

Chemical and Biological Investigations on Mangrove Associated Lichens (Manglicolous Lichens) from India: A Review

V. B. TATIPAMULA, K. N. KILLARI^{1*}, HARITHA POLIMATI¹ AND ALEKHYA KETHA¹

Institute of Research and Development, Duy Tan University, Da Nang 550000, Vietnam, ¹Pharmaceutical Sciences Department, AU College of Pharmaceutical Sciences, Andhra University, Visakhapatnam 530003, India

Tatipamula *et al.*: Studies on Manglicolous Lichens

In 370-285 BC, an itinerant Greek philosopher Theophrastus phrased the term 'Lichen', which is a symbiosis between mycobiont (fungus) and a photobiont (algal) or a cyanobiont (cyanobacterium) where the mycobiont provides the shape and structure, while the photobiont or cyanobiont supply food, i.e., carbohydrates. From the folklore, lichen and its substances are used for various biological activities, which include antimicrobial, anticancer, anti-inflammatory, antioxidant and enzyme inhibitory activities. It is interesting to note that lichens are epiphytes as they synthesis their food by photosynthesis by using plants, rocks and dead logs as a substrate. In particular, lichens that betide to mangroves are termed as "Manglicolous Lichens". Generally, mangroves survive in a unique environment having a high concentration of salt and moisture, low and high tidal water. Hence, lichens growing in such stressful areas may lead to contain variation in their chemical constituents as well as pharmacological actions due to their defensive mechanism from the predators. Keeping the factors mentioned earlier, researchers worked and reported new lichen substances with potent biological properties from manglicolous lichens. The present review, list out all the chemical constituents and biological activities that are reported to date from manglicolous lichens in India. Also, this review remains useful for further research on manglicolous lichens to identify the potential bioactive molecules.

Key words: Mangroves, lichen substances, biological activity, manglicolous lichens, secondary metabolites

Mangroves are the only plants that exist in between land and sea waters. A unique ecosystem is built in and around mangroves with circumstances of anaerobic muddy soils, extreme tides, high temperatures and salinity, strong winds^[1,2]. This unique ecosystem favors assemblage of many species, especially the muddy and sandy soil sediments attract bacteria, fungi, macroalgae and invertebrates and also the aerial roots and twigs host other groups of organisms^[3-7]. Because of the adaptive characteristics, the ecology of mangroves has been extensively studied.

Usually, the harsh stressful environments of mangroves, for instance, variations in tidal water, also effects the epiphytes such as lichens that are habituated on them^[8-12]. Lichen is a symbiotic organism, which is an association of fungi and algae and the lichens growing particularly on the mangroves are named as manglicolous lichens. Generally, the fungal matter present in lichens will die if they come in contact with

saltwater. Hence, it is a sporadic and unique case to find out lichens in mangrove forest regions. Moreover, lichens growing in such stressful areas may also lead to contain variation in their chemical constituents as well as pharmacological actions due to habitat under stress conditions and also their defensive mechanism from the predators^[12,13].

Also, the identification of lichens is always misleading because of the symbiotic association of a microscopic algae or cyanobacterium with a filamentous fungus. Due to this, even they have full applications in human life, the studies carried out on them till now are minute^[14,15]. Furthermore, it is substantial to study manglicolous

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*Address for correspondence

E-mail: kishorenaidu.killari@gmail.com

Accepted 03 March 2021

Revised 02 January 2021

Received 25 February 2020

Indian J Pharm Sci 2021;83(2):186-194

lichens because of climatic factors in mangrove areas, fewer modes of transport and time lagging process due to damp soil, backwater flow and aerial roots (which adjourns the field survey). So a survey of especially manglicolous lichen is an important task^[16]. As the lichens are generally termed as bio-monitors, they are very less resistant towards pollution. Even though there is less possibility of having pollution problems in and around mangrove regions, the lichens are profoundly affected by acid rains (which contain high amounts of sulfur dioxide). Eventually, acid rains affect the availability of lichens in mangroves^[17-20].

The literature on manglicolous lichens is very less. Long back, lichens on mangroves are included either within flora of a region or in monographic studies. Mangroves have rich and interesting lichen flora^[21]. Initially, studies on epiphytic community growing on a mangrove tree *Kandelia candel* were performed which described several lichens and bryophytes, further ecology of mangrove lichens and environmental factors influencing pattern of lichen distribution in mangroves of Southeast Brazil were carried out, which leads to the updates of existing list of lichens of about 81 lichen genera from Southern Brazil and Itanhaem river mangroves. Similarly, unique lichens species of *Peltigera* from mangroves swamps of Colombian Caribbean coast were reported; this exemplifies the lichen richness in mangroves vegetation of tropical belt. Based on these, several studies were framed by several men like favorable growth factors of lichens on mangrove trees in inland streams to those growing near the river mouths and open seas^[22-26].

