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Development and Validation of HPTLC Method for the Estimation of Almotriptan Malate in Tablet Dosage Form

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Suneetha and Syamasundar: HPTLC Method of Almotriptan Malate

A new, simple, precise and accurate high performance thin layer chromatographic method has been proposed for the determination of almotriptan malate in a tablet dosage form. The drug was separated on aluminum plates precoated with silica gel 60 GF₂₅₄ with butanol:acetic acid:water (3:1:1) was used as mobilephase. Quantitative analysis was performed by densitometric scanning at 300 nm. The method was validated for linearity, accuracy, precision and robustness. The calibration plot was linear over the range of 100-700 ng/band for almotriptan malate. The method was successfully applied to the analysis of drug in a pharmaceutical dosage form.

Key words: Almotriptan malate, high-performance thin-layer chromatography, tablets

Almotriptan malate^[1], a selective 5-hydroxytryptamine 1B/1D (5-HT1B/1D) receptor agonist, is chemically designated as

*Address for correspondence E-mail: profbsyamsundar@yahoo.co.in 1-[[[3-[2-(dimethylamino) ethyl]-1H-indol-5-1]methyl] sulfonyl] pyrrolidine-(\pm)-hydroxy butanedioate (1:1), whose empirical formula is C₁₇H₂₅N₃O₂S-C₄H₆O₅, with a molecular weight of 469.56. Almotriptan malate is a water soluble white to slightly yellow crystalline powder, which is used to treat severe

migraine headaches and is available as Axert tablets. Forner et al.^[2] and Bosch et al.^[3] have synthesised almotriptan malate and have published the data in US5565447. A few procedures based on liquid chromatography have been reported for the quantitative determination of almotriptan in human plasma and urine. A HPLC method for the determination of process-related impurities in almotriptan malate was reported earlier^[4]. Jansat *et* al.^[5] have published a method based on HPLC for determining the levels of almotriptan in plasma. Fleishaker et al.^[6] have reported a validated HPLC method for the determination of almotriptan concentrations in urine. Literature survey revealed that no analytical method for determination of almotriptan malate in pharmaceutical dosage forms has been reported thus far. This paper describes a simple, accurate, precise, and sensitive HPTLC method for determination of almotriptan malate in tablet dosage form. The proposed method was optimized and validated in accordance with guidelines suggested by International Conference on Harmonization (ICH)^[7].

Methanol, butanol, toluene, ethyl acetate, acetic acid, hexane, acetone and dichloromethane (all are of Analytical Reagent grade) were obtained from Sisco Research Laboratories, Mumbai, India. Standard bulk drug sample of almotriptan malate (99.8% pure) was obtained as a gift sample from SMS Pharmaceuticals, Hyderabad, India. The pharmaceutical dosage form used in this study was Axert tablets with a declared content of 12.5 mg almotripan malate (Ortho McNell, USA)

A standard stock solution of almotriptan malate was prepared by dissolving 10 mg drug in 100 ml methanol to furnish a concentration of 100 µg/ ml. Chromatography was performed on 10×10 cm aluminum plates precoated with 250-µm layers of silica gel 60 GF₂₅₄ (E. Merck, Darmstadt, Germany). Before use the plates were prewashed with methanol and activated at 110° for 5 min. Samples were applied to the plates as bands 6 mm wide and 10 mm apart by means of a Camag (Switzerland) Linomat V sample applicator equipped with a 100 µl syringe (Hamilton, Bonaduz, Switzerland). Linear ascending development was performed in a 10×10 cm twin trough glass chamber (Camag), with butanol-acetic acid-water 3:1:1 (v/v) as mobile phase and the chamber was presaturated with mobile phase vapour

for 10 min. The development distance was 8.5 cm and the development time was approximately 60 min.

After chromatography the plates were dried in a current of air with the use of a air drier. Densitometric scanning was performed with a Camag TLC Scanner 3 at 300 nm for all measurements. The scanner was operated by Wincats software version 1.2.3. The source of radiation was a deuterium lamp emitting a continuous UV spectrum between 200 and 400 nm. The slit dimensions were 5 mm \times 0.45 mm and the scanning speed was 20 mm/s.

After chromatographic development, bands were scanned over the range 200-400 nm (spectrum scan speed 20 nm/s) so that the drug could be estimated at 300 nm fig. 1, which is ascertained by taking the spectrum at different concentrations between 100–700 ng with 100 ng increment. Further it is also observed that spectra are similar in their behavior.

The standard stock solution of almotriptan malate (100 μ g/ml) was applied on a TLC plate, in the range 1–7 μ l, by use of the Linomat V sample applicator and 100 μ l syringe. The plate was developed and scanned under the conditions described above. Each amount was analyzed five times and peak areas were recorded. A calibration plot of peak area against respective amount was established for almotriptan malate.

Twenty tablets were weighed accurately and finely



Fig. 1: Overlain spectra of almotriptan malate from 200 to 400 nm, recorded *in situ* on the plate --**■**--100 ng/band; --×--200 ng/band; ----300 ng/band;--●-- 400 ng/band; --△--500 ng/band; --*--600 ng/band;--+-- 700 ng/band

powdered. A quantity of powder equivalent to 10 mg almotriptan malate was weighed and transferred to a standard 100 ml volumetric flask containing approximately 50 ml methanol. The mixture was ultra sonicated for 5 min then diluted to volume with methanol. The solution was filtered using Whatmann 41 filter paper. One to seven microlitres of the filtrate was applied to a TLC plate. After development of chromatogram the peak area of the bands were measured at 300 nm and the amount of drug in each tablet was determined from the calibration plot. The analytical procedure was repeated six times for the homogenous powder sample.

