In Situ Single-Pass Intestinal Perfusion in Rats for Intestinal Permeability Investigation of Drugs

WEIYIN HUANG¹, SHUANG CHEN¹, LING SUN², H. WANG² AND HONGQUN QIAO^{1*}

Department of Pharmacy, Zhejiang Provincial People's Hospital Bijie Hospital, Bijie 551700, ¹School of Pharmaceutical Sciences, Nanjing Tech University, ²Jiangsu Provincial Institute of Materia Medica, Nanjing 211816, PR China

Huang et al.: Intestinal Permeability Investigation of Drugs

The aim of this study was to establish the *in situ* single-pass intestinal perfusion method in rats for intestinal permeability investigation of drug. Effective permeability coefficients (P_{off}) of 15 model drugs were investigated in rat jejunal. Krebs-Ringer buffer solution containing drug was perfused at flow rate of 0.2 ml/min and samples were taken from outlet up to 110 min, steady state was achieved after 30 min perfusion without samples taken. Gravimetric method was used to correct the net water flux. Drug concentrations in perfusion samples were determined using high-performance liquid chromatography-ultraviolet method and P_{att} were calculated. The P_{eff} values obtained by *in situ* single-pass intestinal perfusion ranged from 0.0146±0.0026×10⁻⁴ cm/s of enalaprilat to $1.77\pm0.34\times10^{-4}$ cm/s of antipyrine. Among the selected model drugs, the P_{eff} values of high permeability drugs were higher than 0.2×10^{-4} cm/s, and less than 0.03×10^{-4} cm/s for low permeability drugs. The experimental rat Peff was highly correlated to the literature human Peff with correlation coefficient (R²) of 0.99. The observed experimental rat P_{eff} values were highly correlated to the literature human F_{abs} (R²=0.93). All investigated model drugs have similar permeability classification compare with literature permeability classification. The predicted human F_{abs} were linearly correlated with literature human F_{abs} (R²=0.90). This method could be used for investigation of the intestinal absorption of new compounds, and for prediction the absorption fraction in human. Besides, investigation of the permeability classification of generic drug by in situ single-pass intestinal perfusion can be used for application of biowaiver for industry.

Key words: Biopharmaceutics classification system, intestinal permeability, *in situ* single-pass intestinal perfusion, effective permeability coefficient

The Biopharmaceutics Classification System (BCS), first proposed by Amidon et al.^[1] in 1995, is a scientific frame for classifying drugs according to solubility of the drug dose in the Gastrointestinal (GI) milieu and permeability of the drug through the GI membrane. Drugs are divided into 4 categories according to BCS: BCS class I (high solubility-high permeability), BCS class II (low solubility-high permeability), BCS class III (high solubility-low permeability) and BCS class IV (low solubility-low permeability). Before the BCS classification system was proposed, it was difficult for industry to find theoretical support to apply biowaiver from drug regulatory department. Based on this frame, when an immediate-release solid oral dosage form drug with a rapid dissolution compare to gastric emptying and the drug has high solubility, the absorption of drug is most likely to be dependent on the permeability of drug on intestine membrane. Therefore, it is not necessary for drug products containing BCS I and BCS III active ingredients to prove the *in vivo* bioavailability and bioequivalence, as long as the absorption of active ingredients was not significantly affected by the excipients used in the dosage form^[2,3].

Solubility parameters of drugs are readily available compared to permeability parameters. The model for evaluating permeability parameters of drugs

Accepted 11 September 2023 Revised 18 July 2023 Received 09 October 2022 Indian J Pharm Sci 2023;85(5):1429-1435

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms

has two major categories, in vivo and in vitro. In vivo models include absorption studies in humans and animals; in vitro models include Parallel Artificial Membrane Permeability Assay, tissue model (Ussing Perfusion Chamber model), in situ model (Intestinal Perfusion model) and cell model. Among all the permeability study models, the data from in vivo model can truly reflect the absorption of drugs in the body system but it is difficulty to get. Rat in situ intestinal perfusion have become the most reliable and cost-effective model in all permeability study model. Perfusion studies in rat intestine were highly correlated with human data, even though the type of transporters and their expression levels may vary between species^[4,5]. It has been reported that a high correlation was observed between human and rat small intestine permeability ($R^2=0.80-0.95$) for drugs which permeate through the intestinal by carrier-mediated absorption and passive diffusion mechanisms^[6].