Though India has a large coastal area under mangrove vegetation under ten states and Union Territories, independent studies on lichens are rare in these areas, the studies of lichen flora in West Bengal state (a part of Sundarbans and Parmadan Forest) revealed 160 lichen taxa and proves the existence of diverse lichen flora in Sundarbans mangroves. Additionally, an in-deep survey on Sundarbans mangroves identified 550 specimens, which resulted in 165 species, of which 25 species were new records to India. Likewise, 13 and 12 lichens species were recorded from Muthupet lagoon and Pichavaram mangroves, respectively and some impressive records of *Roccellaceae* from the Gujarat mangroves were also recorded. Apart from these, there are no other reports of manglicolous lichens from other areas of India^[26-28].

Keeping all the drawbacks above and aspects, there are limited reports on the chemical and biological

evaluation of particularly mangrove associated lichens. This review, list out all the chemical and biological investigations of associated mangrove lichens, till now. The list of manglicolous lichens that are examined were listed below.

Dirinaria consimilis (Stirton) D. D. Awasthi^[29], Host: twigs of *Avicennia officinalis* Location: Vainateya Island, Godavari estuary, West Godavari District, Andhra Pradesh, India.

Graphis ajarekarii Patw. & C.R. Kulk^[30], Host: twigs of *Avicennia officinalis*, Location: Nagayalanka mangroves, Krishna estuary, Krishna District, Andhra Pradesh, India.

Parmotrema tinctorum (Despr. ex Nyl.)^[29], Host: twigs of *Excoecaria agallocha*, Location: Bhitharkanika Island, Mahanadi estuary, Odisha, India.

Ramalina leiodea (Nyl.) Nyl.^[30], Host: twigs of *Ceriops decandra*, Location: Bhitharkanika Island, Mahanadi estuary, Odisha, India.

Roccella montagnei Bel em. D. D. Awasthi^[29], Host: twigs of *Excoecaria agallocha*, Location: Vainateya Island, Godavari estuary, West Godavari District, Andhra Pradesh, India.

LICHEN SUBSTANCES ISOLATED FROM MANGLICOLOUS LICHENS

The lichen substances that are isolated from the above species are represented in fig. 1.

2'-O-methyl divaricatic acid^[31]:

$C_{22}H_{26}O_7$ (402.44)

White powder

Retention factor (Rf): 0.4 (1:1 ratio of hexane and ethyl acetate)

Wavelength of maximum absorbance (λ_{max}): 212.5 nm

Melting point (Mp): 142-143°

Elemental analysis: C-65.66, H-6.62 (%)

Electrospray ionization-mass spectrometry (ESI-MS)
m/z: 403.05 [M+H⁺]

Proton nuclear magnetic resonance (¹H NMR) (Deuterated chloroform (CDCl₃), 400 MHz): 11.73 (s, 1H), 11.18 (s, 1H), 6.46 (s, 1H), 6.40 (s, 2H), 3.91 (s, 3H), 3.85 (s, 3H), 2.95-3.04 (m, 4H), 1.66-1.80 (m, 4H), 0.93-1.05 (m, 6H).

Carbon-13 nuclear magnetic resonance (¹³C NMR)

