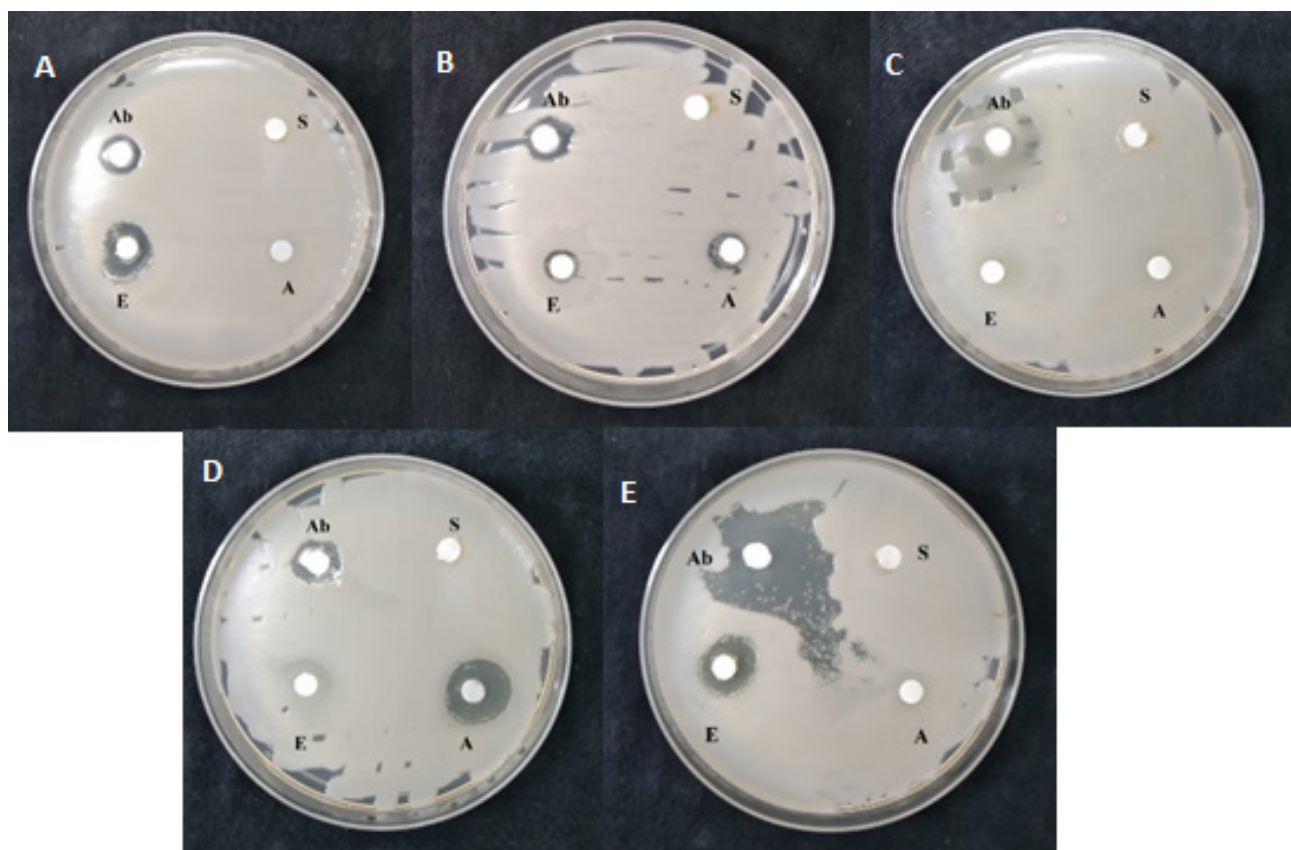
*Staphylococcus aureus* (9 mm) and *Bacillus cereus* (15 mm). However, the extracts were less potent than the standard ampicillin.

