

Development and Validation of a Simultaneous HPLC Method for Estimation of Bisoprolol Fumarate and Amlodipine Besylate from Tablets

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Vora, *et al.*: Simultaneous HPLC Method for Bisoprolol Fumarate and Amlodipine Besylate

A fast, robust and stability indicating RP-HPLC method was developed for simultaneous determination of bisoprolol fumarate and amlodipine besylate in tablets. The mobile phase was mixture of 25 mM ammonium acetate adjusted to pH 5.0 and methanol (65: 35) at 0.8 ml/min. The stationary phase was Luna C18-2 column (3 μ , 50 \times 4.6 mm ID). UV detection was performed at 230 nm. Retention time was 1.45 min and 3.91 min for bisoprolol and amlodipine, respectively. Linearity was established in the range of 8–33 μ g/ml. Mean recovery was 99.1% and 98.6% for bisoprolol fumarate and amlodipine besylate, respectively.

Key words: Bisoprolol fumarate, amlodipine besylate, RP-HPLC

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Bisoprolol fumarate is a synthetic beta₁-selective cardioselective adrenoceptor blocking agent. The

chemical name for bisoprolol fumarate is (\pm)-1-[4-[[2-(1-methylethoxy) ethoxy] methyl]phenoxy]-3-[(1-methylethyl)amino]-2-propanol(*E*)-2-butenedioate (2:1). It is a white crystalline powder, which is readily soluble in water, methanol, ethanol, and chloroform¹. It is official in USP².

Amlodipine besylate, a long-acting calcium channel blocker, is chemically described as 3-ethyl-5-methyl(\pm)-2-[(2-aminoethoxy)methyl]-4-(2-chlorophenyl)-1,4-dihydro-6-methyl-3,5-pyridine dicarboxylate, monobenzenesulphonate. Amlodipine besylate is a white crystalline powder. It is slightly soluble in water and sparingly soluble in ethanol³. It is official in BP⁴. Beta blocker plus calcium channel blocker combinations have utility in certain cardiovascular diseases like angina pectoris, myocardial infarction and hypertension. A tablet formulation containing bisoprolol fumarate and amlodipine besylate has been recently introduced on the market.

Various methods for determination of bisoprolol by fluorimetry^{5,6}, HPLC⁷⁻⁹ and densitometry¹⁰⁻¹² are reported in literature. Also HPTLC¹³⁻¹⁶, HPLC¹⁷⁻²², spectrophotometry²³⁻²⁸ methods are reported for determination of amlodipine alone or in combination with other drugs. But, literature survey did not reveal any method for simultaneous determination of bisoprolol and amlodipine. The aim of this study was to develop a fast, precise, accurate, rugged and robust HPLC method for simultaneous determination of bisoprolol and amlodipine in tablets. Criteria employed for assessing suitability of proposed method was cost effectiveness and speed of analysis.

A liquid chromatographic system comprising of Waters 2695 separation module and Waters 2996 PDA detector (Waters Corporation, Milford, USA) connected to Empower chromatography software for processing the data generated were used. Reference standard of bisoprolol fumarate and amlodipine besylate was kindly supplied by Indoco Remedies along with certificate of analysis, and used as received. HPLC grade acetonitrile was purchased from J. T. Baker, NJ, USA, GR grade ammonium acetate was obtained from Merck, Mumbai, India and ExcelsaR grade glacial acetic acid was supplied by Qualigens Fine Chemicals, Mumbai, India. The filter used in sample preparation was mdi SY25NN which was manufactured by Advanced Microdevices (P) Ltd, Ambala, India. The combination tablets containing

bisoprolol fumarate and amlodipine besylate (Concor AM, Merck) were procured from the market.

A buffer solution was prepared by adjusting the pH of 25 mM ammonium acetate solution to 5.0 with acetic acid. The mobile phase was filtered and degassed mixture of buffer pH 5.0 and acetonitrile (65:35, v/v). Luna C18-2 column, (3 μ , 50 \times 4.6 mm) was used as stationary phase. A constant flow of 0.8 ml/min was maintained throughout the analysis. Detection was carried out using PDA detector at 230 nm.

A combined standard stock solution of bisoprolol fumarate and amlodipine besylate was prepared in methanol (200 μ g/ml). Five ml of standard stock solution was diluted to 50 ml with mobile phase to obtain a 20 μ g/ml solution of bisoprolol and amlodipine and used as working standard for assay analysis. Twenty tablets were weighed and crushed to fine powder. An accurately weighed portion of the powder equivalent to 10 mg of bisoprolol fumarate and 10 mg of amlodipine was taken in 50 ml volumetric flask, about 30 ml of methanol was added to it and flask was kept in an ultrasonic bath for 2 min with intermittent swirling. This solution was then diluted to the mark with methanol and centrifuged. Five ml of the supernatant solution was diluted to 50 ml with mobile phase and mixed. This solution was filtered through mdi SY25NN filter and used for assay analysis. Five μ l of each of working standard and sample solution were injected into the chromatograph and the peak areas were recorded. The amount of each active was computed by external standard quantification.

