# Isolation and Identification of Endophytic Fungi from *Cassia senna* L.: A Medicinal plant

V. N. UPASANI\*, N. V. DHARMIL, NEETA SHRIVASTAVA1, ANSHU SRIVASTAVA1 AND SONAL SHARMA1

Department of Microbiology, M. G. Science Institute, Ahmedabad, Gujarat 380009, <sup>1</sup>B. V. Patel Pharmaceutical Education and Research Development (PERD) Centre, Thaltej, Ahmedabad 380054, Gujarat, India

### Upasani et al.: Endophytic Fungi from Cassia senna L.

Endophytic fungi are present within all plants and more than one million species are predicted to be present. They are widely studied as they promote plant growth as well as resistance to environmental stress, produce bioactive compounds that have industrial applications and play a vital role in biogeochemical cycles. The medicinal plant Cassia (commonly known as seenas) belongs to the Order Fabales Family Caesalpiniaceae. This plant is a native of tropics which is useful as an ornamental plant, cassia gum, cuisines, etc. It is also a source for the laxative drug known as Sennosides complex, which has been used by Egyptians since ancient times. This drug is mainly obtained from Cassia augustifolia and Cassia fistula. The active constituents of this drug are dianthrone glycosides (Sennosides A, B, C, D, etc.), free anthraquinones (aloe-emodin, chrysophanol, rhein) and anthraquinones glycosides. However, due to urbanization the plant source has become scarce, so an attempt has been made to isolate endophytic fungi from this medicinal plant and screen for biosynthesis of Sennosides complex. Plants were collected from six (6) locations/sites at Ahmedabad, Anand (Boriavi), Bavla, Bhuj (Madhapur and Saiyadpur), Dantiwada in Gujarat; and Jodhpur in Rajasthan. 22 isolates were identified based on colony, microscopic/morphological characteristics; and identity of thirteen isolates was also done by phylogenetic analysis. The endophytes belonged to the genera Alterneria, Aspergillus, Cladosporium, Curvularia, Fusarium, Penicillium, Phellinus, etc. The isolates were obtained to screen for production of Sennosides (extracellular). A further study on the potential for their application in sustainable development, and alternative for bioprospecting of bioactive compounds is envisaged.

#### Key words: Alterneria, Cassia senna, Cladosporium, endophytic fungi, Fusarium, Phellinus

Medicinal plants serve as a natural source of drugs with less side-effects and have been exploited for this purpose for thousands of years in traditional medicine. However, the natural plant resources are diminishing (due to endangered species or difficulty in cultivation), and also the yield of drug is gradually decreasing. Therefore, it has become a necessity to find alternative sources for these drugs by plant cell or tissue culture, chemical synthesis, semi-synthetic route, or from endophytes<sup>[1]</sup>. Endophytic microorganisms (namely bacteria and fungi) residing within most vascular plants have sought the attention of researchers since it became known that an endophyte Taxomyces andreanae produced Taxol (an anticancer drug) similar to its host Taxus brevifolia<sup>[2,3]</sup>. The medicinal plant Cassia senna L. is a well-known tropical medicinal plant that produces Sennosides complex, derived from the leaves. It is a major pharmaceutical laxative prescribed worldwide obtained chiefly from Cassia augustifolia and Cassia fistula.

The active constituents of Senna are dianthrone glycosides (Sennosides A, B, C, D, etc.), free anthraquinones (aloe-emodin, chrysophanol, rhein) and anthraquinones glycosides. Other chemical constituents are rhein, emodin, aloeemodin. naphthalene glycosides, flavonoids, beta-sitosterol, saponins, etc.<sup>[4]</sup>. The plant mainly flourishes in the arid or semi-arid region<sup>[5]</sup>. The Cassia plant has become scarce due to urbanization, so an alternative source for sennosides has to be explored. Therefore, an attempt has been made to isolate and identify endophytic fungi from Cassia senna L. plant samples (cultivated as well as wild) collected from Gujarat and Rajasthan; and screen