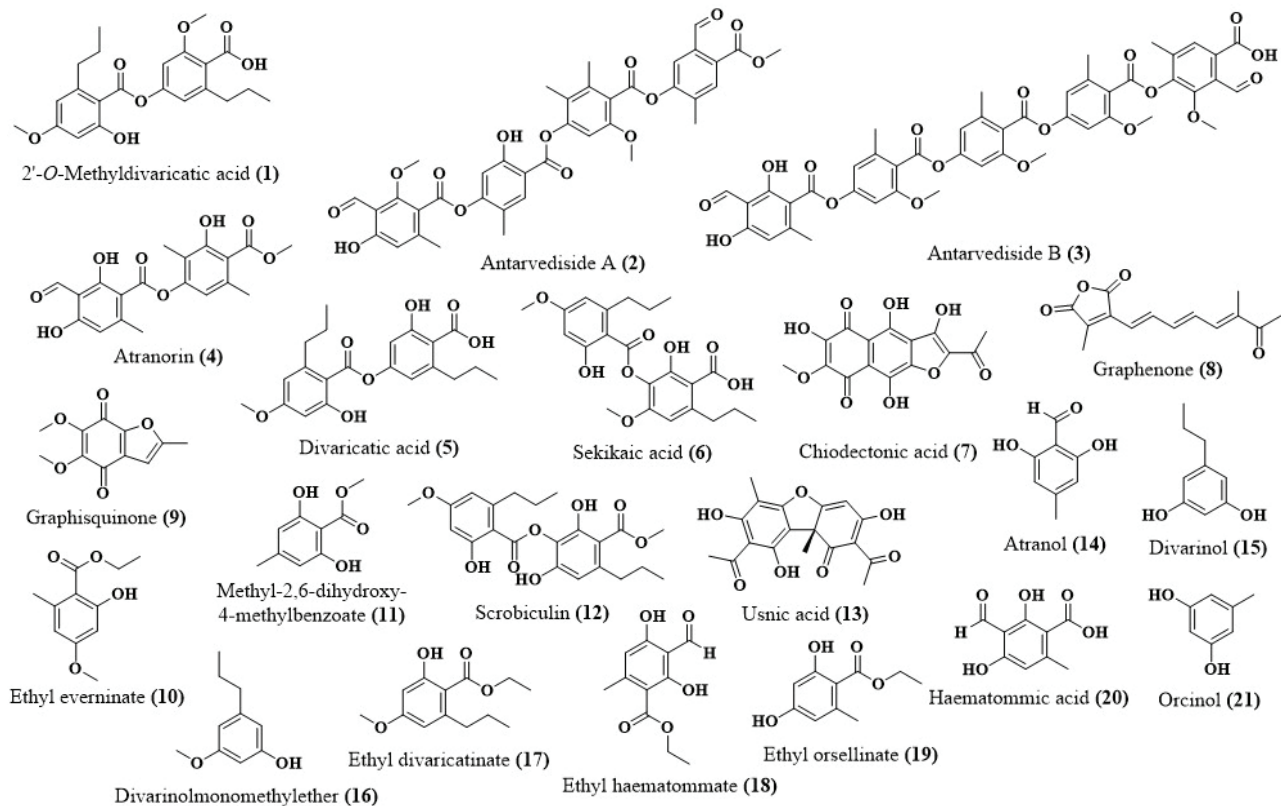


Fig. 1: Chemical structures of identified lichen substances from manglicolous lichens

(CDCl₃, 400 MHz): 14.35, 14.36, 24.84, 25.15, 38.87, 39.10, 55.37, 56.05, 98.89, 104.45, 104.45, 104.65, 106.48, 110.97, 124.94, 147.31, 148.73, 156.40, 156.94, 164.42, 165.55, 168.91 and 175.47

Antarvediside A^[31]:

C₃₈H₃₄O₁₄ (714.68)

Pale pink needles

R_f: 0.7 (9:1 ratio of hexane and dichloromethane)

λ_{max}: 222 nm

Mp: 191-192°

Elemental analysis: C-63.76, H-4.82 (%)

ESI-MS m/z: 715.00 [M+H⁺]

¹H NMR (CDCl₃, 400 MHz): 12.55 (s, 1H), 12.50 (s, 1H), 11.95 (s, 1H), 10.36 (s, 1H), 6.52 (s, 1H), 6.41 (s, 1H), 3.99 (s, 3H), 2.69 (s, 3H), 2.56 (s, 3H), 2.18 (s, 3H), 2.09 (s, 3H).

¹³C NMR (CDCl₃, 400 MHz): 9.39, 23.46, 24.05, 25.60, 29.73, 52.36, 55.67, 55.72, 92.08, 96.78, 98.47, 102.85, 104.13, 108.56, 110.28, 110.53, 112.87, 112.96, 115.46, 116.03, 116.80, 139.89, 143.47, 152.00, 152.45, 156.95, 159.42, 162.89, 163.74, 163.77, 165.82, 167.50, 169.10, 169.71, 172.22, 182.41, 193.86.

Antarvediside B^[31]:

C₄₆H₄₀O₁₈ (880.81)

White solid

R_f: 0.2 (7:3 ratio of hexane and ethyl acetate)

λ_{max}: 219 nm

Mp: 203-204°

Elemental analysis: C-62.76, H-4.54 (%)

ESI-MS m/z: 881.21 [M+H⁺]

¹H NMR (CDCl₃, 400 MHz): 13.39 (s, 2H), 12.55 (s, 1H), 12.50 (s, 1H), 11.95 (s, 1H), 10.36 (s, 1H), 6.67-6.69 (dd, 3H, J=8 Hz), 6.52 (s, 1H), 6.40 (s, 1H), 6.30-6.34 (dd, 3H, J=12, 16 Hz), 3.99 (s, 3H), 3.90 (s, 6H), 3.87 (s, 3H), 2.85 (s, 6H), 2.69 (s, 3H), 2.55 (s, 3H), 2.09 (s, 3H).

