Atypical Log D Profile of Rifampicin

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The distribution coefficient (log D) values of rifampicin, an essential first-line antitubercular drug, at gastrointestinal pH conditions are not reported in literature. Hence determinations were made using n-octanol and buffers ranging between pH 1-7. Also, log D values were predicted using Prolog D. Both the determinations showed opposite behaviour. The atypical experimental log D profile of rifampicin could be attributed to its surface-active properties, which also explained the reported permeability behaviour of the drug in various gastrointestinal tract segments.

Distribution coefficient (log D), measured in two-phase bulk solvent systems at different pH, is one of the main descriptors for prediction of *in vivo* drug permeation¹. Its values for rifampicin, an essential first-line antituberculosis drug, at different gastrointestinal pH are not reported in the literature. Only values at selected pH are known, along with predicted partition coefficient data, which are listed in Table 1²⁻⁷. Accordingly, the purpose of the present study was to determine log D values for rifampicin at various pH between 1 and 7. The log D values were also calculated using Prolog D (Pallas, CompuDrug Chemistry, Budapest, Hungary). Unexpectedly, an opposite behaviour was observed among the experimental and predicted values, which indicated an atypical experimental log D profile for rifampicin. The observed behaviour is discussed critically in this communication, based on the solubility and surface activity data, and related to the known permeability behaviour of the drug in various segments of gastrointestinal tract.

MATERIALS AND METHODS

Rifampicin was a gift sample from M/S Panacea Biotec Ltd., Lalru, India. Buffer materials and all other chemicals were of analytical-reagent (A.R.) grade. n-octanol (A.R. grade) was procured from Central Drug House Pvt. Ltd., New Delhi, India. HPLC grade acetonitrile and methanol were procured from J.T. Baker (Mexico City, Mexico) and Mallinckrodt Baker Inc. (Paris, KY, USA), respectively. Ultra-pure water was obtained from an ELGA water purification unit (Elga Ltd., Bucks, England).

*For correspondence E-mail: ssingh@niper.ac.in pH recordings were made on a research pH meter (MA 235, Mettler Toledo GmbH, Schwerzenbach, Switzerland). Surface and interfacial tensions were measured using a tensiometer (K9, Kruss GMbH, Hamburg, Germany). Other equipment used was a sonicator (Branson Ultrasonic Corporation, Danbury, CT, USA) and a centrifuge (15, Biofuge, Heraeus, Hanau, Germany). The HPLC system consisted of an on-line degasser (DGU-14AM), low-pressure gradient flow control valve (FCV-10AL_{VP}), solvent delivery module (LC-10AT_{VP}), auto injector (SIL-10AD_{VP}), column oven (CTO-10AS_{VP}), UV-visible dual-wavelength detector (SPD-10A_{VP}), system controller (SCL-10A_{VP}) and CLASS-VP software (all from Shimadzu, Kyoto, Japan).

HPLC analyses:

Rifampicin was analyzed by a reported gradient stabilityindicating method⁸ using a Zorbax XDB C-18 column (250×4.6 mm, 5 μ) from Agilent Technologies, Wilmington, USA. The mobile phase was composed of acetonitrile and a buffer consisting of 0.01 M sodium dihydrogen orthophosphate (pH adjusted to 6.8 with dilute orthophosphoric acid).

Preparation of buffers, and saturation of buffers and n-octanol:

The buffers of various pH were prepared according to the formulae given in Table 2⁹. These were saturated with n-octanol for 24 h before use. Octanol was also saturated before the study with respective buffer solutions.

Determination of log D:

The distribution coefficients of rifampicin at different pH

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TABLE 1: REPORTED EXPERIMENTAL AND PREDICTED DISTRIBUTION/PARTITION COEFFICIENT VALUES OF RIFAMPICIN

Medium or software employed	Log D or Log P	Reference
Reported experimental log D values		
n-Octanol-phosphate buffer, pH 7.4	1.19	2
Not mentioned	2.40	3
n-Octanol-Dulbecco's phosphate buffered saline, pH 7.4	1.25	4
Predicted values of log P		
Clog P (Chemdraw ultra)	3.34	-
Alog P	2.95	5
Log K _{w/a} (KOWWIN)	4.24	6
Log P (KOWWIN)	2.50	7
Log P (MDL QSAR)	5.57	7