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for sennosides complex. The isolated strains could be potential candidates for developing sustainable, cost-effective eco-friendly and alternative for bioprospecting of bioactive compounds (Sennosides complex, antimicrobials, anticancer drugs, etc.). Plants (Cassia senna L) were collected from 6 locations/sites at Ahmedabad, Anand (Boriavi), Bavla, Bhuj (Madhapur and Saiyadpur), Dantiwada in Gujarat and Jodhpur in Rajasthan. The various parts of the plants (flowers, leaves, stem and seeds) were collected in plastic bags and stored in the refrigerator. They were processed for isolation of endophytic fungi within 24-48 h after collection. The plant material was surface sterilized by the modified method of Schulz et al.<sup>[6]</sup>. The plant material was cut into an appropriate size, rinsed under running tap water for 10 min, shaken for a few seconds in 70 % alcohol and rewashed with sterile distilled water. Next step involved the treatment with HgCl<sub>2</sub> (0.1-1 % w/v)+Teepol 2 drops/100 ml (sterile distilled water) for 3-5 min with shaking and then rinsed in sterile distilled water. Further treatment was done with Commercial bleach (sodium hypochlorite) 7-15 %+Teepol 2 drops/100 ml (sterile distilled water) for 10-30 min with shaking and the material was rinsed several times with sterile distilled water. The treated plant material was inoculated onto Potato Dextrose Agar ((PDA), Hi-media) plates with streptomycin, and incubated in plant growth chamber at  $25\pm3^{\circ}$  for isolation of endophytic fungi. Pure fungal cultures were obtained by repeated transfers on PDA plates and preserved on PDA slants at 2° to 8° at refrigeration temperature. The controls included: a) 2 PDA plates exposed at the time of inoculation in the laminar air flow and b) inoculating the sterilized water used in the last rinse (one drop) onto one 1 PDA plate. All the 3 controls PDA plates were incubated to check contamination<sup>[7]</sup>. Cultural characteristics were noted after 4-7 d of growth on PDA agar plates. The growth was mounted on a slide in lactophenol and microscopic study was carried out using Coslab phase-contrast microscope with photomicrographic attachment. The sequencing of Internal Transcribed Spacer (ITS) region of the fungal cultures was done using the Universal (ITS1 TCCGTAGGTGAACCTGCGG primers and ITS4 TCCTCCGCTTATTGATATGC) at Chromus Biotech or Eurofins Labs. Pvt. Ltd. The ITS sequences were deposited with GenBank,

BLASTN was used for similarity search and phylogenetic tree were constructed by megablast (Blast Tree View) by Neighbour-Joining method. The fungal isolates were identified based on the colony, morphological characteristics and phylogenetic analysis. In the present paper we have investigated the diverse endophytic fungal isolates from the medicinal plant Cassia senna L. an established source of the drug sennosides which is described in various indigenous Indian medicinal systems. Senna leaves are widely used in treating constipation, loss of appetite, dysentery, splenomegaly, hepatomegaly, jaundice, malaria, wound healing, and has purgative, laxative, expectorant, and carminative properties<sup>[8,9]</sup>. We have reported earlier the isolation of endophytic fungi from *Mucuna pruriens*<sup>[7]</sup>. The characteristics of the Cassia senna L plant are shown in fig. 1. 22 endophytic fungal isolates were obtained and identified based on the colony, morphological characteristics as well as phylogenetic analysis (fig. 2 and Table 1). Phylogenetic analysis based on ITS revealed the presence of endophytic fungi belonging to Ascomycota (Aspergillus, Alterneria, Cladosporium, and Fusarium) and one isolate JF5 belonging to Basidiomycota (Phellinus sp.). The phylogenetic tree constructed with Blast Tree view (Neighbour-joining method) with 20 related species for some of the isolates is depicted in fig. 3 and fig. 4. Two of the isolates *Cladosporium* sp. (CBS02) and Alterneria sp. (CBL01) have been deposited with NCMR (NCCS) with the accession nos. MCC 9038 and MCC 9037, respectively. These isolates may have capacity to produce the drug (sennosides) and provide an economical/ sustainable alternative source for the drug. A review on the endophytic fungi as a potential source of antifungal compounds has been recently been published<sup>[10]</sup>. It has been demonstrated that the alkaloid compounds extracted from Cassia senna L. leaves has more effective antibacterial activity as compared to phenolic and terpenoid compounds<sup>[11]</sup>. It has been shown that the methanolic extracts from Cassia senna inhibited the growth of dermatophytes Thrichophyton rubrum and Trichophyton mentagaphytes<sup>[12]</sup>. Studies on 40 endophytic fungi isolated from 10 species of medicinal plants from Palolo, Central Sulawesi has shown antibacterial and antioxidant activity these findings support the view of the pharmaceutical applications of these organisms<sup>[13]</sup>.