Moreover, some plant extracts could not exhibit antibacterial activity against tested bacterial strains<sup>[52]</sup>. Further, the antimicrobial activity of seaweed species is linked to the presence of phenols and flavonoids<sup>[53]</sup>. The study demonstrated that the ethanol extract was effective against skin bacteria and could be used as a cosmeceutical application to treat acne and bacterial skin infections.

**TABLE 3: IN VITRO ANTIBACTERIAL ACTIVITY OF *H. pannosa* EXTRACTS**

Bacteria	Zone of inhibition in mm		
	Ab	Aqueous extract	Ethanol extract
<i>E. coli</i>	10	-	14.5
<i>Staphylococcus aureus</i>	11	9	8
<i>Bacillus subtilis</i>	8	-	-
<i>Bacillus cereus</i>	11	15	-
<i>Pseudomonas aeruginosa</i>	15	-	14

Note: Ab: Ampicillin used as a positive control



**Fig. 1: In vitro antibacterial activity of the *H. pannosa* extracts**

Note: E: Ethanol extract; A: Aqueous extract; Ab: Ampicillin and S: Seaweed powder

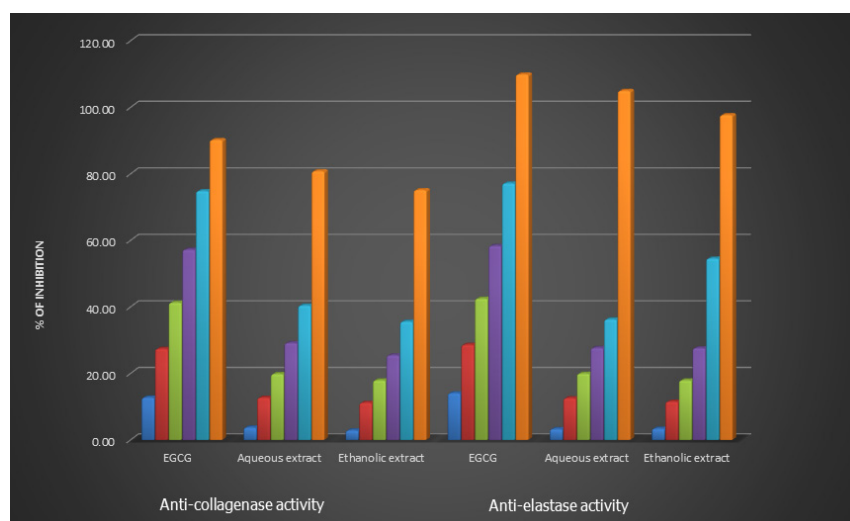
The MTT cytotoxicity assay is used to evaluate the cytotoxicity of seaweed extracts. An active compound with anticancer activity is represented by an  $IC_{50}$  value ( $\mu\text{g/ml}$ ) of less than 100. A weak anticancer activity is represented by an  $IC_{50}$  value ranging from 100 to 300, while an inactive compound has an  $IC_{50}$  value over 300<sup>[54]</sup>. The present study showed that the ethanol extract had an  $IC_{50}$  value of 95.70, while the aqueous extract had an  $IC_{50}$  value of 110.70  $\mu\text{g/ml}$  (Table 4). The results of this assay suggest that the ethanol extract has the potential cytotoxic activity against COLO 829 cancer cell line. The anticancer activity of seaweeds may be related to the presence of various therapeutic compounds that can induce apoptosis through different pathways and molecular mechanisms<sup>[55]</sup>. *In vitro* cytotoxic capacity of the ethanolic extract might be due to its antioxidant activity. In the present study, the cytotoxic effect of the ethanolic extract was probably due to the flavonoids that scavenge free radicals and prevent oxidative damage, which have a powerful anticancer effect and protect cells against all stages of carcinogenesis<sup>[56]</sup>. The present study proved that ethanolic extract of *H. pannosa* are a rich source of anticancer agents as the presence of bioactive

properties is mainly due to the alkaloids, tannins, sterols and terpenoids.

