# Pharmacognostic Standardization of *Aralia racemosa* L. Stem

## D. S. N. B. K. PRASANTH\*, A. SRINIVASA RAO1 AND Y. R. PRASAD2

Department of Pharmacy, JNTUK, Kakinada-533 003, <sup>1</sup>Department of Pharmaceutical Analysis and Quality Control, Shri Vishnu College of Pharmacy, Vishnupur, Bhimavaram-534 202, <sup>2</sup>Department of Pharmaceutical Chemistry, College of Pharmaceutical Sciences, Andhra University, Visakhapatnam-530 003, India

#### Prasanth, et al.: Pharmacognostic Evaluation of Aralia racemosa L. Stem

The present study was taken up to standardize the stem of *Aralia racemosa* L. for quality, purity and sample identification. Various standardization parameters like morphological, microscopical and physicochemical evaluations, preliminary phytochemical screening and fluorescence analysis of the stem of *A. racemosa* were done and qualitative parameters were reported. Macro and microscopic evaluation studies were performed as per the routine laboratory procedures. Phytochemical, physicochemical, florescence analysis, behaviour of powdered drug have been conducted as per the World Health Organization guidelines. Microscopy of stem revealed the presence of phellem, phellogen, phelloderm, medullary rays usually biserrate and powder analysis showed the presence of cork cells, lignified xylem vessels, xylem fibers and parenchyma cells. Phytochemical analysis showed the presence of saponins, flavonoids, tannins, phenols, glycosides and volatile oil. Physicochemical parameters such as moisture content, ash value, extractive value and fluorescent behaviour of stem powder were determined. These parameters are useful tools to differentiate the powdered drug material. The findings of this study will be helpful in the identification of *A. racemosa* stem.

# Key words: Pharmacognostic studies, *Aralia racemosa* L., phytochemical analysis, physicochemical parameters, fluorescence

In accordance with World Health Organization (WHO); traditional medicines are utilized by 70-95% of worldwide population especially in developing countries with regard to their healthcare. Additionally, the usage of herbal drugs has grown incredibly in line with the worldwide trend of folks returning to natural remedies. The increasing utilization of botanicals by the public is forcing moves to evaluate the health claims of these agents and also to establish specifications of quality and manufacture<sup>[1]</sup>.

Herbal drugs exert a key impact in health care programs particularly in developing nations. However hindrance behind the acceptance of herbal medicine in developed nations is the lack of documentation, stringent quality control measures. Therefore, documentation and standardization of the crude materials utilized in herbal medicine is vital for the global compliance of this system of medicine. In this regard, WHO supports the nations to provide conventional remedies and practices in communal and private health services. Construction of monographs are planned fundamentally to promote harmonization in the utilization of herbal drugs with respect to levels of safety, efficacy, and quality control. Pharmacognostic standardization, physicochemical investigation and preliminary phytochemical studies were all over accepted in recognizable proof and confirmation of the genuine plant materials. Standardization of herbal drugs is the strategy of recommending a set of specifications, constant parameters, definitive qualitative and quantitative values, which bring an assurance of quality, efficiency, safety as well as reproducibility. Proper identification and quality assurance of plant materials are essential to ensure reproducible result of herbal medicines, which will add to its safety and efficacy. Pharmacognostic standardization of plant material comprises of

Accepted 05 March 2017 Revised 10 December 2016 Received 03 August 2016 Indian J Pharm Sci 2017;79(2):220-226

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms

its morphological, anatomical and biochemical characteristics<sup>[2]</sup>.

Araliaceae is an extensive family consisting of 50 genera and 1412 species. The plants of this family have a significant contribution in the treatment of respiratory inflammation, diabetes, cancer and parasitic infections<sup>[3]</sup>. The genus Aralia of Araliaceae contains up to 71 species of plants scattered over Asia, North America and South America, a few examples of these species are Aralia armata, A. bipinnata, A. chinensis, A. continentalis, A. cordata, A. dasyphylla, A. echinocaulis, A. elata, A. fargensii and A. nudicaulis. A. racemosa, a perennial herb in this family is distributed in America, Africa, Australia, New Zealand and Pacific Islands<sup>[4]</sup>. The genus Aralia is very closely related to the genus Panax, which consists of Asian and American ginseng (Panax ginseng and P. quinquefolius). Various plants in the genus Aralia are utilized medicinally for different disorders. There are actually hundreds of patents associated to medicinal products that contain plants of genus Aralia<sup>[3]</sup>.

One genus of Aralia found in India i.e. A. racemosa. It is commonly known as American Spikenard. A. racemosa is an aromatic perennial reaches a height of 2 to 5 feet. Root large, thick, spicy-aromatic, bark thick, whitish internally. Stem ligneously herbaceous, smooth, bifurcating, much branched and devoid of prickles. Leaves very large, odd pinnately compound; leaflets ovate to cordate, doubly serrate, acuminate and slightly downy; stipules represented by a serrate stipular membrane at the bifurcation of branches and sometimes at the base of the petiole. Inflorescence numerous axillary, compound; racemose panicles or thyrsi; flowers monoeciously polygamous. Calyx 5 coherent with the ovary; petals 5 epigynous, obovate, acute, reflexed- spreading, caducous; stamens 5, epigynous, situated opposite to the calyx; filaments slender; another is 2-celled opening longitudinally. Ovary globular, 5-celled, somewhat 10-ridged; ovules anatropous suspended one in each cell; styles 5 closely clustered, sometimes united at the base; stigmas capitellate. Fruit globular, aromatic, baccate drupes<sup>[5]</sup>. The genus Aralia is rich in triterpenoid saponins chemically, which exhibit antiinflammatory, antidiabetic, antihyperlipidemic, hepatoprotective and antiproliferative activities<sup>[3]</sup>. In traditional system of medicine various parts of A. racemosa are used in the treatment of rheumatism, whooping cough, skin diseases, pleurisy, pulmonary diseases, asthma, diarrhoea, stimulant, expectorant, syphilis, inflammation and hay fever<sup>[6,7]</sup>. Various

