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## Influence of Phenological Stages on Yield and Quality of Oregano (*Origanum vulgare* L.) Under the Agroclimatic Condition of Doon Valley (Uttarakhand)

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A field experiment was conducted under the agroclimatic conditions of Doon valley, in order to determine the effects of phenological stages on herbage yield and quality of oil in oregano (*Origanum vulgare* L.). Plants were harvested in five phenological stages, *i.e.* early vegetative, late vegetative, flower initiation, full bloom, and fruit set stages. Results showed the significant effects of phenological stages on herbage, yield, and quality of oregano. Harvesting at full bloom stage showed better results in terms of herbage and oil yield. The quality of essential oil was evaluated using GC and GC/MS. Thymol content was rich in all the stages (46.90-62.26%) followed by  $\gamma$ -terpinene (1.11-11.75%) and p-cymene (3.11-5.32%).

Key words: Origanum vulgare L., phenological stages, GC/MS, essential oil, thymol

*Origanum* L. (Lamiaceae), a small genus of perennial and shrubby herb which is distributed in the Mediterranean region and tropical Asia<sup>[1]</sup>. The genus includes some commercially important culinary herbs, including oregano (*Origanum vulgare* L.) and Marjoram (*Origanum majorana* Moench; syn. *Majorana hortensis* L.), which are extensively used for flavoring food products and alcoholic beverages<sup>[2,3]</sup>. In India, the genus is represented by a single species, *O. vulgare*, generally found in subtemperate and temperate Himalaya<sup>[4]</sup>, whereas, the other important species, *O. majorana* is exotic and cultivated in Indian gardens<sup>[5]</sup>.

The chemical composition of oregano oil is very variable in India<sup>[6-10]</sup>. The chemo varieties and the environmental conditions caused to this fact. The major chemo types of oregano oil from Indian origins were found to be thymol<sup>[6,7,10]</sup> and carvacrol<sup>[6-9]</sup>.

In Uttarakhand Himalaya, oregano is wildly grown in forest lands, but due to deforestation, overexploitation, overgrazing the availability of this important traditional medicinal and aromatic herb is limited. The commercial cultivation of this herb is also not in trends, but it is cultivated in some parts of India. The information about harvesting stages, herb, and oil yield of oregano is limited.

Therefore, the purpose of this study was to determine the effects of phenological stages on herbage yield and quality of oil as well as to determine the best harvesting time of wild oregano under the agroclimatic condition of Doon valley of Uttarakhand, India.

The experiment was conducted at the research field of Centre for Aromatic Plants (CAP), Selagui, located at Dehradun, India at an altitude of 680 m above sea level during year 2009-10. Live germplasm of oregano was collected from Bhowali, Nainital (altitude 1700 m) and cultivated for experiment under the climatic conditions of Doon valley. The specimen of oregano was duly identified in Centre for Aromatic Plants (CAP) and deposited with voucher specimens no. (CAP-192). The soil was sandy loam (68.8% sand, 25.6% silt, 5.6% clay) with pH 7.4, EC 0.159 dSm<sup>-1</sup>, 0.03% total nitrogen, 103 ppm available phosphorus, 26.3 ppm available potassium, and 0.47% organic carbon. Five phenological stages, *i.e.* early vegetative, late vegetative, flower initiation, full bloom and fruit

set stages were considered as treatments. Before planting, farmvard manure (200 g/ha) was thoroughly mixed and a basal dose of nitrogen (80 kg/ha), phosphorus (60 kg/ha), and potassium (40 kg/ha) was applied. The rooted plants were transplanted in November, 2009 at 45×45 cm population density. The design of experiment was in randomized block design (RBD) with three replications. In each replication, five plants were randomly selected for the recording of different parameters. Harvesting was done at each phenological stage between the months of February to May, 2010. Growth and yield parameters namely, plant height (cm), number of branch/plant, fresh and dry herb yield (q/ha), fresh and dry oil yield (kg/ha), and oil content (%) were recorded in all the treatments.

