Effects of *Allium Hookeri* Root Extracts on Oxidative Stress-induced Damage in Rat Cardiomyocyte Cells

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The aim of this study was to investigate the bioavailability of *Allium hookeri* root extracts and evaluate its beneficial effects on hydrogen peroxide (H_2O_2) -induced oxidative stress in cultured H9C2 cardiomyocytes. After H9C2 cardiomyocytes were treated with the AHE, a substantial decrease in the H_2O_2 -induced cytotoxicity was observed in a dose dependent manner. These results suggest that AHE has beneficial effects on cardiac dysfunction which can be used as a valuable supplement in the health-care products.

Key words: Allium hookeri, oxidative stress, cardiomyocyte, H9C2 cells

Allium hookeri is a plant species that is native to south Asian countries including India, Burma, Sri Lanka and South western China^[1]. Recently, the plant is successfully cultivated in other countries of South and Southeast Asia such as Korea where it is valued as a food supplement. Furthermore, this plant and/or its extracts are anticipated to have ameliorative properties to several human diseases^[2] such as diabetes, other metabolic syndromes and allergy^[3,4]. However, no reports are available about its effectiveness in complicated disorders of diabetes.

A. hookeri is one of the new imported plant species in Korea where it is known with its common names such as Hooker chives, *SamChe* (in Korea) and is used as an alternative food additive in many countries. However, the effects of this plant, especially the roots (fig. 1) and its extracts on oxidative stress in heart are not recognized so far.

In the present study, the decrease in hydrogen peroxide-induced cytotoxicity was ascertained in rat neonatal cardiomyocytes (H9C2) after the administration of *A. hookeri* roots extracts to determine the ameliorative effect of plants and its extracts on circulation disorders^[5-7]. Then, we examined the ameliorative effects of water extracts of *A. hookeri* (AHE) on hydrogen peroxide-induced cardiomyocytes damaged by cytotoxicity test.

To obtain the rat heart myoblast cell line, H9C2 cells were purchased from the Korean Cell Line Bank (KCLB, Seoul, Korea). All chemical reagents were



Fig. 1: Representative photograph of Allium hookeri roots. These fresh roots were obtained from Hyejeong Farm (Bosung, Korea)

obtained from Sigma-Aldrich. Initially, roots of *A. hookeri* obtained from the Hyejeong Farm (Bosung, Korea) were thoroughly washed with water and freeze dried. For extraction of the sample, 100 g of dried samples were homogenized with distilled water by a Waring blender (HMF-1710; Hanil) to 2-4mm particle size. The freeze-dried sample (moisture content: <5%) was stored in a -80° freezer for further study. To determine the cell viability, EZ-Cytox Cell Viability Assay Kit (Daeil Labservice, Korea) or MTT

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assay was used^[8]. Cells were inoculated overnight in a 96 wells plate in CO₂ incubator and further incubated with various concentration of extracts or H_2O_2 (Sigma) for 24 h. After treatment, cells were coincubated with 10 µl of Ez-CyTox solution for 20 min in a CO₂ incubator. Then, absorbance measurements and calculation of the proliferation rates were carried out according to the manufactures instructions. Data was demonstrated by relative growth inhibition to phosphate buffered saline (PBS, pH 7.4) treated cells. All data were expressed as mean±SEM. Statistical comparisons were performed using oneway ANOVA test. Significant differences between groups were determined using the unpaired Student's t test. Statistical significance was set at P<0.05.

We defined optimum concentration of H_2O_2 by cytotoxicity assay on H9C2 cardiomyocytes model (fig. 2a). The concentration of H_2O_2 that liberates half of the viability (50% viability) as compared to the non-treated group was considered as optimum concentration. A significant decrease in the cell's viability was observed in a dose dependent manner at optimum concentration (0.5 mM, v/v) of H_2O_2 . To determine endogenous cytotoxicity of AHE on H9C2 cells, we measured cell viability with various concentration of AHE for 24 h (0 to 2.0 mg/ml, w/v, fig. 2b). However, no increase in cytotoxicity was observed at all concentrations of AHE, nonetheless

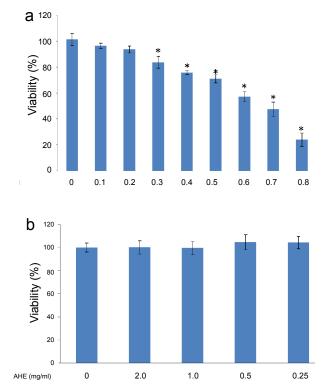


Fig. 2: MTT assay on H9C2 cells.

(A) H_2O_2 -induced cytotoxicity in a dose dependent manner on H9C2 cells and (B) endogeneous cytotoxicity of AHE. H_2O_2 and AHE were incubated with H9C2 cells for 24 h in CO₂ incubator then viable cells were quantified by MTT assay. Data are shown as Mean±SD, *P<0.05 versus no-treated group

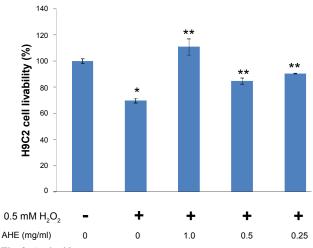


Fig. 3: Antioxidant assay.

Antioxidative stress of water extracts of Allium hookeri. H_2O_2 and extracts (or PBS) were incubated with H9C2 cells for 20 min in CO_2 incubator. Data are shown as Mean±SD, *P<0.05 versus normal group; **P<0.05 versus H_2O_2 treated group

we did not use more high concentration.

The reducing effects of AHE on H_2O_2 -induced H9C2 cell death were compared with that of the normal and the control groups. As shown in fig. 3, the cell viability of the H_2O_2 treated group was markedly decreased than normal group but AHE treated group recovered the cell viabilities at all concentrations. Although, relative high concentration AHE (2 mg/ml, w/v) shows reducing effects of H_2O_2 -induced cell death and serial diluted AHE shows similar effects. These results suggest that AHE has the potential to reduce the risk of heart problem such as hypertensive heart disease and diabetic cardiovascular diseases.

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CONFLICT OF INTERESTS

The author(s) declare(s) that there is no conflict of interest regarding the publication of this paper.

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