Expression of ID2 and Cyclooxygenase 2 Genes in Endometrial Tissues and Their Clinical Significance in the Pathogenesis of Endometrial Polyps

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Wu et al.: Expression of ID2 and COX-2 Genes in endometrial tissues of endometriosis patients

To investigate the expression of Deoxyribonucleic acid binding inhibitor 2 and Cyclooxygenase 2 genes in endometrial polyps patient’s endometrial tissues and analyze their clinical significance is the main objective. 38 patients with endometrial polyps who were treated at the minimally invasive gynecologic and reproductive surgery department of our hospital from October 2020 to December 2020 were selected as the test group, and their polyps and the endometrial tissue around the polyps were taken. 36 cases of women who had undergone intrauterine device placement surgery at the same time were scraped from the endometrium before intrauterine device placement. The messenger ribonucleic acid expression and protein levels of deoxyribonucleic acid binding inhibitor 2 and cyclooxygenase 2 in endometrial tissues were detected by quantitative reverse transcription polymerase chain reaction and western blot. In addition, enzyme-linked immunosorbent assay was used to detect the levels of vascular endothelial growth factor, tumor necrosis factor-alpha, interleukin 1 beta and cancer antigen 125 and to analyze their relationship with the expression of Deoxyribonucleic acid binding inhibitor 2 and cyclooxygenase 2 in endometrial tissues of endometrial polyp’s patients. The results showed that the expression of Deoxyribonucleic acid binding inhibitor 2 and cyclooxygenase 2 messenger ribonucleic acid and protein levels in endometrial tissues were significantly higher in the experimental group compared with the control group, and the difference was statistically significant. Deoxyribonucleic acid binding inhibitor 2 was positively correlated with vascular endothelial growth factor, tumor necrosis factor alpha, interleukin 1 beta and cancer antigen 125 (p<0.05), while cyclooxygenase 2 was positively correlated with interleukin 1 beta and cancer antigen 125 in the endometrial tissues of patients in the test group. The sensitivity of Deoxyribonucleic acid binding inhibitor 2 was 87.32 %, specificity was 64.88 % and cut-off value was 1.68; the sensitivity of cyclooxygenase 2 was 93.15 %, specificity was 84.33 % and cut-off value was 2.03. Deoxyribonucleic acid binding inhibitor 2 and cyclooxygenase 2 in endometrial tissues has a high predictive value for the clinical diagnosis of endometrial polyps. Expression is closely related to the development of endometrial polyps, and both may be candidate specific target genes for endometrial polyps, and both may be used as markers for the clinical diagnosis of endometrial polyps.

Key words: Endometrial polyps, deoxyribonucleic acid binding inhibitor 2, cyclooxygenase 2, clinical significance

Endometrial polyps are one of the important causes of irregular menstruation or postmenopausal vaginal bleeding, which can occur at any age. At present, the incidence of endometrial polyps has an increasing trend. Endometrial carcinoma is one of the three major malignant tumors of female genital tract. It is more common in elderly women, accounting for 20 % to 50 % of female reproductive malignant tumors. The pathogenesis of endometrial polyps is still unclear. Studies have shown that endometrial polyps proliferate and form implanted lesions, and the pathogenesis involves adhesion, invasion and angiogenesis[1,2]. The
pathological process is similar to the proliferation and migration of tumor tissue. At present, it is believed that the factors related to cell biology are involved in the development of endometrial polyposis. Studies have shown[3,4] that Deoxyribonucleic acid binding inhibitor 2 (ID2) is related to the proliferation and migration of tumor cells. Cytochrome P450 aromatase (P450arom) is a speed limiting enzyme in estrogen synthesis. Many studies have confirmed that in endometrial polyps and endometrial cancer, the expression of P450arom in local lesions is positively correlated with cyclooxygenase 2 (COX-2)[5-7]. At present, there are relatively few studies on the expression of ID2 and COX-2 in endometrial tissues of patients with endometrial polyposis. Therefore, this study is to explore the abnormal expression of ID2 and COX-2 in endometrial tissue of patients and their clinical application value.

