Comparison of Umbelliprenin and Auraptene in Cytotoxic Effects and Myeloid Cell Leukaemia Type-1 (Mcl-1) Gene Expression

F. MOVAHEDI MOTLAGH AND O. GHOLAMI1*

Cellular and Molecular Research Center, ¹Department of Physiology and Pharmacology, Faculty of Medicine, Sabzevar University of Medical Sciences, Sabzevar, Iran

Motlagh and Gholami: Comparison of Cytotoxic Effects of Umbelliprenin and Auraptene

Induction of apoptosis in tumour cells is one of the mechanisms of chemotherapy. Myeloid cell leukaemia type 1 is one of the Bcl-2 protein families that make an important role in chemo-resistance to drugs. Thus, down-regulation of myeloid cell leukaemia type 1 gene is one of the aims in chemotherapy. Umbelliprenin and auraptene are naturally occurring coumarins, which exhibit cytotoxic and apoptosis inducing capabilities. In this study, we compared the cytotoxicity of these two coumarins against some cancer cell lines by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay method. In the next step, we tested the effect of auraptene on myeloid cell leukaemia type 1 gene expression in Jurkat cells by real time polymerase chain reaction method. We found that umbelliprenin and auraptene possessed cytotoxic effect against the tested cancer cell lines. Auraptene is more cytotoxic than umbelliprenin. We also found that auraptene down-regulated myeloid cell leukaemia type 1 gene expression in a pattern that was different from umbelliprenin.

Key words: Umbelliprenin, auraptene, cytotoxicity, apoptosis, myeloid cell leukaemia type 1

Cancer is the uncontrolled growth of abnormal cells in the body. Chemotherapy and radiotherapy are highly effective methods of cancer treatment, but these methods exert severe side effects^[1]. Gradual resistance of cancer cells against these treatment is one of the main emerging problems in cancer treatment^[2]. In this regard, achieving a new approach to improve cancer treatment results is one of the aims of immunopharmacological studies^[3]. Apoptosis or programmed cell death is a strictly controlled pathway that is responsible for removal of unwanted cells, old and injured cells^[4]. The induction of apoptosis in tumour cells can lead to their own destruction, so apoptosis has been suggested as an efficient mechanism by which malignant tumour cells can be removed when treated with antineoplastic drugs^[5].

The B-cell lymphoma 2 (Bcl-2) protein family acts as a key regulator in the intrinsic or mitochondrial apoptosis pathway^[6]. Myeloid cell leukaemia type 1 (Mcl-1) is one of the most important members of the Bcl-2 protein family. Mcl-1 is highly expressed in a variety of human cancer cell lines including breast, CNS, colon, lung, ovarian, prostate, renal and melanoma^[7].

Overexpression of Mcl-1 gene is exploited by cancer cells to survive, and as a mechanism for developing resistance to diverse chemotherapeutic agents^[8]. There are multiple drugs that display a mechanism of action that involves down-regulation of Mcl-1. Studies with these drugs show the importance of this protein in maintenance and progression of the cancer phenotype^[8].

Many anticancer drugs are basically derived from herbal plants. Genus *Ferula* includes herbal plants that traditionally used as food and/or medicine in Iran and other Asian countries. Many *Ferula* species, including *F. asafetida*, synthesize terpenyloxy coumarins. Coumarins are a wide class of natural and synthetic compounds that showed variety of pharmacological activities including antiinflammatory^[8], antioxidant^[9], antinociceptive^[10], hepatoprotective^[11], antithrombotic^[12], antiviral^[13], antimicrobial^[14,15],

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antituberculosis^[16], anticarcinogenic^[17], antidepressant^[18], antihyperlipidemic^[19] and anticholinesterase^[20] activities.

Umbelliprenin and auraptene are terpenyloxy coumarins synthesized by *F. asafoetida* and citrus species. These 2 coumarins have similar structure, with the only difference being the higher length of the 7-prenyloxy chain, which contains 15 instead of 10 carbons^[21]. In the present study, we examined the cytotoxicity of umbelliprenin and auraptene on various cancerous cell lines by 3-(4,5-dimethylthiazol-2-yl)-2,5 diphenyltetrazolium bromide (MTT) method and compared those together. We also tested the effect of auraptene on Mcl-1 gene expression in Jurkat cells by real time polymerase chain reaction (PCR) method and compared it with the effect of umbelliprenin on gene expression.