Fresh and shade dried leaves with small twigs (aerial parts) of each sample (300 g) of different harvesting stage (phenological stage) were separately hydro distilled for 3 h using a Clevenger-type apparatus. The collected oil samples were dried over anhydrous sodium sulfate and stored in sealed vials at 4° until analyzed.

GC analyses were carried out by an Agilent Technology 6890 N gas chromatograph data handling system equipped with a split/splitless injector and fitted with FID using N<sub>2</sub> as the carrier gas. The column was HP-5 capillary column (30 m×0.32 mm, 0.25  $\mu$ m film thickness) and temperature program was used as follows: initial temperature of 60° (hold: 2 min) programmed at a rate of 3°/min to a final temperature of 220° (hold: 5 min). Temperatures of the injector and FID were maintained at 210 and 250°, respectively.

The GC-MS analyses were carried out on a Perkin Elmer *Clarus* 500 gas chromatograph equipped with a split-splitless injector (split ratio 50:1) data handling system. The column was an Rtx<sup>®</sup>-5 capillary columns (60 m×0.32 mm, 0.25  $\mu$ m film thickness). Helium (He) was the carrier gas at a flow rate 1.0 ml/min. The GC was interfaced with (Perkin Elmer *Clarus* 500) mass detector operating in the EI + mode. The mass spectra were generally recorded over 40-500 amu that revealed the total ion current (TIC) chromatograms. The temperature program was used as described above. The temperatures of the injector, transfer line, and ion source were maintained at 210°, 210°, and 200°, respectively.

Identification of the individual components was made by matching their recorded mass spectra with the library (NIST/Pfleger/Wiley) provided by the instrument software, and by comparing their calculated retention indices with the literature value<sup>[11]</sup>.

Origanum vulgare crop was assessed for growth, yield, and quality at five stages of development (phenological stages), *i.e.* early vegetative, late vegetative, flower initiation, full bloom, and fruit set stages (Table 1). The plant height significantly increased at all the stages with the advancement of crop age. Number of branches increased up to flowering initiation stage and reduced at the time of maturity, i.e. full bloom and fruit set stages. The fresh and dry herbage yield was significantly increased from early vegetative to full bloom stage of the crop, ranged from 21.74-70.78 q/ha and 4.41-16.93 q/ha in fresh and dry herbage, respectively, whereas in fruit set stage, the herbage yields reduced. Similarly, the fresh and dry oil yield also increased from early vegetative to full bloom stage, ranged from 5.61-35.07 and 8.57-40.33 kg/h in fresh and dry oil, respectively, whereas in fruit set stage, the oil yield reduced. In different developmental stages, the oil content (%) showed some differences. Oil content, obtained by fresh yield, ranged from 0.28-0.55% and recorded maximum at full bloom stage (0.55%), whereas content in dry yields ranged from 2.19 to 2.67%, which was also highest at full bloom stage (2.67%). Verma et al. also reported that the oil content in O. vulgare was increased with the advancement of crop age and became highest at full bloom stage, and afterwards it started to decrease. The oil content also found highest at the time of full flowering in *O. majorana*<sup>[12,13]</sup>.

A total number of 26 components accounting for 86.71-89.27% of the total composition were

identified by GC and GC-MS (Table 2). Among the constituents of oil, thymol (46.90-62.26%) was found as the major component followed by y-terpinene (1.11-11.75%) and p-cymene (3.11-5.32%). Thymol which was found in abundance in the essential oils of all the five growth stages, recorded maximum amount at late vegetative (62.26%) which was slightly but steadily reduced with growth of plant and the lowest percentage in early vegetative stage (46.90%). The maximum (62.26%) and minimum (46.90%) thymol content has moderate difference among the growth stages. The composition of y-terpinene increased with the development of the crop. It was recorded in lesser amount in early and late vegetative stages (1.11 and 4.39%), respectively, whereas at flower initiation, dull bloom and fruit set stage it ranged from 10.43-11.75%. The percentage of p-cymene was slightly higher in full bloom and fruit set stage of the crop as compared with vegetative and flowering initiation stage (fig. 1).