MATERIALS AND METHODS

38 patients with endometrial polyps diagnosed and treated in gynecology department of our hospital from October 2020 to December 2020 were selected as the experimental group. All patients were pathologically diagnosed as endometrial polyp, and the corresponding polyp tissues and endometrial tissues around the polyp were taken. In addition, 36 cases of women who underwent insertion of intrauterine contraceptive device in our hospital during the same period were scraped a little endometrium before placing the intrauterine device (IUD). The samples were taken in the first half of the menstrual cycle, and were confirmed as normal endometrial tissue by pathology. All the women did not receive any hormone therapy 3 mo before operation, and there were no estrogen related diseases such as cervical polyps, uterine fibroids, endometrial polyps, etc. there was no significant difference in age between the two groups (p>0.05). The age of patients in the experimental group was 27-45 (33.42±7.14) y old, and that of the control group was 25-47 (34.76±8.30) y old. This study was approved by the ethics committee of our hospital, and the participants were informed and agreed. Inclusion criteria: Patients who met the diagnostic criteria of endometrial polyps; Patients without hormone therapy; Patients without other serious hormone disorders.

Exclusion criteria: Patients with endometrial tumors, uterine fibroids and other diseases; Patients with severe liver and kidney dysfunction; Patients with severe endocrine, respiratory, digestive and other system diseases; Patients with incomplete clinical data. There was no statistically significant difference in age between the two groups of patients (p>0.05), and they were comparable.

The messenger ribonucleic acid (mRNA) expressions of ID2 and COX-2 in the endometrial tissues of the two groups were detected by Quantitative reverse transcription polymerase chain reaction (qRT-PCR):

Endometrial tissues of the two groups of patients were selected and two samples were retained. The first specimen was washed with phosphate-buffered saline (PBS) and stored in liquid nitrogen tank. The relative mRNA expression levels of ID2 and COX-2 were detected by real-time fluorescence quantitative PCR. Total RNA was extracted with Trizol kit. The complementary Deoxyribonucleic acid (cDNA) was synthesized with reference to the reverse transcription kit. The reaction system was 20 µl: 2x SYBR (Synergy Brands, Inc.) mix 10 µl, cDNA 1 µl and double-distilled water (ddH₂O) 8 µl. The upstream and downstream primers were 0.5 µl each, with Beta-Actin (β-actin) as internal reference. The primers were synthesized by Wuhan Biomisp Co., Ltd. And the design of primers is as follows (Table 1). The reaction set-up procedure was as follows: pre-denaturation at 95º for 1 min; 95º for 15 s, 95º for 15 s, 60º for 15 s, 72º for 2 min, with a total of 40 cycles. The expression levels of ID2 and COX-2 mRNA in endometrium were quantitatively analyzed by 2ΔΔCT.

The expressions of ID2 and COX-2 were detected by western blot assay:

The second endometrial tissue samples retained in 1.2.1 were ground and added into protein lysis buffer containing protease inhibitor to extract the total protein of endometrial tissue and determine the total protein. The levels of ID2 and COX-2 protein in endometrium were detected by western blot with β-actin as internal reference. The detection was carried out strictly in accordance with the operation instructions. The

<table>
<thead>
<tr>
<th>Gene</th>
<th>Forward</th>
<th>Reverse</th>
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<tbody>
<tr>
<td>ID2</td>
<td>5’-TCTACGACATGAACGGCTG-3’</td>
<td>5’-GGTCGCCCTGATGTAGTCGAT-3’</td>
</tr>
<tr>
<td>COX-2</td>
<td>5’-TTCAAATGAGATTGTGGAAAAAT-3’</td>
<td>5’-AGATCATCTCCTGCTGAGTATCTT-3’</td>
</tr>
<tr>
<td>β-actin</td>
<td>5’-TGCTGTCCCTGTATGCT-3’</td>
<td>5’-TGATGTCACGCACGATT-3’</td>
</tr>
</tbody>
</table>
images were taken by Tanon software, and the protein expression level was analyzed semi quantitatively

**Detection and observation indicators:**

5 ml of fasting venous blood was taken from two groups and divided into two parts immediately. One sample was centrifuged, and the supernatant was aspirated. The serum levels of vascular endothelial growth factor (VEGF) and tumor necrosis factor-alpha (TNF-α) were determined by enzyme-linked immunosorbent assay (ELISA) according to the instructions of the test kit. The other sample was placed in the anticoagulant tube, and the detection reagents of interleukin 1 beta (IL-1β) and cancer antigen 125 (CA 125) were added. The expression level of IL-1β and CA 125 was detected by colloidal gold method. The levels of VEGF, TNF-α, IL-1β and CA 125 in the two groups were observed, and the correlation between the expressions of ID2 and COX-2 and the levels of VEGF, TNF-α, IL-1β and CA 125 were analyzed. The correlation between ID2 and COX-2 expression and clinical indicators of patients was compared and analyzed.