In situ Single-Pass Intestinal Perfusion (SPIP) model provides the advantages of precise experimental control, intact blood supply, and ability to investigate regional factors influencing intestinal absorption of compounds^[7-9]. It has been widely utilized to predict the extent of drug absorption and to clarify absorption mechanisms^[6,10]. A list of 15 drugs, which proposed as model drugs by Food and Drug Administration (FDA) for investigation intestinal permeability, were chosen with different permeability characteristics: low, moderate and high permeability. In this work, we investigated the effective permeability coefficients (Peff) of these model drugs and compared with literature human data to establish the study model for intestinal permeability of drug.

MATERIALS AND METHODS

Materials:

Enalaprilat, lisinopril, amiloride, aciclovir, phenytoin, propranolol, metoprolol, ketoprofen, chlorothiazide, ranitidine, atenolol, hydrochlorothiazide, terbutaline, metformin and antipyrine were purchased from National Institutes for Food and Drug Control (Beijing, China). Acetonitrile, methanol (High-Performance Liquid Chromatography (HPLC) grade) were obtained from TEDIA (Ohio, USA). Deionized water was generated by Milli-Q system from Millipore (Bedford, USA). All other chemicals were analytical reagent grade.

In situ single-pass intestinal perfusion in rat:

All experimental protocols were supervised and managed by Jiangsu Provincial Drug Safety Evaluation Center Agency Committee (protocol code: JA-18-009) and the experiments were conducted in accordance with the guide for the related laws and regulations and Institutional Animal Care and Use Committee (IACUC). Male Sprague Dawley rats weighing 250-350 g (Zhejiang Vital River Laboratory Animal Technology Co., Ltd., Zhejiang, China) were maintained on a 12 h lightdark cycle and had free access to water and food, six replicates for each drug. Rats were fasted for 12 h (water ad libitum) prior to each experiment, and then anaesthetized with pentobarbital solution (30 mg/kg, i.p.). Then placed on a heated slide warmer and under a heating lamp to maintain normal body temperature. The abdomen was opened with a midline incision and jejunal segment of approximately 10 cm was measured, isolated and cannulated with plastic tubing^[11-13]. Cares was taken to avoid disturbance of the circulatory system and the exposed segment was kept moist with 37° saline. Initially, the intestinal segment was rinsed with isotonic saline (37°) until the outlet solution was clear. Krebs-Ringer buffer solution, consisted of 130 mM NaCl, 5mM KCl, 1.27 mM MgSO₄, 0.95 mM CaCl₂, 5 mM glucose and 10 mM NaH₂PO₄, was used as blank perfusion solution. Perfusion solution was prepared by dissolving drugs in Krebs-Ringer buffer solution and then perfused through the intestinal segment at a flow rate of 0.2 ml/min. Blank perfusion solution was perfused for 30 min without sample taken to ensure steady state conditions, followed by additional 80 min of perfusion with samples taken every 20 min. Gravimetric method was used to correct the Net Water Flux (NWF)^[14-17]. All samples were centrifuged at 16 000 rpm for 2 min before analyzed by HPLC. The length of the perfused intestinal segment was measured at the endpoint of the experiment.

