# HPTLC Method for the Determination of Gatifloxacin in Human Plasma

#### G. SOWMIYA\*, M. GANDHIMATHI, T. K. RAVI AND K. R. SIREESAA

Department of Pharmaceutical Analysis, Sri Ramakrishna Institute of Paramedical Sciences, College of Pharmacy, 395, Sarojini Naidu Road, Coimbatore - 641 044, India.

A high performance thin layer chromatographic method for the determination of gatifloxacin in human plasma using paracetamol as internal standard is proposed. Gatifloxacin in plasma was extracted with dichloromethane at pH 4.5 (phosphate buffer). The dichloromethane layer was evaporated and residue was reconstituted with ethanol and quantified using the mobile phase consisting of chloroform:methanol:ethanol:ammonia (4:2:2:2% v/v/v/v). The limit of quantitation was 50 ng/spot. The % RSD of the intra-day and inter-day was less than 15. The method was found to be simple, sensitive, rapid and could be used in the pharmacokinetic study involving human volunteers.

Gatifloxacin (±1-cyclopropyl-6-fluoro-1,4-dihydro-8methoxy-7-{3-methyl-1-piperazinyl} 4-oxo-3-quinoline carboxylic acid) is an advanced general antibacterial agent<sup>1,2</sup>, has broader spectrum of antibacterial activity than the other fluoroquinolones<sup>3</sup>. The different analytical methods reported for the estimation of gatifloxacin includes HPLC<sup>4-7</sup>, LC/SI-MS/MS<sup>8</sup>, spectrophotometry<sup>9</sup>, and HPTLC<sup>10</sup>. Estimation from human plasma<sup>11</sup>, serum and urine<sup>12</sup> were also reported. These methods are lacking from higher detection limit and time consuming. The present HPTLC method using paracetamol as internal standard is simple and cost effective for the determination of gatifloxacin in human plasma with ease and higher sensitivity.

Gatifloxacin was supplied by Cipla Pharmaceuticals Ltd, Mumbai. Paracetamol pure sample was a gift from Medoform Pharmaceuticals, Chennai. Dichloromethane and ethanol (S. D. Fine Chemical Ltd, India) were of AR grade. Sodium dihydrogen phosphate was purchased from E. Merck (India) Ltd, Mumbai. Plasma was collected from healthy human volunteers.

An HPTLC system (Camag, Muttez, Switzerland) equipped with Linomat V semi automatic sample applicator, twin trough developing chamber and TLC scanner3 with WINCATS evaluation, ATS software (version 1.2.6) were used for the analysis, silica gel  $60GF_{254}$  plates (10 × 10), layer thickness 0.2 mm, (E-Merck, Darmstadt, Germany) were used as stationary phase. A mobile phase consisting of chloroform:methanol:ethanol:ammonia (4:2:2:2% v/v/v/v) was used for development of chromatogram. The spot was scanned using 291 nm.

A stock solution of 1 mg/ml of gatifloxacin and paracetamol were prepared separately in ethanol. Appropriate dilutions of gatifloxacin were made to produce working standard solutions in the range of 50 to 400  $\mu$ g/ml. Paracetamol solution was diluted to get 250  $\mu$ g/ml.

To 500  $\mu$ l of plasma sample, 500  $\mu$ l of working standards of gatifloxacin and paracetamol solution (250  $\mu$ g/ml) were added and mixed. Phosphate buffer pH 4.5 (1000  $\mu$ l) was added along with 5 ml of dichloromethane and the contents were vortexed for 1 min and centrifuged at 2000

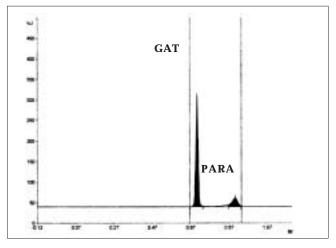


Fig. 1: Typical HPTLC chromatogram for analysis of gatifloxacin Plasma sample spiked with 400 ng/spot. Rf value of gatifloxacin is 0.70 and paracetamol is 0.90. GAT stands for gatifloxacin and PARA stands for paracetamol.

<sup>\*</sup>For correspondence E-mail: sowmi53@yahoo.co.in

rpm for 5 min at 35°. An aliquot of (1000  $\mu$ l) of the supernatant was taken and evaporated to dryness with nitrogen gas. The residue was reconstituted with 1000  $\mu$ l of ethanol. Aliquots of 10  $\mu$ l each of standard extract was spotted on TLC plates.