In order to optimize the LC separation of bisoprolol and amlodipine, initially, mobile phases of buffer and acetonitrile were used. The retention behavior of both the drugs was studied with respect to pH of buffer solution in the range of 3.0–6.8, and aqueous composition of mobile phase. Retention of both the drugs was found slightly dependant on pH of buffer (slight increase in retention with increase in pH). Bisoprolol was found relatively less sensitive to aqueous composition as against amlodipine, which was found more sensitive to aqueous composition. A ten percent increase in aqueous composition resulted in 1.6 and 3.3 times increase in retention for bisoprolol and amlodipine respectively. The buffer solution of pH 5.0 and mobile phase composition of buffer:acetonitrile (65:35) was found most appropriate

for separation of bisoprolol and amlodipine on Luna C18-2 column. Flow rate was optimized based on capacity factor and column efficiency. Bisoprolol and amlodipine were well resolved in reasonable time of about 5 minutes. The retention times were 1.45 min and 3.91 min, respectively. The resolution between bisoprolol and amlodipine was 14.2. The final dilution of analytes with mobile phase helped to minimize the interference due to blank peaks. The wavelength of 230 nm was selected for the UV detection because at this wavelength there was maximum overlap of the spectra of bisoprolol and amlodipine. The peak purity of the peak due to bisoprolol and amlodipine was tested using PDA detector and were found to be pure.

To ascertain effectiveness of system suitability test, five replicate injections of freshly prepared working standard solution were injected into the chromatograph and relative standard deviation (RSD) of peak areas was calculated. The data is presented in Table 1. System suitability parameters such as tailing factor, resolution factor, capacity factor and theoretical plates of a typical chromatogram are tabulated in Table 2. Linearity (described by equation and corresponding correlation coefficient) was determined using five calibration levels for both the compounds (at 50-150% levels). The concentrations of calibration solutions of both the drugs were from 8 to 33 µg/ml. The method of linear regression was used for data evaluation. Peak area of standard compounds was plotted against respective concentrations. The content of actives found in the commercial brand of tablets (Concor AM, Merck) by proposed method is shown in Table 1. The low values of RSD, indicates that method is precise.

Intermediate precision was studied using different column, HPLC instrument and performing the analysis on different day. The results are presented in Table 1, along with repeatability data. Sample solution injected after 24 h of preparation did not show any appreciable change in assay value. To confirm the accuracy of the proposed method, recovery experiments were carried out by standard addition technique. Three different levels of standards were added to pre-analyzed tablet samples in triplicate. The mean percentage recoveries of bisoprolol and amlodipine were 99.1% and 98.6%, respectively. The results are shown in Table 1, which indicates that the method is accurate and precise and also there is no interference due to excipients present in the tablets. To ascertain the suitability of filter used

TABLE 1: METHOD VALIDATION RESULTS FOR INDIVIDUAL COMPOUND

Parameter	Bisoprolol	Amlodipine
System precision ^a (% RSD)	0.5	0.8
Repeatability ^b (% assay)	98.5	99.9
Repeatability ^c (% RSD)	0.3	0.4
Intermediate precision ^b (% assay)	99.5	99.9
Intermediate precision ^c (% RSD)	0.5	0.4
Linearity ^d (correlation coefficient)	0.99999	0.99999
Linearity ^d (equation)	y = 8922.1x - 1466.6	y = 14126.9x - 5056.6
Accuracy ^e (% RSD)	0.8	0.4
Accuracy ^e (% recovery)	99.1	98.6
Selectivity ^f	No interference	No interference
Stability—ambient[%] ^g	99.5	98.7
Filter recovery ^h	99.8	99.6

^aDetermined on five replicate injections of working standard solution.

^bDetermined on six real samples of Concor AM tablets and average is reported.

^cPercent RSD of six values of % assay of Concor AM tablets. ^dDetermined at five

levels, from 50-150% of working standard concentration. ^eDetermined at three

levels with triplicate determination at each level. Mean of 9 values and % RSD

is reported. ^fDemonstrated by forced degradation and peak purity of main

peaks in degraded samples. ^gPercent correlation of assay after 24 h of sample

preparation against freshly prepared sample. ^hPercent correlation of assay of

filtered sample against assay of centrifuged sample.