MATERIAL AND METHODS

Cell lines were prepared from national cell bank of Iran (Pasteur institute, Tehran, Iran). HeLa (cervical cancer cell line), Jurkat (T cell leukaemia cell line), MCF-7 (breast cancer cell line) and KYSE-30 (oesophageal carcinoma cell line) were cultured in RPMI 1640 (Gibco[™], Cat. Number: 11875-093) and incubated at 95% humidity, with 5% CO₂ at 37° in the culture medium containing 10% foetal bovine serum (FBS, Gibco[™], Cat.Number:10082-147) and 50 U/ml penicillin/streptomycin (Sigma Aldrich, Cat. Number: 4458). After that cells were frozen in FBS containing 10% dimethyl sulfoxide (DMSO) and stored in liquid nitrogen (5×10⁶ cells/vial). The viability of cryopreserved cells was determined by trypan blue staining, immediately upon thawing. Only cells whose viability exceeded 93% (range of 93.4-99%) were used in this study. Umbelliprenin and auraptene preparations were obtained from Biotechnology Research Center and School of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran.

MTT assay:

The effects of auraptene and umbelliprenin on the growth of cell lines were examined using the MTT assay. Cells were sub-cultured in 96-well plates at a density of 10^4 cells (for MCF-7, HeLa, KYSE-30) or 5×10^4 cells (Jurkat) per well with or without auraptene and umbelliprenin (10, 20, 40 µg/ml) for 24 and 48 h in a final volume of 100 µl. Auraptene and umbelliprenin were dissolved in DMSO. Immediately before use, it was diluted in the culture medium to obtain a final

DMSO concentration of 0.5% (v/v). Then, the medium was changed with fresh medium supplemented with 20 μ l of MTT (5 mg/ml in phosphate-buffered saline (PBS)). Plates were incubated at 37° for 2 h. After addition of 100 μ l DMSO to each well, plates were agitated for 1 min. Spectrophotometric absorbance at 570 nm was measured. Cell viability was calculated using the formula. %viable cells=(OD of drug-treated sample/OD of untreated sample)×100.

RNA isolation and real-time quantitative PCR analysis:

Jurkat cells were incubated by auraptene (10, 20, 40 μ g/ml) in 37° and 5% CO₂ for 1, 2 and 3 h. Total RNA was isolated using RNX-Plus kit in accordance with the manufacturer's instructions (Sinagene[®]). The quantity and quality of the total RNA was verified with the Pico Drop spectrophotometer (Alpha Biotech, Cambridge, UK) according to the manufacturer's instructions. Complementary DNA (cDNA) was synthesized from 2 μ g total RNA treated with DNase I using TaKaRa kit (PrimeScript First Strand cDNA Synthesis Kit).

Real-time PCR was performed in 3 replicates in 20 µl reaction volumes using 0.2 µl cDNA, 10 µ SYBR Green master mix (YEKTA TAJHIZ AZMA, SYBR Green qPCR Master Mix 2x) and 0.4 µl of each primer on a Bio-Rad real time CR System. PCR was performed using Mcl-1 primers (5'-CCA AGA AAG CTG CAT CGA ACC AT-3' and 5'-CAG CAC ATT CCT GAT GCC ACC T-3') and GAPDH primers (5'-GGA AGG TGA AGG TCG GAG T-3' and 5'-GTC ATT GAT GGC AAC AAT ACC ACT-3') as internal control. Conditions of real-time PCR steps are presented in fig. 1.

Statistical analysis:

One way analysis of variance (ANOVA) test was used for statistical analysis by SPSS16 software. The P-value was considered significant when it was <0.05. Error bars indicate standard deviations. Number of readings (N) for MTT assay and real-time PCR were 3 replicate. Calculations were done with GraphPad Prism 5 (GraphPad Software, San Diego).