Fig. 1: Comparison of three major components at various phenological stages.

The percentage occurrence of three major components (- $\blacksquare$ - thymol; - $\bullet$ -  $\gamma$ -terpinene; and- $\blacktriangle$ - p-cymene) present in the essential oils of *O. vulgare* analyzed at 5 different phenological stages of plant cycle.

TABLE 1: INFLUENCE OF DIFFERENT PHENOLOGICAL STAGES ON GROWTH, YIELD AND OIL CONTENT OF O. VULGARE

Treatments (Phenological stages)	Plant height (cm)	No. of branch/plant	Herb yield (q/ha)		Oil yield (kg/ha)		Oil content (%)	
			Fresh	Dry	Fresh	Dry	Fresh	Dry
Early vegetative	12.44	37.22	21.74	4.41	5.61	8.57	0.28	2.19
Late vegetative	21.78	75.16	32.01	7.30	9.29	16.07	0.32	2.48
Flower initiation	41.58	76.61	59.97	13.89	24.87	30.88	0.47	2.52
Full bloom	49.61	55.19	70.78	16.93	35.07	40.33	0.55	2.67
Fruit set	56.61	46.33	59.27	14.93	23.49	33.15	0.45	2.50
CD at 5% ( <i>P</i> =0.05)	12.03	16.76	20.41	5.71	12.87	13.09	0.19	0.17

Values are given in average

Compounds	RI	Percentage in oil						
		Early vegetative	Late vegetative	Flower initiation	Full bloom	Fruit set		
α-thujene	931	0.89	0.87	1.73	2.02	1.23		
α-pinene	939	0.33	0.45	0.76	0.89	0.67		
Camphene	953	-	0.12	0.13	-	-		
Sabinene	969	1.19	0.21	0.17	0.48	0.62		
1-octan-3-ol	974	0.91	2.14	-	-	-		
B-myrcene	991	2.92	2.73	3.59	3.09	2.82		
α-terpinene	1018	0.13	0.80	2.05	2.17	1.89		
B-phellandrene	1025	0.83	-	0.21	-	0.91		
p-cymene	1026	3.51	4.60	3.11	4.69	5.32		
Limonene	1030	0.08	0.14	0.26	0.44	0.82		
γ-terpinene	1062	1.11	4.39	11.75	10.83	10.43		
Linalool	1095	0.59	0.57	0.33	0.32	0.41		
Trans-sabinene hydrate	1097	0.25	0.32	0.28	-	-		
Terpinen-4-ol	1177	0.20	-	0.42	0.80	0.91		
Thymol methyl ether	1235	-	-	0.34	0.17	0.53		
z-citral	1240	8.71	0.99	-	-	-		
Trans-geraniol	1249	0.90	0.17	-	-	-		
Thymol	1290	46.90	62.26	59.31	57.17	57.11		
e-citral	1338	10.73	1.47	-	-	-		
Neryl acetate	1359	0.65	-	-	-	-		
Trans-caryophyllene	1422	2.13	2.65	3.18	3.24	3.07		
B-farnesene	1440	0.02	0.05	-	-	-		
Germacrene-D	1484	0.21	0.12	0.53	0.76	0.48		
B-bisabolene	1509	1.58	-	0.79	0.66	0.39		
Caryophyllene oxide	1582	1.03	0.94	-	0.22	0.71		
Globulol	1590	3.47	0.72	-	-	-		
Monoterpene hydrocarbons		10.99	14.31	23.76	24.61	24.71		
Oxygenated monoterpenoids		69.84	67.92	60.68	58.46	58.96		
Sesquiterpene hydrocarbons		3.94	2.82	4.5	4.66	3.94		
Oxygenated sesquiterpenoids		4.5	1.66	-	0.22	0.71		
Identified compounds (%)		89.27	86.71	88.94	87.95	88.32		