TABLE 2: COMPOSITION	OF E	BUFFERS	USED	IN THE	STUDIES
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pН	HCl(ml)	Citric acid (g)	Na ₂ HPO ₄ (g)	$KH_2PO_4(g)$	NaOH(g)	NaCl(g)	Water
1	9.2	-	-	-	- 77 -	6.1	q.s. to make II
2	6.7	6.3	-	-	2.1	4.2	
3	5.7	8.4	-	-	3.2	3.4	
4	3.8	12.6	-	-	4.8	2.1	
5	-	21.0	-	-	8.0	-	
6	-	-	2.4	8.7		6.6	
7	-	-	14.3	3.6	\dot{C}	4.3	

HCl, hydrochloric acid; Na₂HPO₄, disodium hydrogen phosphate; KH₂PO₄, potassium dihydrogen phosphate; NaOH, sodium hydroxide; NaCl, sodium chloride.

were determined by the standard shake-flask method¹⁰. For this purpose, rifampicin solution at a concentration of 500 µg/ml was prepared in buffer pre-saturated n-octanol to which an equal portion of octanol pre-saturated buffer was added. The tube was shaken horizontally for 100 fores in 5 min, centrifuged at 5,000 rpm (2200 × g) for 10 rnn and 56 organic layer was analyzed by HPLC. The drug concentration in aqueous medium (Ca₁) was calculated by subtracting the drug concentration is octand layer (Coct), from the initial drug concentration. Log D was expressed as log (Corg/Caq). The experiment was carried out in triplicate for all investigated pH values. Determinations were also made by oppositely preparing arug solutions in octanol pre-saturated buffers, to which buffer pre-saturated n-octanol was added, followed by shaking, centrifugation and analysis of the organic layer.

The determination of log D values of rifampicin at different pH by shake flask method was validated by repetition of the determinations by three different analysts. The experiments were also carried out at two other drug concentrations (100 and 1000 μ g/ml), and additionally at a ratio of 1:10 of n-octanol to buffer.

Determination of surface and interfacial behaviour:

Surface tension of rifampicin solutions (500 μ g/ml) at pH 1 to 7 was determined using a Du-Nuoy Tensiometer. For the measurements, the suitability of the instrument was first assessed by determining the surface tensions of four

standard liquids, *viz.*, water, methanol, ethanol and ethylene glycol. The values for these liquids were found o be within the limit of±1 mN/m from the reported values, hence allowing direct use of the equipment. Subsequently, the ring was immersed into the sample, and the instrument reading was set at zero. Thereafter, the sample support was lowered carefully until a film was formed between the ring and the liquid surface. The measurement of surface tension was completed when the maximum value was shown on the display. This was repeated three times for each sample. All determinations were made between 22° and 25°. The surface activity was expressed as surface pressure¹¹, which was calculated by subtracting the surface tension of drug solution (γ_{drug}) from the surface tension of blank buffer ($\gamma_{subsert}$).

The same instrument was also used for the determination of interfacial tension of n-octanol-buffer systems. For this purpose, the ring of the tensiometer was first dipped into n-octanol and the reading was set at zero. Thereafter, the container was removed and replaced by buffer solutions containing the drug at a concentration of 500 μ g/ml. After dipping the ring, n-octanol was poured on the top of the buffer through the side of the container. The ring was then slowly lifted and the reading was noted when it crossed the interface of the buffer and n-octanol bilayer.

Prediction of log D:

Log D values of rifampicin at different pH were also

TABLE 3: LOG D VALUES OF RIFAMPICIN DETERMINED BY PREPARING DRUG SOLUTIONS IN OCTANOL AND BUFFER, AND INTER-ANALYST VARIATION OF VALUES

Observed pH	$Log D \pm SD (n = 3)$				
	Analyst 1	Analyst 2	Analyst 3	Analyst 1	
	Drug solution* in n-octanol		Drug solution in buffer		
1.12	1.606±0.042	1.409±0.049	1.530±0.037	1.525±0.062	
2.06	1.317±0.018	1.258±0.009	1.248±0.067	1.260±0.002	
2.97	1.171±0.035	1.156±0.029	1.139±0.047	1.132±0.004	
4.03	1.049±0.012	1.078±0.036	1.032±0.026	1.087±0.005	
5.06	0.992±0.001	0.977±0.032	0.969±0.083	1.008±0.001	
6.08	0.765±0.020	0.755±0.028	0.847±0.057	0.896±0.036	
7.06	0.736±0.002	0.705±0.021	0.806±0.043	0.881±0.001	

*Drug concentration was 500 μ g/ml in each case. The organic phase:buffer ratio was 1:1. SD, standard deviation.

predicted using Prolog D, a module of Pallas Cluster, package from CompuDrug Chemistry (Budapest, Hungary). The software allows prediction of ADME-Tox parameters based on the structural formulae of the organic compounds.