The present study was conducted to test the elastase inhibitory effects of *H. pannosa* extract using the SK-MEL human melanoma cell line. The results showed that the extract inhibited elastase in a dose-dependent manner. It was noted that the ethanol extract was more effective compared to the aqueous extract, with an  $IC_{50}$  value of 48.53 and 48.65  $\mu\text{g/ml}$ , respectively (Table 4, fig. 2 and fig. 3a). However, it was observed that both extracts were found to be more potent than the standard catechin, Epigallocatechin Gallate (EGCG) (49.63  $\mu\text{g/ml}$ ). Studies have shown that plant extracts have been identified as elastase inhibitors<sup>[57]</sup>, but there have been limited investigations on the same from algae resources. Research has indicated that polyphenols extracted from plants possess strong elastase activity<sup>[58]</sup>. Elastase activity might be significantly inhibited by phenols such as epicatechin, catechin and catechol<sup>[45]</sup> and flavonoids such as kaempferol, quercetin, and myricetin<sup>[59]</sup>. Therefore, the study proved that the ethanolic extracts of *H. pannosa* could be used to develop cosmetic agents to prevent skin aging.

**TABLE 4:  $IC_{50}$  VALUES OF ANTICANCER, ANTI-ELASTASE AND ANTI-COLLAGENASE ACTIVITY OF *H. pannosa* EXTRACTS**

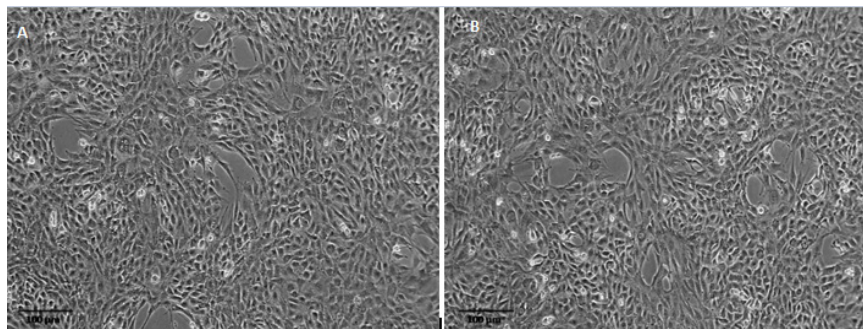
Bioactive properties	$IC_{50}$ value ( $\mu\text{g/ml}$ ) of aqueous extract	$IC_{50}$ value ( $\mu\text{g/ml}$ ) of ethanol extract
Anti-cancer	110.70	95.70
Anti-elastase	48.65	48.53
Anti-collagenase	49.32	48.76



**Fig. 2: *In vitro* anti-collagenase and anti-elastase activity of *H. pannosa* extracts and positive control (EGCG)**

**Note:** (■): 6.25; (■): 12.50; (■): 25.00; (■): 50.00; (■): 100.00 and (■): 200.00





**Fig. 3:** SK-MEL cells treated with *H. pannosa* extract; (a): For *in vitro* anti-elastase activity and (b): For *in vitro* anti-collagenase activity