RESULTS AND DISCUSSION

For comparison of cytotoxicity of auraptene and umbelliprenin, we incubated various concentrations (10, 20 and 40 μ g/ml) of these compounds with HeLa (cervical cancer cell line), Jurkat (T cell leukaemia cell line), MCF-7 (breast cancer cell line) and KYSE-30 (oesophageal carcinoma cell line) for 24 and 48 h.

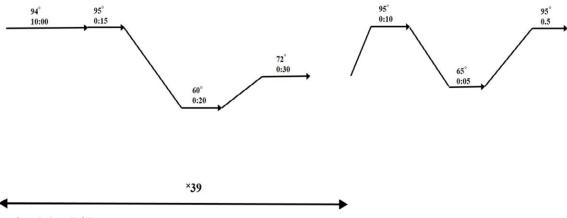


Fig. 1: Steps of real-time PCR.

After these incubation periods the extent of cytotoxicity is estimated using the MTT assay. Auraptene was more cytotoxic than umbelliprenin against these cell lines (figs. 2 and 3). IC_{50} of these two compounds have been compared in Table 1. As shown in Table 1 cytotoxic effect is dose and time dependent. Moreover, auraptene is more potent than umbelliprenin in various concentrations and times (Table 1).

Mcl-1 protein has an obvious role in the chemoresistance of certain malignancies^[22-24]. Forced Mcl-1 downregulation induce apoptosis in a number of cancer cell types^[25]. Thus, Mcl-1 is a potential and attractive therapeutic target in a number of malignancies and is the focus of a number of studies reviewed^[26]. In our study, we tested the effect of auraptene on Mcl-1 gene expression. We incubated Jurkat cells with 10, 20 and 40 µg/ml of auraptene for 1, 2 and 3 h. After that this effect was examined by real-time PCR method. 10 µg/ ml of auraptene did not have significant effect on Mcl-1 gene in other concentrations and times of incubation (fig. 4).

Cancer is one of the most complicated diseases of this century. Natural products are very promising compounds in treatment of cancer. Among natural products, coumarins show cytotoxic effect *in vitro* and *in vivo*. Based on previous findings we decided to compare cytotoxic effect of two natural coumarin compounds (auraptene and umbelliprenin) *in vitro*.

Auraptene and umbelliprenin belongs to the class of natural terpenyloxy coumarins. Umbelliprenin has a different structure from auraptene by the presence of an acyclic sesquiterpenene group in place of the geranyl group at C_7 -OH of the 1,2-benzopyrone ring. The cytotoxicity of auraptene and umbelliprenin has been found to be related to the presence of the

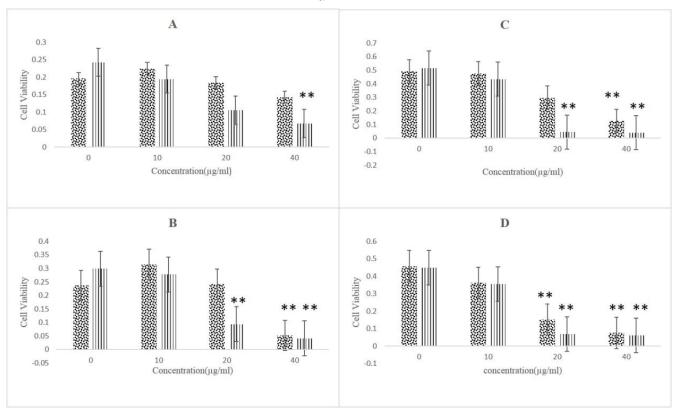
aliphatic sesquiterpenoid group linked at C_7 -OH. Cytotoxicity and apoptosis induction of auraptene and umbelliprenin on some of these cancerous cell lines were reported by other researchers^[5,27,28]. Since, none of them compared the cytotoxicity of these two compounds together, we decided to do this. We showed that auraptene and umbelliprenin have doseand time-dependent cytotoxicity against these cell lines. In general, auraptene is more potent (has less IC₅₀) than umbelliprenin in cytotoxic effects (Table 1). Umbelliprenin and auraptene have no cytotoxic effect on peripheral blood mononuclear cells (PBMCs) from healthy donors^[5,29].