TABLE 2: SYSTEM SUITABILITY PARAMETERS AND ROBUSTNESS

Component	Robustness parameter	k' ^a	T ^b	R ^c	N ^d	%Assay	
Bisoprolol	No change (repeatability)	1.08	1.48	-	2596	98.5	
	Organic in mobile phase (+3%)	1.20	1.38	-	1522	98.4	
	Organic in mobile phase (-3%)	1.31	1.39	-	1928	98.8	
	pH of buffer (+0.2 units)	1.18	1.50	-	2524	98.5	
	pH of buffer (-0.2 units)	1.14	1.41	-	2770	98.5	
	Column temperature 35°	1.31	1.39	-	3217	98.3	
	Flow (+0.1 mL)	1.05	1.52	-	2733	98.1	
	Flow (-0.1 mL)	1.20	1.43	-	2628	98.4	
	Amlodipine	No change (repeatability)	4.58	1.53	14.23	4985	99.9
		Organic in mobile phase (+3%)	3.79	1.54	9.23	3545	100.3
Organic in mobile phase (-3%)		5.25	1.52	13.26	4618	99.7	
pH of buffer (+ 0.2 units)		4.53	1.52	13.24	4835	99.8	
pH of buffer (- 0.2 units)		4.42	1.50	13.71	5062	99.8	
Column temperature 35°		4.96	1.51	14.61	5614	99.4	
Flow (+ 0.1 mL)		3.92	1.50	12.16	4738	99.1	
Flow (- 0.1 mL)		4.29	1.55	12.65	4933	99.6	

^aCapacity factor determined for individual peak. ^bTailing factor determined for individual peak. ^cResolution factor determined between bisoprolol and amlodipine peaks. ^dColumn efficiency expressed as number of theoretical plates for bisoprolol and amlodipine peaks.

TABLE 3: FORCED DEGRADATION DATA

Degradation condition	Bisoprolol			Amlodipine		
	% Assay	Purity angle ^a	Purity threshold	% Assay	Purity angle ^a	Purity threshold
No degradation (Control)	98.5	1.177	1.598	99.9	0.480	1.278
Acid hydrolysis (1N HCl, 80°, 20 min)	95.3	1.191	1.357	94.1	0.380	0.981
Alkali hydrolysis (1N NaOH, 80°, 10 min)	95.4	0.725	1.421	93.9	0.406	0.942
Oxidation (30% H ₂ O ₂ , 80°, 30 min)	91.5	1.327	1.478	91.6	0.417	0.974
Thermal (105°, 1 d)	79.7	3.501	26.787	84.6	1.126	18.860
Photolytic (UV@254 nm, 1 d)	93.9	1.791	29.484	98.5	0.845	20.924

^aFor the peak to be pure, purity angle is required to be less than purity threshold.

for filtering sample preparation, the aliquot of sample solution was centrifuged and another aliquot of same sample solution was filtered through SY25NN filter. The percentage assay result of filtered sample was in close agreement with result of centrifuged sample, indicating that there was no adsorption of analytes on the filter. In order to evaluate specificity and stability indicating capability of the proposed method forced degradation studies were performed. The powdered samples of tablets were exposed to acidic, alkaline, strong oxidizing, heat and UV light conditions. Also, standard of bisoprolol and amlodipine were exposed to the above stress conditions, individually and in combination with each other to identify source of degradation peaks, if any. All the exposed standards and tablet samples were then analyzed by the proposed method. The results are given in Table 3. The assay values found lowered for bisoprolol in all the degradation conditions, however in case of amlodipine the assay values found lowered in all conditions except light. The assay values of both the ingredients were found decreased significantly in case of samples exposed to strong heat. Peaks due to bisoprolol and amlodipine in the chromatogram of all exposed samples were investigated using PDA detector and were found spectrally pure. The proposed method was subjected to robustness studies with respect to change in pH of buffer (± 0.2 units), change in mobile phase composition ($\pm 3\%$), change in column temperature (35°) and change in flow rate (± 0.1 ml). The results are presented in Table 2. The method was found robust with respect to variability in above conditions.

The proposed method is fast, precise, accurate, rugged and robust for the simultaneous determination of bisoprolol and amlodipine from tablets. Hence it can be easily and conveniently adopted for the routine quality control analysis for assay as well as dissolution and content uniformity testing.