**Analysis and Statistics:**

Statistical software, Statistical Package for the Social Sciences SPSS 22.0 was used to analyze the data. The quantitative data were represented as mean value±standard deviation, and the comparison between the two groups was performed by t test. In addition, the qualitative data were described by the number of cases (%), while the comparison between groups was conducted by F test (statistical test). Moreover, the correlation analysis between variables was performed by person correlation analysis. p<0.05 was significant difference.

**RESULTS AND DISCUSSION**

The results of PCR showed that compared with the control group, the expressions of ID2 and COX-2 mRNA in the experimental group were significantly increased, and the difference was statistically significant (p<0.05), as shown in fig. 1.

According to the results of western blot, the protein levels of ID2 and COX-2 in endometrial tissue of the experimental group were significantly higher than those of the control group (p<0.05), and the difference was statistically significant (p<0.05), as depicted in fig. 2.

Compared with the control group, the levels of VEGF, TNF-α, IL-1β and CA 125 in the experimental group were significantly higher than those in the control group (p<0.05), as shown in fig. 3.

In the endometrial tissues of the experimental group, ID2 was positively correlated with VEGF, IL-1β and CA 125, while COX-2 was positively correlated with IL-1 and CA 125, with statistically significant differences (p<0.05), as shown in Table 2. The results showed that ID2 and COX-2 in endometrial tissue had high predictive value for clinical diagnosis of endometrial polyps, among which the sensitivity of ID2 was 87.32 %, specificity was 64.88 % and truncation value was 1.68. The sensitivity and specificity of COX-2 were 93.15 %, 84.33 %, and the truncation value was 2.03, as shown in Table 3.

Endometrial polyps can cause chronic inflammatory changes in the internal environment of pelvic inflammatory disease, resulting in irregular menstruation or postmenopausal vaginal bleeding. Its occurrence and development can lead to a variety of adverse clinical outcomes[8].

Noninvasive investigations
such as transvaginal ultrasonography, with the use of 3-dimensional ultrasonography remain the mainstay of first-line investigation. At present, hysteroscopic surgery is used to remove endometrial polyps, but there is a high recurrence rate after Hysteroscopic resection is the most effective management for endometrial polyps and allows histologic assessment. Preutthipan et al. reported that the recurrence rate can reach 15%[9]. Studies have shown that the changes of cell biology related factors are related to the pathogenesis of endometrial polyps, which can accelerate the infiltration process of glandular cells by affecting the proliferation and invasion of glandular cells in endometriosis tissue, and eventually lead to the continuous deterioration of the disease[10]. Therefore, it is of great significance to seek specific molecular markers for the diagnosis and treatment of endometrial polyps. Studies have shown that ID2 and COX-2 are related to the pathogenesis of ovarian endometrial tumor, and both of them are directly or indirectly involved in the process of cell migration, invasion and angiogenesis[11]. There are few studies on the expression of ID2 and COX-2 in patients with endometrial polyps. Therefore, this study mainly discusses the expression levels of ID2 and COX-2 in endometrial tissues of patients with endometrial polyps, which can provide theoretical basis for the pathogenesis of endometrial polyps.