The P_{eff} (cm/sec) of model drugs through the rat gut wall was determined according to the following equations:

$$C_{out (corrected)} = C_{out} Q_{out} / Q_{in} (1)$$

$$P_{eff} = -Q ln (C_{out (corrected)} / C_{in}) / 2\pi RL (2)$$

Where C_{out} and C_{in} are the outlet and the inlet concentration of drug respectively, Q_{out} and Q_{in} are the volume that collected at the outlet and the volume that perfusion solution reduced at the inlet respectively, $C_{out(corrected)}$ is the outlet concentration of drug that has been corrected NWF by Gravimetric method, Q is the perfusion buffer flow rate (0.2 ml/min), R is the radius of the intestinal segment, and L is the length of the perfused intestinal segment.

Analytical methods:

All *in situ* SPIP samples were quantified by a Essentia LC-15C HPLC (Shimadzu, Japan). More details about analytical methods of each model drug can be find in supporting information. Processing and analysis of chromatogram were performed on LabSolutions Essentia software.

Statistical analysis:

Values are expressed as mean \pm Standard Deviation (SD). The independent t test and one-way Analysis of Variance (ANOVA) were used to assess differences comparison of permeation parameter. Differences were considered statistically significant when p<0.05. The statistical comparison was made using the statistical package SPSS, V.23.

RESULTS AND DISCUSSION

Permeability values of model drugs were investigated by *in situ* SPIP in rat at steady-state. During the perfusion, the collected sample were centrifuged, and then injected into the HPLC system for analysis. Gravimetric method was used to correct the NWF of rat intestine during the perfusion process. The corrected outlet concentration of each drug was obtained from equation (1), P_{eff} value was calculated from equation (2). The permeability values were presented in fig. 1, and summarized in Table 1. The P_{eff} of different model drugs ranged from 0.0146±0.0026×10⁻⁴ cm/s for enalaprilat and 1.77±0.34×10⁻⁴ cm/s for antipyrine, respectively.

During perfusion, the intestine will absorb and secrete water, which affects the calculation of the P_{eff}. It is necessary to correct the NWF. NWF correction during the SPIP perfusion was usually involves the co-perfusion of a "non-absorbed" marker^[14]. Since phenol red is an ionic compound, it is conventionally considered to be not absorbed in the intestine, and thus it has long been used as a correcting substance for correcting the NWF. However, it has been reported in the literature that phenol red is absorbed in the intestine^[18]. For drugs with high permeability, the influence caused by phenol red has little effect on the test results, which is negligible, but for drugs with low permeability, the influence caused by phenol red cannot be ignored. Therefore, gravimetric method was used to correct the NWF during the perfusion in this study.



Fig. 1: The permeability values of 15 different model drugs obtained by *in situ* SPIP method (mean±SD, n=6)

Drug	Concentration (µg/ml)	P _{eff} (10⁻₄ cm/s)	-
Antipyrine	120	1.77±0.34	_
Ketoprofen	120	1.09±0.56	
Propranolol	120	1.01±0.05	
Metoprolol	120	0.613±0.249	
Phenytoin	30	0.526±0.141	
Metformin	120	0.198±0.0114	
Amiloride	120	0.190±0.094	
Terbutaline	120	0.185±0.014	
Atenolol	120	0.159±0.117	
Hydrochlorothiazide	120	0.140±0.041	
Ranitidine	120	0.0668±0.0147	
Aciclovir	120	0.0282±0.0208	
Chlorothiazide	120	0.0254±0.0212	
Lisinopril	120	0.0161±0.0028	
Enalaprilat	120	0.0146±0.0026	

TABLE 1: THE PERFUSION CONCENTRATIONS AND PERMEABILITY VALUES OF DIFFERENT MODEL DRUGS OBTAINED BY *IN SITU* SPIP METHOD (MEAN±SD, n=6)