25 endophytic fungi have been isolated and identified based on macroscopic and microscopic studies from leaves of 10 medicinal plants from two locations in Surat, India<sup>[14]</sup>. A more extensive study on the diversity of endophytic fungi from 7 medicinal plants of Western Ghats (Maharashtra, India) has been reported by Nalini et al.<sup>[15]</sup>. It was interesting to note that out of the 1525 isolated ascomycetes represented only 3 %, whereas majority of the isolates belonged to coelomycetes (65 %). However, this study did not include medicinal plants belonging to the genus Cassia. An endophytic fungus Fusarium tricinctum was isolated from the roots of Lithospermum officinale L, which produced shikonin (a naphthoquinone with bioactive properties)<sup>[16]</sup>. Thus, the present paper reports the isolation and identification of endophytic fungi from different parts of the medicinal plant Cassia senna L. These isolates may be a useful source for antifungal, antibacterial, anti-cancer, sennosides and other compounds of biotechnological importance. To our knowledge this is the first study on endophytic fungi from this medicinal plant from Gujarat and Rajasthan.

Exploration of endophytic fungi from *Cassia* senna L has benefits both in pharmaceutical and ecological fields. In order to assess the endophytic fungal diversity of this medicinal plant, samples were collected from different regions having diverse eco-geographical conditions in Gujarat and Rajasthan. The 22 fungal isolates obtained belonged to the genera *Alterneria*, *Aspergillus*, *Cladopsorium*, *Fusarium*, *Penicillium*, *Phellinus*, etc. Further studies on the bioprospecting of these isolates would provide new bioactive metabolites including sennosides for biotech industry.



Fig. 1: Different plant parts Cassia senna; A: Aerial parts; B: Branch; C: Flowers; D: Pods; E: Leaves

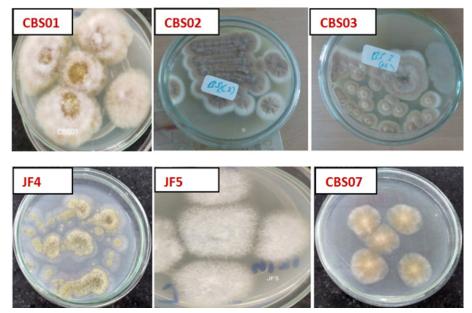


Fig. 2: Colony characteristics of some endophytic fungal isolates from Cassia senna on PDA plate after 5-7 d incubation

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## TABLE 1: LIST OF ENDOPHYTIC FUNGI ISOLATED FROM Cassia senna L. PLANT SAMPLES