¹³C NMR (CDCl₃, 400 MHz): 9.39, 23.47, 24.04, 25.60, 29.73, 52.36, 55.67, 55.73, 62.14, 92.10, 96.79, 98.49, 100.44, 108.57, 109.00, 112.88, 115.47, 115.56, 116.03, 119.36, 143.50, 149.52, 152.46, 153.44, 162.90, 163.76, 164.17, 167.51, 168.72, 182.44, 185.78, 193.87.

Atranol^[32]:

C₈H₈O₃ (152.15)

Yellowish powder

R_f : 0.8 (7:3 ratio of dichloromethane and ethyl acetate)

λ_{\max} : 225 nm

Mp: 125-126°

Elemental analysis: C-63.50, H-5.52 (%)

ESI-MS m/z: 153.10 [M+H⁺]

¹H NMR (400 MHz, dimethylsulfoxide-d₆ (DMSO-d₆)): 11.29 (s, 1H), 10.66 (s, 1H), 9.94 (s, 1H), 6.12 (s, 2H), 2.23 (s, 3H).

¹³C NMR (400 MHz, DMSO-d₆): 22.46, 100.83, 107.93, 110.58, 141.15, 161.46, 192.87.

Atranorin^[31]:

C₁₉H₁₈O₈ (374.35)

Colorless needles

R_f : 0.4 (1:1 ratio of dichloromethane and ethyl acetate)

λ_{\max} : 210 nm

Mp: 196-197°

Elemental analysis: C-60.76, H-4.74 (%)

ESI-MS m/z: 375.11 [M+H⁺]

¹H NMR (CDCl₃, 400 MHz): 12.58 (s, 1H), 12.52 (s, 1H), 11.97 (s, 1H), 10.39 (s, 1H), 6.54 (s, 1H), 6.43 (s, 1H), 4.01 (s, 3H), 2.71 (s, 3H), 2.57 (s, 3H), 2.12 (s, 3H).

¹³C NMR (CDCl₃, 400 MHz): 9.37, 24.02, 25.58, 52.34, 102.85, 108.56, 110.27, 112.87, 116.02, 116.08, 139.88, 152.00, 152.44, 162.89, 167.50, 169.10, 169.71, 172.21, 193.36.

Chiodectonic acid^[33]:

C₁₅H₁₀O₉ (334.24)

Red color crystals

R_f : 0.4 (1:4 ratio of hexane and ethyl acetate)

λ_{\max} : 287 nm

Mp: 296-297°

Elemental analysis: C-60.76, H-4.74 (%)

ESI-MS m/z: 335.08 [M+H⁺]

¹H NMR (CDCl₃, 400 MHz): 5.84 (s, 1H), 5.63 (s, 1H), 5.15 (s, 1H), 3.79 (s, 3H), 2.69 (s, 3H), 1.41 (s, 1H).

¹³C NMR (CDCl₃, 400 MHz): 25.13, 61.34, 111.68, 115.55, 118.60, 129.90, 145.69, 146.38, 148.59, 155.94,

159.76, 158.46, 179.37, 184.07, 188.53.

Divaricatic acid^[31]:

C₂₁H₂₄O₇ (388.42)

Pale pink solid

R_f : 0.2 (1:1 ratio of dichloromethane and ethyl acetate)

λ_{\max} : 213 nm

Mp: 138-139°

Elemental analysis: C-64.76, H-6.34 (%)

ESI-MS m/z: 387.00 [M+H⁺]

¹H NMR (CDCl₃, 400 MHz): 11.36 (s, 2H), 6.78 (s, 1H), 6.67 (s, 1H), 6.40-6.42 (s, 2H), 3.85 (s, 3H), 2.96-3.04 (m, 4H), 1.70-1.72 (m, 4H), 0.97-1.05 (m, 6H).

¹³C NMR (CDCl₃, 400 MHz): 14.19, 14.23, 24.83, 25.28, 38.54, 39.18, 55.43, 99.03, 103.58, 108.50, 108.99, 111.59, 116.27, 148.11, 149.62, 154.98, 164.89, 165.35, 166.61, 169.39, 174.67.

Divarinol^[32]:

C₉H₁₂O₂ (152.19)

Pinkish powder

R_f : 0.8 (1:1 ratio of hexane and ethyl acetate)

λ_{\max} : 274 nm

Mp: 51-52°

Elemental analysis: C-71.87, H-7.82 (%)

ESI-MS m/z: 153.0 [M+H⁺]

¹H NMR (400 MHz, DMSO-d₆): 6.66 (s, 1H), 6.40-6.42 (dd, 1H, J=8, 9.2 Hz), 6.18 (s, 1H), 3.53 (brs, 2H), 2.95-3.04 (m, 2H), 1.65-1.76 (m, 2H), 0.97-1.04 (m, 3H).