In this work, the P_{eff} values investigated by in situ SPIP in rat were ranged from 0.0146±0.0026×10⁻ ⁴ cm/s for enalaprilat to $1.77\pm0.34\times10^{-4}$ cm/s for antipyrine. Drug with $P_{eff} < 0.03 \times 10^{-4}$ cm/s in the rat small intestine is classified as low permeability, whereas drug with $P_{eff} > 0.2 \times 10 - 4$ cm/s is completely absorbed (classified as high permeability)^[4]. Among the selected model drugs, the P_{eff} of high permeability drugs obtained in this experiment were higher than 0.2×10^{-4} cm/s and P_{eff} of low permeability drugs were less than 0.03×10^{-4} cm/s, indicating a high correlation of in situ SPIP model to permeability classification of drug. For industry with desire in generic drug development, the investigation in permeability of drugs by a appropriate method is crucial. The FDA has approved the biowaiver for immediate-release solid oral dosage forms based on a BCS system, which divided drugs in different categories according to solubility and permeability. The in situ SPIP method could be suitable for investigation of permeability.

Table 2 summarizes the permeability values for 8 different model drugs obtained by in-situ SPIP and literature human P_{eff} data^[19-22]. The correlation between the two sets of data, the *in situ* SPIP *vs*. human P_{eff} value, was presented in fig. 2. It can be seen that an excellent correlation was obtained between the experimental rat P_{eff} and literature human P_{eff} , as evident by correlation coefficient

 (R^2) of 0.99, which indicating the validity of this work. These results indicated that *in situ* SPIP method could be used for investigation of the intestinal absorption of new compounds, and for predicting the absorption fraction in human, which is important in early drug development. A strong correlation was observed between experimental rat P_{eff} data and literature human F_{abs}

 $(R^2=0.93)$. Fig. 3 fitting to the follow equation;

$$F_{abs} = 1 - e^{-7.06Peff}$$
 (3)

Table 3 summarizes the literature human F_{abs} data, and the corresponding predicted F_{abs} by the experimental model using above-mentioned equation, and BCS permeability class^[19,23]. According to FDA, when there is no evidence to suggest the instability in gastrointestinal tract, drugs can be considered as high permeability drug when the fraction absorbed in human gastrointestinal tract is investigated to be 85 % or more^[2]. This means that drugs with F_{abs} higher than 85 % can be considered as high permeability drug. As shown in Table 3, F_{abs} of 5 different model drugs obtained by equation (3) were higher than 85 %, indicating a high correlation of absorption fraction between this work and literature. It can be seen in Table 3 that the F_{abs} of metformin was 86 %, which was very nearly to the boundary between high permeability and moderate permeability. It was

www.ijpsonline.com

classed as moderate permeability by FDA. Similar situation have occurred in ranitidine, which literature F_{abs} was 50 %, nearly to the boundary between moderate permeability and low permeability. Similar finding also have been found in other literature^[6]. In this work, the predicted F_{abs} of metformin and ranitidine using above-mentioned equation was 75 % and 38 %, respectively. When drugs with F_{abs} nearly

to the boundary between different permeability classification, the application of the *in situ* SPIP model may not distinguish them well, which may be a limitation of the model. In this study, all the investigated model drugs by *in situ* SPIP have similar permeability classification compare with literature permeability classification. The predicted human F_{abs} by *in situ* SPIP model were linearly correlated with literature human F_{abs} (R²=0.90) (fig. 4).

TABLE 2: PERMEABILITY VALUES FOR 8 OF 15 DIFFERENT MODEL DRUGS OBTAINED BY IN SITU SPIP AND LITERATURE HUMAN P of DATA. DATA PRESENTED AS MEAN±SD

Drug	P _{eff} (10 ⁻⁴ cm/s)					
Diug	Human, literature	Rat, experimental				
Antipyrine	5.6±1.6	1.77±0.34				
Propranolol	2.8±1.3	1.01±0.05				
Metoprolol	1.5±0.9	0.613±0.249				
Terbutaline	0.3±0.3	0.185±0.014				
Atenolol	0.15±0.2	0.159±0.117				
Hydrochlorothiazide	0.04±0.05	0.140±0.041				
Ranitidine	0.273±0.247	0.0668±0.0147				
Enalaprilat	0.1±0.3	0.0146±0.0026				



Fig. 2: The correlation between literature human Per and experimental rat Per for 8 of 15 different model drugs