TABLE 2: COMPARATIVE ESSENTIAL OIL COMPOSITION OF O. VULGARE AT DIFFERENT PHENOLOGICAL STAGES

RI=Retention indices relative to homologous series of n-alkanes (C8-C24) hydrocarbons on non polar Rtx5 capillary column

On the basis of the above findings, the harvesting of herbage at full bloom stage had almost all the parameters slightly greater than those harvested at early and late vegetative, flower initiation, and fruit set stages. Therefore, the study revealed that harvesting of the crop at full bloom is optimum time to get better herbage and oil yield. The oil vield obtained through dry herbage also recorded better results as compared with fresh herbage. In all the developmental stages the quality of the oil found similar having some quantitative variations, as the occurrence of  $\gamma$ -terpinene, which constituted 1.11-4.39% in vegetative stages and increased up to 10.43-11.75% with the advancement of the crop age. The harvesting of the crop in three phenological stages, i.e. flower initiation, full bloom stage, and fruit set stage showed similarity in quality and

quantity of the oil composition as compared with vegetative stages.

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## REFERENCES

- Anonymous. The Wealth of India-Raw Materials. Vol. 7. Council of Scientific and Industrial Research, New Delhi, India; 1966;105-6.
- Baydar H, Sagdic O, Ozkan G, Karadogan T. Antibacterial activity and composition of essential oils from *Origanum*, *Thymbra* and *Satureja* species with commercial importance in Turkey. Food Control 2004;15:169-72.
- Filippo-Dantuono L, Galletti GC, Bocchini P. Variability of essential oil content and composition of *Origanum vulgare* L. populations from a north Mediterranean Area (Liguria Region, Northern Italy). Ann Bot

2000;86:471-8.

- Mukerjee SK. A revision of Labiatae of the Indian Empire. Rec Bot Surv India 1940;14:94-5.
- Anonymous. The Wealth of India-Raw Materials. Vol. 6. Council of Scientific and Industrial Research, New Delhi, India; 1962;226-7.
- Bisht D, Chanotiya CS, Rana M, Semwal M. Variability in essential oil and bioactive chiral monoterpenoid compositions of Indian Oregano (*Origanum vulgare* L.) populations from northwestern Himalaya and their chemotaxonomy. Ind Crop Prod 2009;30:422-6.
- Pande C, Tewari G, Singh S, Singh C. Chemical markers in *Origanum vulgare* L. from Kumaon Himalayas: A chemosystematic study. Nat Prod Res 2012;26:140-5.
- Verma RS, Padalia RC, Chauhan A, Verma RK, Yadav AK, Singh HP. Chemical diversity in Indian oregano (*Origanum vulgare* L.). Chem Biodivers 2010;7:2054-64.
- Verma RS, Rahman L, Verma RK, Chanotiya CS, Chauhan A, Yadav A, et al. Changes in the essential oil content and composition of *Origanum* vulgare L. during annual growth from Kumaon Himalaya. Curr Sci 2010;98:1010-2.

- Verma RS, Padalia RC, Chauhan A. Volatile constituents of *Origanum vulgare* L., 'thymol' chemotype: Variability in North India during plant ontogeny. Nat Prod Res 2012;26:1358-62.
- 11. Adams RP. Identification of essential oil components by Gas chromatography/Mass Spectrometry. 4th ed. Carol Stream, IL, USA: Allured Business Media; 2009.
- Nurzynska-Wierdak R, Dzid AK. Influence of plant density and term of harvest on yield and chemical composition of sweet marjoram (*Origanum* majorana L.). Acta Sci Pol Technol Aliment 2009;8:51-61.
- Sellami IH, Maamouri E, Chahed T, Wannes WA, Kchouk ME, Marzouk B. Effect of growth stage on the content and composition of the essential oil and phenolic fraction of sweet marjoram (*Origanum majorana* L.). Ind Crop Prod 2009;30:395-402.

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