¹³C NMR (400 MHz, DMSO-d₆): 12.40, 23.11, 35.21, 101.49, 108.59, 141.81, 155.76.

Divarinolmonomethylether^[32]:

C₁₀H₁₄O₂ (166.22)

Pale yellow oil

R_f : 0.6 (9:1 ratio of hexane and dichloromethane)

λ_{\max} : 273 nm

ESI-MS m/z: 166.90 [M+H⁺]

¹H NMR (400 MHz, DMSO-d₆): 6.61 (d, 1H, J=3 Hz), 6.49 (s, 1H), 6.24 (d, 1H, J=2.1 Hz), 4.39 (brs, 3H), 3.76 (brs, 1H), 2.66-2.74 (m, 2H), 1.41-1.52 (m, 2H), 0.74-0.82 (m, 3H).

^{13}C NMR (400 MHz, DMSO- d_6): 12.40, 23.11, 35.21, 52.81, 101.49, 108.59, 111.23, 141.81, 155.76, 162.12.

Ethyl divaricatinatate^[32]:

$\text{C}_{13}\text{H}_{18}\text{O}_4$ (238.28)

Sharp colorless needles

R_f : 0.4 (9:1 ratio of hexane and ethyl acetate)

λ_{max} : 274 nm

Mp: 51-52°

Elemental analysis: C-65.16, H-7.26 (%)

ESI-MS m/z: 239.0 $[\text{M}+\text{H}^+]$

^1H NMR (400 MHz, DMSO- d_6): 11.36 (s, 1H), 6.79 (d, 1H, $J=2.8$ Hz), 6.67 (d, 1H, $J=2.3$ Hz), 3.87 (s, 3H), 2.98-3.04 (m, 2H), 2.38-2.39 (m, 2H), 1.70-1.72 (m, 2H), 0.97-1.05 (m, 6H).

^{13}C NMR (400 MHz, DMSO- d_6): 14.50, 14.60, 24.87, 31.17, 55.75, 56.53, 99.46, 106.21, 108.59, 144.12, 162.92, 166.74, 172.65.

Ethyl everninate^[34]:

$\text{C}_{11}\text{H}_{14}\text{O}_4$ (210.23)

Greenish solid

R_f : 0.4 (9:1 ratio of hexane and dichloromethane)

λ_{max} : 219 nm

Mp: 74-76°

Elemental analysis: C-62.87; H-6.82 (%)

ESI-MS m/z: 210.0 $[\text{M}+\text{H}^+]$

^1H NMR (400 MHz, Acetone- d_6): 6.37 (d, 1H), 6.33 (d, 1H), 3.83 (s, 3H), 2.09 (s, 1H), 2.06 (t, 3H), 1.62 (m, 2H), 0.95 (t, 3H).

^{13}C NMR (400 MHz, Acetone- d_6): 14.47, 25.85, 54.96, 55.80, 99.70, 111.10, 149.10, 165.20, 167.20, 174.00.

Ethyl haematommate^[32]:

$\text{C}_{11}\text{H}_{12}\text{O}_5$ (224.21)

Greenish solid

R_f : 0.6 (7:3 ratio of dichloromethane and ethyl acetate)

Mp: 112-113°

λ_{max} : 209.5 nm

Elemental analysis: C-58.64, H-5.52 (%)

ESI-MS m/z: 224.90 $[\text{M}+\text{H}^+]$

^1H NMR (400 MHz, DMSO- d_6): 13.75 (s, 1H), 11.46 (s, 1H), 10.59 (s, 1H), 9.68 (s, 1H), 6.42 (s, 1H), 2.54 (s, 3H), 1.59-1.65 (m, 2H), 0.93-0.97 (t, 3H).

^{13}C NMR (400 MHz, DMSO- d_6): 14.28, 20.14, 68.63, 106.38, 109.34, 114.78, 155.22, 163.90, 167.04, 173.42, 191.73.

Ethyl orsellinate^[32]:

$\text{C}_{10}\text{H}_{12}\text{O}_4$ (196.20)

Pale yellow solid

R_f : 0.4 (7:3 ratio of dichloromethane and ethyl acetate)

λ_{max} : 219 nm

Mp: 131-132°

Elemental analysis: C-61.26, H-6.55 (%)

ESI-MS m/z: 197.10 $[\text{M}+\text{H}^+]$

^1H NMR (400 MHz, DMSO- d_6): 10.59 (s, 1H), 9.68 (s, 1H), 6.42 (s, 2H), 2.54 (s, 3H), 1.46-1.52 (m, 2H), 0.80-0.84 (m, 3H).