Fig. 3: The correlation between literature human F_{abs} and experimental rat P_{eff} of 15 different model drugs

Davia	Literatur	re human data	Predicted data			
Drug –	F _{abs} (%)	Permeability class	F _{abs} (%)	Permeability class		
Antipyrine	100	Н	100	Н		
Ketoprofen	100	Н	100	Н		
Propranolol	100	Н	100	Н		
Metoprolol	95	Н	99	Н		
Phenytoin	90	Н	98	Н		
Metformin	86	Μ	75	Μ		
Amiloride	50	Μ	74	Μ		
Terbutaline	73	Μ	73	Μ		
Atenolol	50	Μ	67	Μ		
Hydrochlorothiazide	67	Μ	63	Μ		
Ranitidine	50	Μ	38	Μ		
Aciclovir	16	L	18	L		
Chlorothiazide	13	L	16	L		
Lisinopril	28	L	11	L		
Enalaprilat	10	L	10	L		

TABLE	3:	LITERATURE	HUMAN	F _{abs}	DATA	AND	CORRESPONDING	PREDICTED	F _{abs}	AND	BCS
PERME/	ABI	LITY CLASS	OF 15 DIFF	ERE	NT MO	DEL D	RUGS		455		

Note: H: High; M: Moderate and L: Low



Fig. 4: The correlation between literature human F_{abs} and predicted human F_{abs} of 15 different model drugs

In this work, we investigated the P_{eff} of 15 model drugs with different permeability classification by *in situ* SPIP method. The P_{eff} of high permeability drugs were higher than 0.2×10^{-4} cm/s and P_{eff} of low permeability drugs were less than 0.03×10^{-4} cm/s. The P_{eff} values obtained in this work showed a high correlation with those in human, and a strong correlation was observed between human F_{abs} and experimental rat P_{eff} . Besides, the literature human F_{abs} and predicted human F_{abs} was strongly correlated. The results indicating that *in situ* SPIP method could be used for investigation of the intestinal absorption of new compounds and for prediction the absorption fraction in human, which is crucial in drug development.

Acknowledgements:

This study was financially supported by the Association Fund Project of the Science and Technology Department of Bijie Prefecture (No. [2023] 61).

Conflict of interests:

The authors declare that they have no competing interest.

REFERENCES

- 1. Amidon GL, Lennernäs H, Shah VP, Crison JR. A theoretical basis for a biopharmaceutic drug classification: The correlation of *in vitro* drug product dissolution and *in vivo* bioavailability. Pharm Res 1995;12:413-20.
- 2. Waiver of *in vivo* bioavailability and bioequivalence studies for immediate-release solid oral dosage forms based on a biopharmaceutics classification system guidance for industry. Food and Drug Administration. 2017.
- Morais JA, Lobato MD. The new European Medicines Agency guideline on the investigation of bioequivalence. Basic Clin Pharmacol Toxicol 2010;106(3):221-5.
- Fagerholm U, Johansson M, Lennernäs H. Comparison between permeability coefficients in rat and human jejunum. Pharm Res 1996;13:1336-42.
- Kim JS, Mitchell S, Kijek P, Tsume Y, Hilfinger J, Amidon GL. The suitability of an *in situ* perfusion model for permeability determinations: Utility for BCS class I biowaiver requests. Mol Pharm 2006;3(6):686-94.
- Lozoya-Agullo I, Zur M, Wolk O, Beig A, González-Álvarez I, González-Álvarez M, *et al. In situ* intestinal rat perfusions for human F_{abs} prediction and BCS permeability class determination: Investigation of the single-pass vs. the Doluisio experimental approaches. Int J Pharm 2015;480(1-2):1-7.
- Kuang G, Yi H, Zhu M, Zhou J, Shang X, Zhao Z, et al. Study of absorption characteristics of the total saponins from Radix *Ilicis Pubescentis* in an *in situ* Single-Pass Intestinal Perfusion (SPIP) rat model by using Ultra Performance Liquid

Chromatography (UPLC). Molecules 2017;22(11):1867-84.