Induction of apoptosis by auraptene and umbelliprenin in Jurkat cells have previously shown^[5,29]. These studies showed that umbelliprenin and auraptene, both of them, induced apoptosis in Jurkat cells by activation of caspase-8 (hallmark of extrinsic pathway of apoptosis) and caspase-9 (hallmark of intrinsic pathway of apoptosis)^[5,29]. In our review article, we compared proapoptotic properties of umbelliprenin and auraptene. We notified that umbelliprenin promoted G1 cell cycle arrest but auraptene promoted G1/S cell cycle arrest in cancerous cell lines. Moreover, we noticed that umbelliprenin decreased Bcl-2 and Mcl-1 gene expression but auraptene decreased Bcl-XL gene expression (Table 1)^[30].

One of the most important proteins that were upregulated in leukemic cells like Jurkat is Mcl-1 protein. We previously showed that Mcl-1 gene was upregulated by umbelliprenin (50 μ M \approx 20 μ g/ml) from 1 to 3 h incubation in Jurkat cells^[31]. In this study, we showed that Mcl-1 was down-regulated by auraptene (20 and 40 μ g/ml) from 1 to 3 h incubation in Jurkat cells.

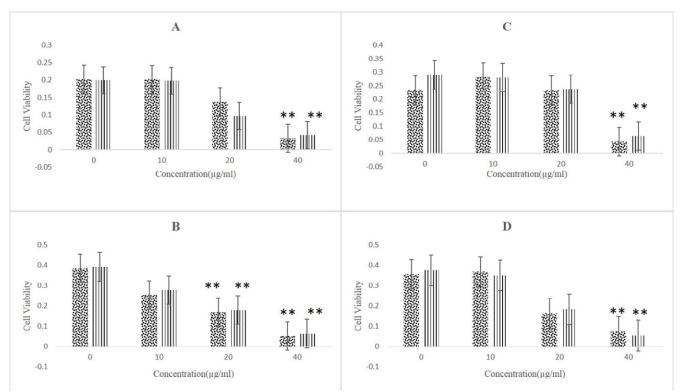
Umbelliprenin increased expression of Mcl-1 mRNA

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After 24 (A) and 48 h (B) incubation of HeLa cells and after 24 (C) and 48 h (D) incubation of Jurkat cells in the presence of either umbelliprenin or auraptene (0, 10, 20 and 40 µg/ml). Auraptene is more cytotoxic than umbelliprenin. Data are shown as mean±Standard deviation. **P<0.01 compared with control. umbelliprenin; || auraptene.

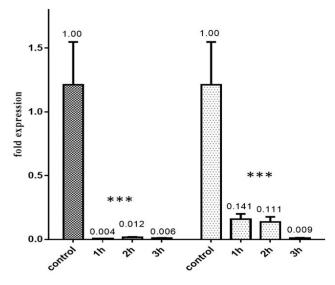




After 24 (A) and 48 h (B) incubation of KYSE-30 cells and after 24 (C) and 48 h (D) incubation of MCF-7 cells in the presence or absence of umbelliprenin and auraptene (0, 10, 20 and 40 µg/ml). Auraptene is more cytotoxic than umbelliprenin. Data are shown as mean±Standard deviation. **P<0.01 compared with control. Umbelliprenin; auraptene.