^{13}C NMR (400 MHz, DMSO- d_6): 14.28, 20.14, 68.63, 105.25, 109.34, 114.78, 155.22, 163.90, 173.42.

Graphenone^[33]:

$\text{C}_{14}\text{H}_{14}\text{O}_4$ (246.26)

Yellowish needles

R_f : 0.6 (1:1 ratio of hexane and ethyl acetate)

λ_{max} : 290 nm

Mp: 167-168°

Elemental analysis: C-68.71, H-5.72 (%)

ESI-MS m/z: 247.18 $[\text{M}+\text{H}^+]$

^1H NMR (CDCl_3 , 400 MHz): 7.05 (s, 1H), 6.43-6.53 (m, 5H), 2.34 (s, 3H), 2.12 (s, 3H), 1.93 (s, 3H).

^{13}C NMR (CDCl_3 , 400 MHz): 12.34, 12.61, 27.54, 125.00, 128.70, 129.02, 131.40, 131.97, 132.65, 134.73, 143.30, 166.28, 168.61, 200.83.

Graphisquinone^[33]:

$\text{C}_{11}\text{H}_{10}\text{O}_5$ (222.20)

Red needles

R_f : 0.4 (1:1 ratio of hexane and ethyl acetate)

λ_{max} : 228 nm

Mp: 130-131°

Elemental analysis: C-59.12, H-4.02 (%)

ESI-MS m/z: 223.21 [M+H⁺]

¹H NMR (CDCl₃, 400 MHz): 7.17 (s, 1H), 3.75 (s, 3H), 3.71 (s, 3H), 2.38 (s, 3H).

¹³C NMR (CDCl₃, 400 MHz): 14.37, 61.33, 104.36, 118.91, 144.94, 150.85, 153.57, 154.24, 176.98, 183.80.

Haematommic acid^[32]:

C₉H₈O₅ (196.16)

Pale yellow needles

R_f: 0.4 (1:1 ratio of hexane and ethyl acetate)

λ_{max}: 219.5 nm

Mp: 172-173°

Elemental analysis: C-55.64, H-4.52 (%)

ESI-MS m/z: 198.30 [M+H⁺]

¹H NMR (400 MHz, DMSO-d₆): 13.75 (s, 1H), 11.46 (s, 1H), 10.59 (s, 1H), 9.68 (s, 1H), 6.42 (s, 1H), 2.54 (s, 3H).

¹³C NMR (400 MHz, DMSO-d₆): 17.10, 105.25, 109.34, 155.22, 163.90, 167.04, 173.42, 191.73.

Methyl 2,6-dihydroxy-4-methylbenzoate^[32]:

C₉H₁₀O₄ (182.18)

Faint yellow crystals

R_f: 0.6 (1:1 ratio of hexane and ethyl acetate)

λ_{max}: 219.5 nm

Mp: 143-144°

Elemental analysis: C-59.36; H-5.52 (%)

ESI-MS m/z: 182.0 [M+H⁺]

¹H NMR (400 MHz, DMSO-d₆): 10.65 (s, 1H), 9.94 (s, 1H), 6.12 (d, 2H), 3.75 (s, 3H), 2.23 (s, 3H).

¹³C NMR (400 MHz, DMSO-d₆): 22.5, 52.2, 100.8, 107.9, 110.6, 141.2, 161.5, 170.6.

Orcinol^[32]:

C₇H₈O₂ (124.14)

Colorless needles

R_f: 0.7 (1:1 ratio of hexane and ethyl acetate)

λ_{max}: 215.5 nm

Mp: 108-109°

Elemental analysis: C-67.59, H-6.52 (%)

ESI-MS m/z: 125.10 [M+H⁺]

¹H NMR (400 MHz, DMSO-d₆): 6.30 (s, 1H), 6.23 (s, 2H), 3.34 (s, 2H), 2.46 (s, 3H).

¹³C NMR (400 MHz, DMSO-d₆): 27.93, 98.36, 109.34, 155.22, 163.90, 157.52.