Determination of solubility profile:

Solubility of rifampicin was determined between pH 1 and 7 by the shake flask method. For the same, an excess amount of drug was placed in 30 ml vials and 5 ml buffer solution of respective pH was added. The vials were incubated together in a shaking water bath at 37°. Samples were withdrawn after saturation was achieved and analyzed by HPLC after appropriate dilution. All the studies were conducted in triplicate.

RESULTS AND DISCUSSION

Table 3 lists the log D values of rifampicin determined at different pH by three analysts. The drug solutions in this case were made in n-octanol. It shows that the values determined by three different analysts were consistent to each other. It also depicts that log D values decreased as the pH increased from 1 to 7. Fig. 1 shows the photograph of the Eppendorf tubes containing the two liquid phases after shaking and centrifugation. It depicts that at all the pH, most of the drug partitioned into the



Fig. 1: Partitioning of rifampicin in octanol/buffer layers at various pH

The photograph shows that the drug preferentially partitions into octanol (upper layer) between pH 1-7, though a slight increasing amount of drug remains in the aqueous layer beyond pH 4. octanol layer, which explains the positive log D values in Table 3. The photograph shows a slight and increasing yellow colour in aqueous layer at pH>3, which justifies the decrease in log D values with increase in pH. A same behaviour was also observed when the experiment was repeated by preparing drug solutions in buffer, instead of n-octanol. The data for the reverse experiment are also given in Table 3, which clearly show similarity of values with those obtained by preparing drug solutions in n-octanol. Visibly also, most of the drug was found to be immediately transferred from the aqueous layer to the organic layer, after addition of organic layer to the buffer containing the drug. A similar observation was also made by Mannisto¹², who reported that dissociated and undissociated forms of rifampicin were always found in butanol layer, when water-lipid partition experiment was done at wide range of pH conditions. Unfortunately, the report neither gives the profile nor values of log D of rifampicin at various pH in butanol/buffer system.

Table 4 gives the log D values determined at 100 and 1000 μ g/ml. Evidently, there was no significant influence of the drug concentration on the determined log D values. Table 5 lists the log D values obtained using 1:10 ratio of n-octanol:buffer. This study with lesser amount of octanol was justified as rifampicin partitioned preferentially

TABLE 4: INFLUENCE	OF DRUG CONCENTRATION*
ON LOG D VALUES OF	

Observed pH	$Log D \pm SD (n = 3)$			
	100 μg/ml	1000 μg/ml		
1.12	1.456±0.070	1.549±0.045		
2.06	1.323±0.034	1.276±0.044		
2.97	1.247±0.006	1.159±0.016		
4.03	1.074±0.010	1.095±0.031		
5.06	1.042±0.019	1.037±0.020		
6.08	0.871±0.069	0.852±0.044		
7.06	0.842±0.061	0.843±0.042		

*Solutions were prepared in n-octanol saturated for 24 h with buffer phase. The organic phase:buffer ratio was 1:1. SD, standard deviation.

into octanol. The change in the ratio of octanol:buffer from 1:1 to 1:10 did not result in change in log D values or even the trend of decrease in log D values with an increase in pH (Table 3).

Fig. 2 shows the comparison of the log D values determined by shake flask method and the predicted data using Prolog D. It is evident that the two profiles were exactly opposite. In case of experimental determination, the log D values were positive and decreased as the pH was increased. On the other hand, the log D values predicted from the software were all negative, and the curve was sigmoidal in nature, which rose sharply beyond pH 3.

The solubility of rifampicin at different pH is given in Table 6. Evidently, the solubility of the drug was very high at acidic pH (pH \leq 2) and decreased as the pH was increased. The profile conformed to the known ionization behaviour of the drug². Accordingly, it was expected that rifampicin would remain in water in acid region and partition to a lesser extent into the organic phase. But an opposite pattern was observed in the present study. As apparent from the data in Tables 3-6, rifampicin partitioned more strongly into the organic layer at lower pH. Thus solubility data did not explain the atypical log D behaviour of rifampicin.

In that situation, the amphoteric nature of rifampicin² and hence its surface activity was considered as the reason for the atypical log D behaviour and quantitative transfer of the drug into organic medium between pH 1-7. Such a

TABLE 5: LOG D OF RIFAMPICIN* DETERMINED USING 1:10 RATIO OF N-OCTANOL-BUFFER SYSTEM

Observe	ed pH	Log D
1.12		1.522±0.007
2.06	0	1.325±0.005
2.97		1.291±0.007
4.03		1.210±0.004
5.06		1.161±0.003
6.08		0.955±0.001
7.06		0.888±0.004

*The concentration of drug solution employed was 500 μ g/ml.