ID2 is a member of the differentiation /DNA binding inhibitor family. It forms heterodimer by binding with basic helix loop helix transcription factors, resulting in ID2 deletion, affecting its binding with target genes, and then inhibiting the expression of related genes[10]. Studies have shown that abnormal expression of ID2 can promote the proliferation and migration of cancer cells, which is closely related to cell adhesion, protein glycosylation, cell invasion and angiogenesis[12]. Due to the similarity between endometrial polyps and cancer, the expression of ID2 mRNA was detected by qRT-PCR in the experimental group and the control group, and the protein expression level was detected by western blot assay. The results showed that the expression of ID2 mRNA and protein in the experimental group were higher than those in the control group, suggesting that ID2 was up-regulated in endometrial tissues of endometrial polyps. The relationship between ID2 in the endometrial tissue of the experimental group and the clinical indicators of the patients was observed, and the results showed that ID2 in the endometrial tissue of the experimental group was positively correlated with VEGF, IL-1β and CA 125, among which the high expression of VEGF can increase the infiltration capacity of heterotopic gland cells[13]. IL-1β and CA 125 are highly expressed in patients with endometrial polyps, which can improve the adhesion ability of glandular cells by damaging mitochondria in endometrial polyps, and eventually damage the epithelial tissue of basement membrane seriously[14-16]. It is suggested that the expression of ID2 in endometrial tissue can be used as an index to predict the severity of endometrial polyps.

Cyclooxygenase includes COX-1 and COX-2 isozymes. COX-1 catalyses the release of prostaglandin E2 (dinoprostone) by cells at rest, which mainly maintains cell self-stability. COX-2 was almost not expressed under normal conditions. When inflammation and cell proliferation and differentiation appeared, it was induced by cytokines, growth factors and tumor promoters, and its expression could be rapidly increased by 8-10 times. COX-2 is a rate-limiting enzyme of prostaglandin production, which is widely considered as one of the important inducible enzymes in inflammatory process[17]. Recent studies have found that the biological function of COX-2 can enhance the angiogenesis of subunit fibroblast growth factor and VEGF by stimulating the mitosis of human endothelial cells[18]. Based on this, the expression of COX-2 in endometrial tissues of patients with endometrial polyps and normal endometrial tissues was detected by qRT-PCR.

![Fig. 3: Comparison of clinical indicators between the two groups](image)

TABLE 2: RELATIONSHIP BETWEEN ID2, COX-2 AND CLINICAL INDEXES

<table>
<thead>
<tr>
<th>Indexes</th>
<th>ID2</th>
<th>COX-2</th>
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</thead>
<tbody>
<tr>
<td>VEGF</td>
<td>0.593</td>
<td>0.017</td>
</tr>
<tr>
<td>TNF-α</td>
<td>0.081</td>
<td>0.503</td>
</tr>
<tr>
<td>IL-1β</td>
<td>0.575</td>
<td>0.021</td>
</tr>
<tr>
<td>CA 125</td>
<td>0.331</td>
<td>0.032</td>
</tr>
</tbody>
</table>

TABLE 3: THE PREDICTIVE VALUE OF ID2 AND COX-2 FOR CLINICAL DIAGNOSIS

<table>
<thead>
<tr>
<th>Gene</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>Truncation value</th>
</tr>
</thead>
<tbody>
<tr>
<td>ID2</td>
<td>87.32 %</td>
<td>64.88 %</td>
<td>1.68</td>
</tr>
<tr>
<td>COX-2</td>
<td>93.15 %</td>
<td>84.33 %</td>
<td>2.03</td>
</tr>
</tbody>
</table>
PCR and western blot assay in this study. The results illustrated that the expression of COX-2 in endometrial tissues of patients with endometrial polyps in the experimental group was higher than that in the control group, indicating that COX-2 was highly expressed in endometrial tissues of patients with endometrial polyps. Further studies showed that COX-2 was positively correlated with IL-1β and CA 125, which suggested that COX-2 might be involved in the occurrence and development of endometrial polyps and accelerate the deterioration of the disease through the interaction with serum related factors. The results of this study indicated that ID2 and COX-2 in endometrial tissue have high predictive value for clinical diagnosis of endometrial polyps, indicating that ID2 and COX-2 in endometrial tissue can be used as predictive factors for clinical diagnosis of endometrial polyps. At the same time, this study analyzed the related factors of endometrial polyps by logistic regression analysis. The results showed that ID2 and COX-2 in endometrial tissue were risk factors for patients with endometrial polyps, indicating that both of them can interact and promote the development of the disease.

In summary, the high expression of ID2 and COX-2 in endometrial tissue is closely related to the occurrence and development of endometrial polyps. Both of them may be candidate specific target genes for endometrial polyps, and both of them can be used as markers for clinical diagnosis of endometrial polyps.

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Conflict of interests:

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REFERENCES