S No	Isolates	Morphological identification based on Microscopic study	Identification based on ITS Sequence	GenBank Accession Number	Location / Place
1	CBS01	Fusarium sp.	Fusarium sp.	MN128229	Bhuj 23.2420° N, 69.6669° E
2	CBS02	Cladosporium sp.	Cladosporium sp	MH156215	
3	CBS03	Cladosporium sp.	Cladosporium sp	MN156205	
4	CBS04	Alterneria sp.	Alterneria sp.	MH156196	
5	CBS07	Fusarium sp.	Fusarium sp	MN973665	
6	CBL01	Alterneria sp.	Alterneria brassicicola/ tenuissmia	MH156192	
7	BL1	Aspergillus sp	Aspergillus neoellipticus	MN266885	Boriavi (Anand)
8	BP3	Aspergillus sp	Aspergillus terreus	MN133937	22°35'54.9"N 72°56'00.4"E
9	JF4	Aspergillus sp.	Aspergillus sp	MN128226	Jodhpur 26° 15'49.8"N 72° 58'34.7"E
10	JP2	Fusarium sp.	Fusarium sp	MN266886	
11	JF5	Fusarium sp.	Phellinus caribaeo- quercicola	MT108197	
12	JP3	Aspergillus sp.	Aspergillus caespitosus	MT107145	
13	CBS05	Cladosporium sp.	Nd	NA	Bhuj 23.2420° N, 69.6669° E
14	CBS06	Alterneria sp.	Nd	NA	
15	CBS08	Neurospora sp	Nd	NA	
16	BL2	Aspergillus sp.	Nd	NA	Boriavi (Anand)
17	BP1	Helminthosporium sp.	Nd	NA	22°35'54.9"N 72°56'00.4"E
18	BP2	Fusarium sp.	Nd	NA	
19	BS01	Penicillium sp.	Nd	NA	
20	BS02	Aspergillus sp.	Nd	NA	
21	JF1	Fusarium sp	Nd	NA	Jodhpur 26°15'49.8"N 72°58'34.7"E
22	JP1	Penicillium sp.	Nd	NA	
23	MGS001	Aspergillus sp.	Aspergillus sp.	NA	Ahmedabad 23.0225° N, 72.5714° E

Note: Nd: Not done; NI: Not Identified; NA: Not Applicable

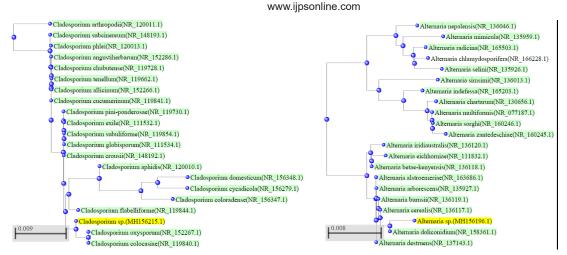


Fig. 3: Phylogenetic trees reconstructed by Blast Tree View (neighbour-joining method) depicting the interrelationships of ITS sequence of the endophytic fungi: CBS02 (MN156215) with *Cladosporium* spp. and CBS04 (MH156196.1) with *Alterneria* spp.

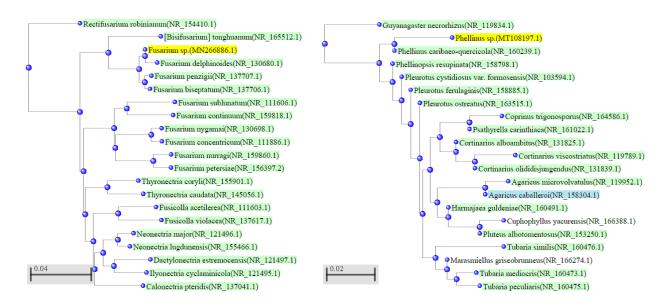


Fig.4: Phylogenetic trees reconstructed by Blast Tree View (neighbour-joining method) depicting the interrelationships of ITS sequence of the endophytic fungi: JP2 (MN266886.1) with *Fusarium* sp. and JF5 (MT108187.1) with *Phellinus* sp. and other basidomycetes

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## **Conflict of interests:**

The authors declared no conflict of interest.

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