TABLE 6: SOLUBILITY OF RIFAMPICIN AT VARIOUS pH

Observed pH	Solubility \pm SD ($n = 3$) (mg/ml)
1.12	127.20±12.2
2.06	19.20±1.57
2.97	0.24±0.03
4.03	0.19±0.02
5.06	0.44±0.05
6.08	0.70±0.04
7.06	0.85±0.13
CD standard doviation	

SD, standard deviation



Fig. 2: Log D-pH profiles of rifampicin

Plots of experimentally determined (\blacksquare) and predicted (\blacktriangle) log D values of rifampicin at different pH. The experimental values are the mean of three determinations made by different analysts (Table 3). Evidently, the profiles are opposite in nature.



Fig. 3: Profiles for surface and interfacial tensions of rifampicin at various pH

Plots of pH *versus* surface tension (\blacktriangle), and interfacial tension at the octanol:buffer interface (\blacksquare). The two show a parallel behaviour. Surface and interfacial tensions are expressed in mN/m. Drug concentration was 500 µg/ml in both the cases.





The figure shows the parallel relationship between the profiles of log D (\blacksquare) and surface pressure (\blacktriangle) of rifampicin at different pH.

characteristic is known for other amphoteric drugs like celecoxib, meloxicam and nimesulide, which possess surface-active properties¹¹. Fig. 3 shows the plots of surface pressure (difference between the surface tension of the drug solution to that of blank buffer) of rifampicin and its log D values at various pH conditions. The plots were parallel in nature, thus confirming the contention that higher experimental log D values for rifampicin in acidic pH range were due to higher surface activity of the drug. In addition, the parallel behaviour between surface and interfacial tension (fig. 4) indicated that not only the surface activity, but interfacial tension also contributed towards preferential transfer of rifampicin to the organic (octanol) layer at acidic pH.

This reason also explains the higher permeability of rifampicin through stomach as compared to intestine¹³. It is reported that drugs with surface-active property reduce the surface tension between drug solution and biological membrane, resulting in concentration of more number of drug molecules at the biological interface, where they interact with charged carriers to form neutral species. These neutral molecules cross the biological membrane, and hence show good absorption through stomach¹⁴. The same is postulated to happen in the case of rifampicin.

REFERENCES

...d Kontturi, K., Eu Malkia, A., Murtomaki, L., Urtti, A. and Kontturi, K., Eur. 1.

Pharm. Sci., 2004, 23, 13.

- Gallo, G.G. and Radaelli, P. In; Florey, K., Eds., Analytical Profiles of 2 Drug Substances, Academic Press, London, 1976, 467.
- Washington, N., Lamont, G., Wilson, C.G., Washington, C. and 3. Withington, R., Int. J. Pharm., 1994, 108, 125.
- 4 Available from http://www.cerep.fr/cerep/users/pages/downloads/ Documents/Marketing/ Pharmacology 20% & 20% ADME/Application 20% notes/ PartitionCoefficient.pdf (Accessed on 15-08-2006).
- Barry, C.E., Slayden, R.A., Sampson, A.E. and Lee, R.E., Biochem. 5. Pharmacol., 2000, 59, 221.
- 6. Rodriques, C., Gameiro, P., Reis, S., Lima, J.L.F.C. and Castro, B.D., Biophys. Chem., 2001, 94, 97.
- Available from http://chemdb.niaid.nih.gov/struct_search/lr/ 7. LR_HIV_OI.asp?LITREF=12951 (Accessed on 15-08-2006).
- 8. Mohan, B., Sharda, N. and Singh, S., J. Pharm. Biomed. Anal., 2003, 31, 607.
- Koizumi, T., Arita, T. and Kakemi, K., Chem. Pharm. Bull., 1964, 9 12, 413.
- 10. EPA. Prevention, pesticides and toxic substances (7101): Product properties test guidelines OPPTS 830.7550. Partition coefficient (noctanol/water), shake flask method. EPA 712-C-96-038, United States Environmental Protection Agency, 1996, 1.
- 11. Seedher, N. and Bhatia, S., Indian J. Pharm. Sci., 2004, 66, 254.
- 12. Mannisto, M.D.P., Clin. Pharmacol. Ther., 1976, 21, 370.
- 13. Mariappan, T.T. and Singh, S., Int. J. Tuberc. Lung Dis., 2003, 7, 797.
- 14. Fiese, G. and Perrin, J.H., J. Pharm. Sci., 1969, 58, 599.

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