TABLE 1: COMPARING THE CYTOTOXICITY OF AURAPTENE AND UMBELLIPRENIN

Cell line	Compound	Time	IC ₅₀
HeLa	Aur	24 h	13.33
		48 h	13.87
	Umb	24 h	20.22
		48 h	27.26
Jurkat	Aur	24 h	11.3
		48 h	11.49
	Umb	24 h	19.64
		48 h	14.19
KYSE-30	Aur	24 h	11.75
		48 h	15.24
	Umb	24 h	29.51
		48 h	24.32
MCF-7	Aur	24 h	29.66
		48 h	17.26
	Umb	24 h	29.76
		48 h	15.10



Time after treated

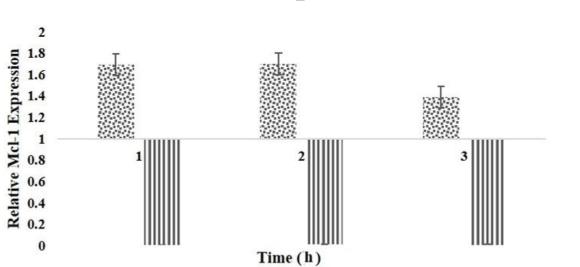
Fig. 4: Effect of auraptene on Mcl-1 gene expression after 1, 2 or 3 h incubation in Jurkat cells. Effect of auraptene (20 and 40 µg/ml) on Mcl-1 gene expression

after 1, 2 or 3 h incubation in Jurkat cells. Auraptene down-

regulated Mcl-1 gene expression at different concentrations

and times of incubation. ***P<0.001 compared with control.

Umbelliprenin and Auraptene (10, 20 and 40 µg/ml) were incubated with HeLa, Jurkat, MCF-7 and KYSE-30 cell lines for 24 and 48 h. Cytotoxicity was tested by MTT method. IC₅₀ was calculated for each cell line and time of incubation. As it shows cytotoxic effect is dose and time dependent. Auraptene is more cytotoxic than umbelliprenin in various cell lines and times (time and IC₅₀ have been indicated as h and µg/ml, respectively)



20; 40.

from 1 to 3 h incubation, but this increase has a scale down pattern^[31]. Auraptene decreased expression of Mcl-1 mRNA for same incubation times. In fig. 5, we compared the extent of Mcl-1 mRNA changes by umbelliprenin (50 μ M \approx 20 μ g/ml) and auraptene (20 μ g/ml) after 1, 2 and 3 h incubation.

Extent of Mcl-1 gene down-regulation by auraptene is different for various concentrations and times of incubation. This can be related to the different extent of apoptosis/necrosis induction by auraptene. As it shows in fig. 4, the extent of Mcl-1 gene down-regulation in 20 μ g/ml is more than 40 μ g/ml. This shows that auraptene induce apoptosis more than necrosis in 20 μ g/ml concentration. In 40 μ g/ml auraptene induce necrosis more than apoptosis. Our findings are consistent with those reported by June (fig. 3)^[5]. More studies are needed to get further clarity. Finally, it may

be concluded that umbelliprenin (but not auraptene) falls into the category of apoptosis inducing agents that initially up-regulates Mcl-1 gene.

It also appeared important to verify if auraptene and umbelliprenin promote anticoagulant effects and modulate immune functions. Anticoagulant effects may lead to side effects. However, auraptene and umbelliprenin may also contribute to decrease metastasis because interference with the fibrinolytic system affects angiogenesis. At least in one study, certain coumarins exhibiting anticoagulant effects inhibited metastasis in several animal models^[32].

In a review regarding the structure-activity relationships and anticancer effects of natural and synthetic coumarins, the author reported that coumarins may enhance lymphocyte response to mitogen and induce immune-induced antitumor effects. The author further suggested that the effects of auraptene (and possibly umbelliprenin) would partly be due to an enhancement of immune function^[32]. However, this stimulatory effect needed to be demonstrated.

In conclusion, we showed that umbelliprenin and auraptene have cytotoxic effects against some cancer cell lines. In this regard, auraptene was more cytotoxic than umbelliprenin against these cell lines. This study also demonstrated that auraptene down-regulated Mcl-1 mRNA expression in a pattern that was different from umbelliprenin

Since, auraptene has a shorter length 7-prenyloxy chain than umbelliprenin, it has more favourable pharmacokinetic properties than umbelliprenin. Moreover, auraptene is more cytotoxic than umbelliprenin against cancerous cell lines. In conclusion, auraptene is more attractive compound for future studies (for example *in vivo* studies) than umbelliprenin.

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Conflicts of interest:

There are no conflict of mentioned.

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