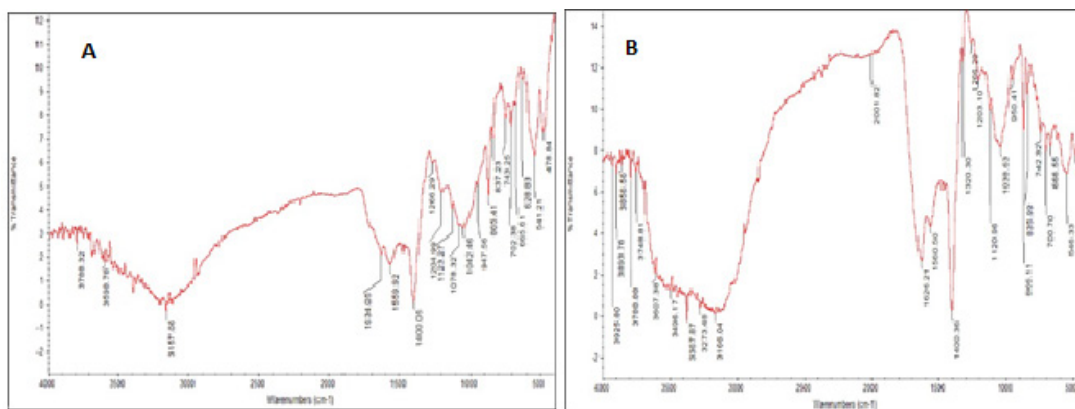
The degradation of collagen occurs with ageing due to collagenase activity resulting in wrinkles on the skin<sup>[60]</sup>. The present study investigated the effects of *H. pannosa* extracts on collagenase inhibition using the SK-MEL human melanoma cell line. Results suggest that the extracts inhibit collagenase in a dose-dependent manner. The ethanol extract proved more effective than the aqueous extract, with an  $IC_{50}$  value of 48.76 compared to 49.32  $\mu\text{g/ml}$  (Table 4, fig. 2 and fig. 3b). However, both extracts were more potent than the standard catechin, EGCG (49.74  $\mu\text{g/ml}$ ). The presence of polyphenols in the seaweed extract may be responsible for its ability to inhibit collagenase activity. Based on these findings, the ethanolic extracts of *H. pannosa* show promising natural bioactive agents for future formulations aimed at reducing wrinkles and skin sagging.

FT-IR analysis was conducted to identify and characterize the functional groups present in the extracts. FT-IR spectra can provide valuable information for identifying samples with specific functional ingredients<sup>[61]</sup>. The results of FT-IR analysis confirmed the presence of phenols, carboxylic acids, aromatics, ketones, ethers, amides/amines and sulfonates compounds at varying concentrations. The ethanol extract exhibited wave patterns 3925, 3893, 3856 and 3788  $\text{cm}^{-1}$  corresponding to OH stretching (Hydroxy) compounds, while the aqueous extract exhibited wave patterns 3788.32, 3598.78  $\text{cm}^{-1}$  corresponding to OH stretching (carboxylic acid) (fig. 4). However, Rafiquzzaman *et al.*<sup>[23]</sup>, also used FT-IR to identify several phytochemicals such as phenols, carboxylic acids, ketones, ethers, aromatics, amides and sulfonates from *H. musciformis*. Therefore, it was concluded that the ethanolic extract contains high levels of phenolic compounds, making them valuable for

cosmeceutical applications.

In conclusion, it is clear from the results presented here that the type of seaweed used for extraction and the solvent choice substantially impact the availability of functionally bioactive compounds, as demonstrated by *in vitro* screening and FT-IR analysis. The results showed that seaweed extracts possess various phytochemical substances and potent antioxidant properties, with varying levels of total phenolics, flavonoids and DPPH activity. Among the solvents, the ethanolic extract was the most effective in obtaining an antioxidant-rich extract with high phenolics compared to the aqueous extract, possibly due to high polyphenols, flavonoids and polysaccharides. The bioactive properties of the extract, including its *in vitro* antibacterial, anticancer, anti-elastase and collagenase activity, were also tested. The ethanol extract had good antibacterial, anti-ageing and anticancer properties compared to the aqueous extract. FT-IR analysis revealed that ethanolic extract is abundant in polyphenolic compounds, which have implications for cosmeceuticals. Using ethanol extract as an ingredient in cosmetic products could boost their efficacy. Further research should investigate the purification of each extract and the effects of different combinations. Finally, the nutrient content, health benefits and chemical composition of seaweed can differ depending on its origin, with factors such as water temperature, salinity and environmental pollutants playing a role. It is important to choose seaweed harvested from safe and sustainable sources. The present study has revealed that the red seaweed *H. pannosa*, sourced from the Mandapam coast, boasts advantageous cosmeceutical properties. Notably, the seaweed's ethanolic extract could serve as a natural antioxidant in cosmetics and pharmaceuticals.





**Fig. 4:** FT-IR analysis of *H. pannosa* extracts

**Note:** (A): Aqueous extract and (B): Ethanolic extract

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### Conflict of interests:

The authors declared no conflict of interests.

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