The chromatogram of plasma spiked with gatifloxacin and paracetamol is shown in fig. 1. No endogenous interfering peaks were visible in blank plasma at the spot of gatifloxacin and the internal standard there by confirming the specificity of the analytical method. Both the analyte and the internal standard were well separated with Rf values of  $0.70\pm0.03$  and  $0.90\pm0.02$ , respectively.

The resolution (Rs) between gatifloxacin and paracetamol was 3.3. The efficiency of the method is studied by calculating number of theoretical plates and was found as 14,400. Peak area ratios of standard gatifloxacin to that of internal standard were measured. A representative calibration graph of peak area ratio versus gatifloxacin concentration (50 to 400 ng/spot) resulted in regression equation y=0.0133 x+0.9082 (r = 0.9900).

The intra-day precision was determined by analyzing 4 spiked plasma samples at each concentration is different time on the same day. For the determination of inter-day precision, spiked samples were analyzed on three different days (Table 1). The precision at each concentration was calculated as the % RSD. The vales were found to be with in the specific limit.

The recovery from plasma samples was detected by

## TABLE 1: INTRA AND INTER DAY PRECISION OF DETERMINATION OF GATIFLOXCIN IN PLASMA

Amount spiked ng/spot	Measured		% average accuracy
	Mean (ng/spot)	RSD	
Inter day variation			
50	43.34	2.5	
100	90.66	3.7	
300	267.02	0.61	87.72
400	347.24	1.11	
Intra day variation			
50	44.02	1.76	
100	97.27	7.14	
300	265.51	1.71	86.92
400	346.53	0.29	

RSD: Related standard deviation and mean value of 3 determinations

comparing the recovery of standards that are obtained without plasma matrix but processed similarly. The accuracy was calculated as a percentage of spiked/ nominal concentration

These experiments confirm that the HPTLC method for the determination of gatifloxacin in plasma samples is specific, accurate, and precise and has high sensitivity. The method can be used for analysis of gatifloxacin plasma samples during pharmacokinetic studies in human volunteers.

### ACKNOWLEDGEMENTS

The authors thank Cipla Pharmaceuticals Ltd, Mumbai for the gift sample of gatifloxacin and Medoform Pharmaceuticals Ltd., Chennai for paracetamol and M/S SNR and Sons Charitable trust, Coimbatore for providing facilities to carry out the work.

## REFERENCES

- 1. Budavari, S., Eds., In; The Merck Index, 13th Edn., Merck & Co., Inc., Whitehouse Station, NJ, 2001, 4388.
- 2. Parfitt, K., Eds., In; Martindale, The complete drug reference, 33rd Edn., The Pharmaceutical Press, London, 2002, 3078J.
- 3. Percy, C.M., Ormrod, D., Hurst, M. and Onrust, S.V., Drugs, 2002, 62, 169.
- Lober, S., Zieje, S. and Lode, H., Antimicrob. Agents Chemother., 1999, 43, 1067.
- Liang, H., Kays, M.B and Sowinski, K.M., J. Chromatogr. B. Anal. Tech. Biomed. Life Sci., 2002, 772, 53.
- Holser, B.R. Kays, M.B and Sowinski, K.M., J. Chromatogr. B. Anal. Tech. Biomed. Life Sci., 2003, 798, 167.
- 7. Lakshmi, S and Muthukumaran. A., Indian J. Pharm. Sci., 2005, 67, 367.
- Vishwanathan, K., Bartlett, N.G and Stewart, J.T., Rapid Commun. Mass Spectrom., 2001, 15, 195.
- Patel, P.U., Suhagia, B.N., Patel, C.N., Patel, M.M., Patel, C.G. and Patel, M.G, Indian J. Pharm. Sci., 2005, 67, 356.
- Shah, S.A., Rathod, I.S., Suhagia, B.N. and Bardaniya, M., Indian J. Pharm. Sci., 2004, 66, 306.
- 11. Nabel, K., Sorgel, K. and Nable, G.K., Antimicrob. Agents Chemother., 2001, 45, 293.
- 12. Bornel, K., Hartweg, H. and Lode., J. Chromatogr., 2000, 52, 105.

Accepted 12 April 2007	
Revised 11 August 2006	
Received 23 November 2005	
Indian J. Pharm. Sci., 2007, 69 (2): 301-302	