Scrobiculin^[34]:

C₂₂H₂₆O₈ (418.44)

Colorless crystals

R_f: 0.6 (7:3 ratio of dichloromethane and ethyl acetate)

λ_{max}: 217.5 nm

Mp: 134-136°

Elemental analysis: C-63.16; H-6.26 (%)

ESI-MS m/z: 418.0 [M+H⁺]

¹H NMR (400 MHz, DMSO-d₆): 11.85 (s, 1H), 10.50 (s, 1H), 6.61 (s, 1H), 6.39-6.41 (dd, 2H), 3.84 (s, 3H), 3.78 (s, 3H), 2.82-2.87 (dd, 4H), 2.51 (t, 1H), 1.59-1.65 (dd, 4H), 0.88-0.97 (m, 6H).

¹³C NMR (400 MHz, DMSO-d₆): 14.5, 14.6, 24.8, 25.2, 36.9, 37.8, 55.8, 56.5, 99.5, 106.2, 108.6, 108.8, 109.5, 125.2, 144.1, 146.0, 154.3, 154.7, 160.6, 162.9, 166.7, 172.7.

Sekikaic acid^[31]:

C₂₂H₂₆O₈ (418.44)

White solid

R_f: 0.6 (1:1 ratio of dichloromethane and ethyl acetate)

λ_{max}: 219 nm

Mp: 220-221°

Elemental analysis: C-63.16, H-6.24 (%)

ESI-MS m/z: 419.33 [M+H⁺]

¹H NMR (CDCl₃, 400 MHz): 11.40 (s, 2H), 10.39 (s, 1H), 6.77-6.78 (d, J=3.8 Hz, 1H), 6.66-6.67 (d, J=3.2 Hz, 1H), 6.40-6.42 (dd, J=4, 8 Hz, 2H), 3.91 (s, 3H), 3.85 (s, 3H), 2.95-3.04 (m, 4H), 1.69-1.72 (m, 4H), 0.97-1.05 (m, 6H).

¹³C NMR (CDCl₃, 400 MHz): 174.34, 169.40, 166.60, 165.33, 164.88, 154.92, 149.53, 148.11, 116.22, 111.58, 108.55, 103.60, 99.03, 55.43, 39.18, 38.53, 25.28, 24.84, 14.23, 14.18.

Usnic acid^[34]:

C₁₈H₁₆O₇ (344.32)

TABLE 1: LIST OF SECONDARY METABOLITES IDENTIFIED FROM MANGLICOLOUS LICHENS

Manglicolous lichen	Isolated Metabolites
<i>Dirinaria consimilis</i> (Stirton) D. D. Awasthi ^[31]	2'-O-methyl divaricatic acid (1) Antarvediside A (2) Antarvediside B (3) Atranorin (4) Divaricatic acid (5) Sekikaic acid (6) Atranorin (4)
<i>Graphis ajarekarii</i> Patw. & C.R. Kulk ^[33,35,40]	Chiodectonic acid (7) Graphenone (8) Graphisquinone (9) Ethyl everninate (10)
<i>Parmotrema tinctorum</i> (Despr. ex Nyl.) ^[35]	2'-O-methyl divaricatic acid (1) Ethyl everninate (10)
<i>Ramalina leiodea</i> (Nyl.) Nyl. ^[34]	Methyl-2,6-dihydroxy-4-methyl benzoate (11) Scrobiculin (12) Usnic acid (13) Atranol (14) Divarinol (15) Divarinolmonomethylether (16)
<i>Rocella montagnei</i> Bel em. D.D. Awasthi ^[32]	Ethyl divaricatinatate (17) Ethyl haematommate (18) Ethyl orsellinate (19) Haematommic acid (20) Methyl-2,6-dihydroxy-4-methyl benzoate (11) Orcinol (21)

Yellowish needles

R_f: 0.4 (9:1 ratio of hexane and dichloromethane)

λ_{max}: 220 nm

Mp: 201-202°

Elemental analysis: C-63.16; H-6.26 (%)

ESI-MS m/z: 345.00 [M+H⁺]

¹H NMR (400 MHz, CDCl₃): 13.31 (s, 1H), 11.03 (s, 1H), 5.98 (s, 1H), 2.68 (s, 3H), 2.67 (s, 3H), 2.11 (s, 3H), 1.76 (s, 3H).

¹³C NMR (400 MHz, CDCl₃): 7.6, 27.9, 31.3, 32.2, 59.1, 98.4, 101.6, 104.0, 105.3, 109.3, 155.2, 157.5, 163.9, 179.4, 191.7, 198.1, 200.4, 201.8.

BIOLOGICAL EXAMINATION OF EXTRACTS AND SECONDARY METABOLITES OF MANGLICOLOUS LICHENS

Extracts of manglicolous lichens showed different biological properties, including antioxidant, antibacterial, anti-cancer, antifungal, anti-inflammatory, anti-tubercular, hypo and hyperglycemic activities^[35-39].