The limit of detection of a compound is defined as the smallest level of analyte that gives a measurable response. LOD was determined by using the equation $LOD= 3.3 \times \sigma/S$. The limit of quantitation is the lowest concentration of a compound that can be quantified with acceptable precision and accuracy. LOQ was determined by employing the relationship $LOQ=10\times\sigma/S$, where σ = the standard deviation of the response and S = the slope of the calibration curve.

To study intra-day and inter-day precision, three different concentrations of sample solutions were prepared and applied to the TLC plates. All the solutions were analyzed in triplicate on the same day and on three different days to record intra-day and inter-day variations in the results, respectively.

To check the accuracy of the method, recovery was measured by the addition of standard drug solution at three different levels (50, 100 and 150%) to preanalyzed sample solution (200 ng/band for almotriptan malate was selected so that after standard addition sample would be in the linear range). Three replicate estimations were carried out for each concentration level.

The effect of small, deliberate variation of the analytical conditions on the peak areas of the drugs was examined in order to assess the robustness of the method. The robustness of the method was checked for 200 and 700 ng/band for almotriptan malate.

For optimization of method, different mobile phase compositions were employed to achieve good separation. The method development was initiated by using different proportions of mobile phase consisting variety of solvents like: toluene:methanol:ethylacetate,

TABLE 1: ANALYSIS OF MARKETED FORMULATION OF ALMOTRIPTAN MALATE BY HPTLC METHOD

			70	
Almotriptan malate	12.5 mg	12.42 mg	99.36	0.52

*Average of six determinations

TABLE 2: RESULT FROM RECOVERY STUDIES OF ALMOTRIPTAN MALATE

Sample	Excess of drug added to the analyte (%)	Theoretical content (ng)	Recovery* %	RSD %
Almotriptan	50	300	99.54	0.21
malate	100	400	99.82	0.35
	150	500	101.1	0.61

*Average of three determinations

Parameters	Results of almotriptan malate		
Linearity (ng/band)	100-700		
Correlation coefficient	0.9994		
LOD (ng/band)	28.19		
LOQ (ng/band)	93.99		
Precision			
Interday (% RSD)	0.51		
Intraday (% RSD)	0.23		
Specificity	specific		



Fig. 2: Densitogram obtained from a sample solution of almotriptan malate

dichloromethane:methanol, butanol:aceticacid:water, toluene:methanol:acetone, toluene:methanol:ethyla cetate:aceticacid, hexane:ethylacetate. Of these the mixture butanol-acetic acid-water 3:1:1 (v/v) was found to be suitable for the studies. The R_F value of almotriptan malate was 0.53±0.01. The proposed HPTLC method was validated in terms of linearity, precision, accuracy, specificity and robustness. The calibration plot was found to be linear over the range 100–700 ng/band for almotriptan malate, with a correlation coefficient of 0.9994±0.0002. The LOD

and LOQ were found to be 28.19 and 93.99 ng/ band, respectively. The values of percent relative standard deviations were found to be 0.23 and 0.51 for Intraday and inter day precision studies, respectively which indicate that the method is precise. The method was also evaluated by assay of commercially available tablets (Axert) containing almotriptan malate. The resulting densitogram for a sample solution of almotriptan malate is presented in fig. 2 indicating a concentration of 700 ng/band. Six replicate analyses were performed on accurately weighed amount of the tablets (Table 1). The percent assay was found to be 99.36±0.522 for almotriptan malate. To study the accuracy of the method, recovery studies were performed. For almotriptan malate, recovery ranged from 99.54 to 101.1% with values of percent RSD ranging from 0.21 to 0.61 indicating that the proposed HPTLC method is highly accurate (Table 2). To confirm the specificity of the proposed method, the solution of formulation was spotted on TLC plate, developed and scanned. It was observed that the excipients present in the formulation did not interfere with the sample peak. Study of the robustness of the method revealed that the peak areas were unaffected (RSD < 2%) by small changes in composition and volume of mobile phase indicating appreciable robustness of the method. The method validation parameters are presented in Table 3.

The validated HPTLC method for almotriptan malate is simple, rapid, accurate, precise, sensitive, specific and robust and can thus be used for routine analysis of almotriptan malate in a tablet dosage form.

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REFERENCES

- 1. Maryadele JO, Heckelman PE, Koch CB, Roman KJ. Editors. The Merck Index: an Encyclopedia of Chemicals, Drugs and Biologicals. 14th ed. Whitehouse Station NJ: Merck and Co Inc; 2006.
- 2. Forner DF, Duran CP, Soto JP, Noverola AV, Maruti JM. Indole derivaties. US Patent 5565447. 1996.
- 3. Bosch J, Roca T, Armengol M, Forner DF. Synthesis of 5-(Sulfamoyl methyl) indoles. Tetrahedron 2001;57:1041-48.
- Kumar AP, Ganesh VR, Rao DV, Anil C, Rao BV, Hariharakrishnan VS, *et al*. A Validated Reverse phase HPLC method for the determination of process related impurities in almotriptan malate API. J Pharm Biomed Anal 2008;46:792-98.
- Jansat JM, Tobed AM, Garcia E, Cabarrocas X, Costa J. Effect of food intake on the bioavilability of almotriptan, an anti migrane compounds in healthy volunteers: An open, randomized, crossover, single-dose clinical trial. Int J Clin Pharmacol Ther 2006;44:185-90.
- Fleishaker JC, Ryan KK, Jansat JM, Carel BJ, Bell DJ, Burke MT, *et al.* Effect of MAO-A inhibition on the pharmacokinetics of almotriptan, an anti migrane agents in humans. Br J Clin Pharmacol 2001;51:437-41.
- International Conference on Harmonization (2005) ICH harmonized tripartite guidelines (Nov 2005), Validation of Analytical Procedures: Text and Methodology Q2 (R1).ICH, Geneva.

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