REFERENCES

1. Available from: <http://www.rxlist.com/cgi/generic3/bisoprolol.htm>. October 2006.
2. The United States Pharmacopoeia, 29th ed. Rockwell, MD: The United States Pharmacopoeial Convention, Inc.; 2006.
3. Available from: <http://www.rxlist.com/cgi/generic/amlod2.htm>. October 2006.
4. The British Pharmacopoeia, London: British Pharmacopoeial Commission; 2005.
5. Yang XM, Wang CB. Determination of bisoprolol in Urine by fluorometry. Fenxi Shiyanshi (Chinese) 2001;20:54-5.
6. Braza AJ, Modamio P, Lastra CF, Marino EL. Development validation and analytical error function of two chromatographic methods with fluorimetric detection for the determination of bisoprolol and metoprolol, in human plasma. Biomed Chromatogr 2002;16:517-22.
7. Agapova NN, Vasileva E. HPLC method for determination of bisoprolol and potential impurities. J Chromatogr A 1993;654:299-302.
8. Kintz P, Lohner S, Tracqui A, Mangin P, Lugnier A, Chaumont AJ. Rapid HPLC determination of bisoprolol in human plasma. J Anal Chem 1990;336:517-9.
9. Buehring KU, Garbe A. Determination of the new beta blocker bisoprolol and of metoprolol, atenolol and propranolol in plasma and urine by HPLC. J Chrom Biomed Appl 1986;55:215-24.
10. Krzek J, Kwiecien A. Application of densitometry for determination of beta-adrenergic-blocking agents in pharmaceutical preparations. J Planar Chromatogr Mod TLC 2005;18:308-13.
11. Witek A, Hopkala H, Matysik G. TLC densitometric determination of bisoprolol labetalol and propafenone as dabsyl derivatives in pharmaceutical preparations. Chromatographia 1999;50:41-4.
12. Witek A, Hopkala H, Przyborowski L. Chromatographic separation of bisoprolol labetalol and propafenone, in the form of dabsyl derivatives. Chemia Analityczna (Warsaw) 1998;43:817-22.
13. Meyyanathan SN, Suresh B. HPTLC method for the simultaneous determination of amlodipine and benazepril in their formulations. J Chromatogr Sci 2005;43:73-5.
14. Gawri N, Vaidhyalingam V, Santha A. HPTLC method for the simultaneous estimation of amlodipine besylate and benazepril HCl tablets. Indian Drugs 2003;40:645-8.
15. Ilango K, Kumar PB, Lakshmi KS. Simple and rapid HPTLC estimation of amlodipine and atenolol from pharmaceutical dosages. Indian Drugs 2000;37:497-9.
16. Argekar AP, Powar SG. Simultaneous determination of atenolol and amlodipine in tablets by HPTLC. J Pharm Biomed Anal 2000;21:1137-42.
17. Rao JR, Kadam SS, Mahadik KR. Reverse phase HPLC determination of amlodipine and benazepril HCl in tablets. Indian Drugs 2002;39:378-81.
18. Gowri N, Vidhyalingam V, Santha A. Simultaneous estimation of amlodipine and benazepril from tablet by RP- HPLC. Indian Drugs 2001;38:332-5.
19. Zarakar SS, Kanyawar NS. Simultaneous estimation of amlodipine and losartan potassium in pharmaceutical dosage by RP-HPLC. Indian

- Drugs 2002;39:341-3.
20. Zarpkar SS, Kolte SS, Rane SH. High performance liquid chromatographic determination of amlodipine and atenolol simultaneously from pharmaceutical preparations. *Indian Drugs* 1997;34:350-3.
 21. Dhorda UJ, Shetkar NB. Reverse phase HPLC determination of ramipril and amlodipine in tablets. *Indian Drugs* 1999;36:638-41.
 22. Halkar UP, Bhandari NP, Rane SH. High performance liquid chromatographic Simultaneous determination of amlodipine and enalapril maleate from pharmaceutical preparations. *Indian Drugs* 1998;35:168-9.
 23. Meyya SN, Nathan GV, Ramasarma, Suresh B. Simultaneous spectrophotometric estimation of benazepril and amlodipine besylate in their dosage form. *Indian Pharmacist* 2003;2:100-1.
 24. Prasad CVN, Parihar C, Chowdhary TR, Purohit S, Parimoo P. Simultaneous determination of atenolol-amlodipine and haloperidol-trihexyphenidyl in combined tablet preparations by derivative spectroscopy. *Pharm Pharmacol Comm* 1998;4:325-30.
 25. Prasad CVN, Saha RN, Parimoo P. Simultaneous determination of amlodipine-enalapril maleate and amlodipine-lisinopril in combined tablet preparations by derivative spectrophotometry. *Pharm Pharmacol Comm* 1999;5:383-8.
 26. Dake AS, Kasture VS and Syed MR. Spectrophotometric estimation of amlodipine besylate and enalapril maleate in tablet. *Indian Drugs* 2002;39:14-7.
 27. Jain HK, Agrawal RK. Spectrophotometric method for simultaneous estimation of amlodipine besylate and lisinopril in tablet. *Indian Drugs* 2000;37:196-99.
 28. Mashru RC, Parikh PP. Development of method of simultaneous estimation of amlodipine besytale and lisinopril in their in combined dosage form. *East Pharm* 2000;43:111-2.

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