Ethyl acetate and acetone extracts of manglicolous lichen *Dirinaria consimilis* showed a reasonable specificity against human lung adenocarcinoma cell line (A549), human colon cancer cell line (DLD-1), a hypopharyngeal carcinoma cell line (FADU), immortal

cell line (HeLa) and breast cancer cell line (MCF-7)^[38,39]. Also, the acetone extract of manglicolous lichen *Dirinaria consimilis* revealed prominent inhibition of protein denaturation, as well as inflammation in rat paw edema^[39]. Ethanolic extract of manglicolous lichen *Graphis ajarekarii* showed fibrinolytic (*in vitro*) and anti-inflammatory activities (*in vitro* and *in vivo*) and a better specificity against human prostate cancer cell line (DU145), FADU, leukemia cell line (K-562), MCF-7, epithelial, human breast cancer cell line (MDA-MB-231) and human colon cancer cell line (SW620)^[33,40]. Additionally, ethyl acetate extract of mangrove associated lichen *Graphis ajarekarii* is active against *Bacillus subtilis*, *Micrococcus luteus*, *Salmonella Typhi*, *Candida albicans* and *Mycobacterium tuberculosis*, free radicals (2,2-diphenyl-1-picrylhydrazyl (DPPH), (2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid)) (ABTS) and superoxide) and also showed prominent hypo and hyperglycemic activities in albino rats^[35,36].

Manglicolous lichen *Parmotrema tinctorum* ethyl acetate extract is active against *S. Typhi*, free radicals (DPPH, ABTS and superoxide) and also showed prominent hypo and hyperglycemic activities in albino rats. Acetone extract of manglicolous lichen *P. tinctorum* is active against *B. subtilis* and *M. luteus*^[35,36]. Ethyl acetate and acetone extract of mangrove associated lichen *Ramalina leiodea* showed functional inhibition

of protein denaturation and a better specificity against A549, DLD-1, FADU, HeLa and MCF-7^[38]. Chloroform and methanol extracts of mangrove associated lichen *Roccella montagnei* were active against *S. aureus*, DLD-1 and MCF-7. Ethyl acetate extract of manglicolous lichen *R. montagnei* is active against *B. subtilis*. Acetone extract of mangrove associated lichen *R. montagnei* showed functional inhibition of free radicals (DPPH, ABTS and superoxide) and a significant specificity against A549, DLD-1, FADU and HeLa^[37]. List of secondary metabolites identified from manglicolous lichens is shown in Table 1.

It was reported that 1 and 3 were active against DPPH, ABTS and superoxide free radicals and pronounced specificity against A549, DLD-1, FADU, HeLa and MCF-7, while 2, 11, 16 and 18 were active against ABTS, DPPH and superoxide free radicals. 4, 18 and 20 were active against protein denaturation, whereas 5 and 6 reported having a noticeable specificity against DLD-1, FADU and HeLa. 7, 8, 9, 13, 15, 20 and 21 were active against DPPH and superoxide free radicals, while 7 and 9 reported having a prominent specificity against FADU, MDA-MB-231 and SW620. 10 was active against superoxide free radicals only and 12 was reported to have a distinct specificity against A549, DLD-1, FADU, HeLa and MCF-7. 14 was reported to be active against ABTS and superoxide free radicals. 17 was reported to have a noticeable specificity against MCF-7 and HeLa, while 18 and 20 reported having a prominent specificity against A549, FADU and HeLa. 19 and 20 were reported to have moderate active against *Mycobacterium tuberculosis* H37Rv strain, while 19 reported having a marked specificity against DLD-1^[31-35,40].

CONCLUSION

The chemical and biological investigation on lichens has exploited that they contain highly oxygenated aromatic derivatives with a wide range of biological activities. Also, more than 1000 secondary metabolites have been identified and reported from different lichen species, till now. However, there are no studies related to chemical and pharmacological examination on stressed lichens like manglicolous lichens. In many cases, it is proved that stressed organisms have a chance of obtaining novel compounds with potent biological activities.

Similarly, manglicolous lichens (stressed under mangrove ecosystems) are proved to have unique lichen substances with prominent pharmacological activities, which were listed out in this review. Moreover, as

there are very few isolations and biological evaluation reports on manglicolous lichens, there is still a need to carry out extensive studies on manglicolous lichens, to discover unexplored substances of lichens along with their pharmacological evaluation. Hence, this is a first review report on the chemical and pharmacological evaluation of manglicolous lichens, which assist academic researchers and pharmaceutical scientists in working not only on manglicolous lichens but also on stressed organisms.

Conflicts of Interest:

The authors declared no conflict of interest.

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