- Jagabalan JY, Murugaiyah V, Zainal H, Mansor SM, Ramanathan S. Intestinal permeability of mitragynine in rats using *in situ* absorption model. J Asian Nat Prod Res 2019;21(4):351-63.
- 9. Al Shaker HA, Qinna NA, Badr M, Al Omari MM, Idkaidek N, Matalka KZ, *et al.* Glucosamine modulates propranolol pharmacokinetics *via* intestinal permeability in rats. Eur J Pharm Sci 2017;105:137-43.
- Patel JR, Barve KH. Intestinal permeability of lamivudine using single pass intestinal perfusion. Indian J Pharm Sci 2012;74(5):478-81.
- Athukuri BL, Neerati P. Enhanced oral bioavailability of domperidone with piperine in male wistar rats: Involvement of CYP3A1 and P-gp inhibition. J Pharm Pharm Sci 2017;20:28-37.
- Sirisha K, Achaiah G, Prasad N, Bhasker S, Umachander L, Reddy VM. Multidrug resistance reversal activity of some new dihydropyridines studied by *in situ* Single-Pass Intestinal Perfusion (SPIP) method in rat. Pharm Chem J 2018;52:8-14.
- 13. Wu L, Bi Y, Wu H. Formulation optimization and the absorption mechanisms of nanoemulsion in improving baicalin oral exposure. Drug Dev Ind Pharm 2018;44(2):266-75.
- 14. Sutton SC, Rinaldi MT, Vukovinsky KE. Comparison of the gravimetric, phenol red, and 14C-PEG-3350 methods to determine water absorption in the rat single-pass intestinal perfusion model. Aaps Pharmsci 2001;3:93-7.
- 15. Reis JM, Dezani AB, Pereira TM, Avdeef A, Serra CH. Lamivudine permeability study: A comparison between PAMPA, ex vivo and *in situ* Single-Pass Intestinal Perfusion (SPIP) in rat jejunum. Eur J Pharm Sci 2013;48(4-5):781-9.
- 16. Li Y, Zhang B, Liu M, Zhang X, Shi D, Guo L, *et al.* Further study of influence of *Panax notoginseng* on intestinal absorption characteristics of triptolide and tripterine in rats with *Tripterygium wilfordii*. Pharmacogn Mag 2018;14(53):95-102.
- 17. Ates M, Kaynak MS, Sahin S. Effect of permeability enhancers on paracellular permeability of acyclovir. J Pharm Pharmacol 2016;68(6):781-90.
- Cui SM, Zhao CS, He ZG. Study on absorption mechanism of puerarin using rat everted gut sac. Lishizhen Med Mater Med Res 2008;19:1715-6.
- 19. Salphati L, Childers K, Pan L, Tsutsui K, Takahashi L. Evaluation of a single-pass intestinal-perfusion method in rat for the prediction of absorption in man. J Pharm Pharmacol 2001;53(7):1007-13.
- 20. Lennernas H, Nylander S, Ungell AL. Jejunal permeability: A comparison between the ussing chamber technique and the single-pass perfusion in humans. Pharm Res 1997;14(5):667-71.
- 21. Winiwarter S, Bonham NM, Ax F, Hallberg A, Lennernäs H, Karlén A. Correlation of human jejunal permeability (*in vivo*) of drugs with experimentally and theoretically derived parameters. A multivariate data analysis approach. J Med Chem 1998;41(25):4939-49.
- 22. Takamatsu N, Kim ON, Welage LS, Idkaidek NM, Hayashi Y, Barnett J, *et al.* Human jejunal permeability of two polar drugs: Cimetidine and ranitidine. Pharm Res 2001;18:742-4.
- 23. Skolnik S, Lin X, Wang J, Chen XH, He T, Zhang B. Towards prediction of *in vivo* intestinal absorption using a 96-well Caco-2 assay. J Pharm Sci 2010;99(7):3246-65.