pharmacological activities of *A. racemosa* have been reported such as antioxidant, antidiabetic<sup>[8,9]</sup>, antitubercular<sup>[10]</sup> and antibacterial<sup>[11]</sup>. The genus *Aralia* is rich in acetylenic lipids, triterpenoid saponins, steroid saponins, phytosterols and triterpene carboxylic acid. Phytochemical investigation on *A. racemosa* revealed the presence of triterpenoid saponins i.e., oleanolic acid, sterols i.e.,  $\beta$ -sitosterol and diterpenoids i.e., entkaurenoic acid, continentalic acid, acanthoic acid<sup>[3]</sup>. Hence, in this work we attempted to standardize *A. racemosa* to study the morphological, anatomical, physicochemical and preliminary phytochemical analysis of stem was done as prescribed in WHO guidelines.

# **MATERIALS AND METHODS**

*A. racemosa* plant was procured in the month of September 2015 from Sri Venkateswara University, Andhra Pradesh. It was identified and authenticated in the Department of Botany, Sri Venkateswara University, Tirupati, Andhra Pradesh and a voucher specimen of the plant (No 1489) was deposited at the herbarium of Sri Venkateswara University, Tirupati, for future reference.

## **Preparation of extract:**

Stem of *A. racemosa* was shade dried and powdered using an electric mixer-grinder and sieved through sieve no. 22 with an average aperture size of 710  $\mu$ m. Then coarsely powdered leaves were macerated with solvents in the increasing order of polarity, chloroform, methanol and water. After 24 h, the extracts were filtered through a Whatman filter paper and concentrated in a rotary evaporator followed by removal of the residual moisture in a desiccator.

# Macroscopic and microscopic analysis:

Macroscopic characteristics of the plant were analysed by WHO quality control methods of herbal medicine<sup>[12]</sup>. Stem were cut into thin sections manually with the sharp cutting edge of a blade using the free hand. The sections were then transferred to slides, cleared by warming with chloral hydrate to remove chlorophyll, stained with phloroglucinol and concentrated HCl to detect the lignified tissues and mounted in glycerin. Powder microscopy was performed according to the method of Khandelwal<sup>[13]</sup>. The photomicrographs of different magnifications were taken with Olympus CX-21i triocular microscope under illumination of a halogen lamp.

### **Physicochemical parameters:**

Physicochemical parameters such as ash value, moisture content and extractive values were determined according to the procedures mentioned in WHO quality control methods for herbal materials as follows.

#### **Determination of loss on drying:**

About 10 g of powdered drug (without preliminary drying) was weighed and placed in a tared evaporating dish and dried at 105°. The drying and weighing were being carried out at 1 h intervals till the variance between two successive weighing's had not been more than 0.25%. A consistent weight was supposed to have reached when two successive weighing after drying for 30 min and cooling for 30 min in a desiccator, showed not more than 0.01 g difference<sup>[14,15]</sup>.

#### Determination of total ash:

About 2.0 g of the powdered drug was weighed accurately and incinerated in a silica crucible at a temperature not exceeding 450° until free from carbon. The resulting ash was cooled and weighed. The process was repeated to obtain a constant weight. The percent of total ash with reference to the air-dried drug was calculated<sup>[14,15]</sup>.

#### Water soluble ash:

The ash was obtained according to the method defined above, boiled for 5 min with 25 ml of water, filtered and the insoluble matter was collected on an ash less filter paper. It was additionally washed with hot water and inflamed for about 15 min at a temperature not exceeding 450° and weighed. The difference in weight of inflamed and total ash represented the water-soluble ash. The percent water soluble ash was determined with reference to the air-dried drug<sup>[14,15]</sup>.

#### Determination of acid insoluble ash:

Ash was obtained according to the method described above, boiled for 5 min with 25 ml of 2 M hydrochloric acid, filtered and insoluble matter was collected on an ash less filter paper. It was further washed with hot water and ignited for 15 min at a temperature not exceeding  $450^{\circ}$  and weighed. The percent acid insoluble ash was calculated with reference to the air-dried drug<sup>[14,15]</sup>.

#### Water soluble extractive value:

Four grams of the air-dried coarsely powdered drug was macerated with 100 ml of water in a closed flask for 24 h, shaken repeatedly during first 6 h and then allowed to stand for 18 h. It was filtered; 25 ml of filtrate was evaporated in a flat shallow dish, dried at 105° and weighed. Percent water-soluble extractive value was calculated with reference to air-dried drug<sup>[14,15]</sup>.

#### Methanol soluble extractive:

Four grams of the air-dried coarsely powdered drug was macerated with 100 ml of ethanol in a closed flask for 24 h, shaken frequently during first 6 h and allowed to stand for 18 h. Then it was filtered, during filtration precaution was taken against loss of methanol; 25 ml of filtrate was evaporated in a flat shallow dish and dried at 105° and weighed. Percent methanol soluble extractive value was calculated with reference to air-dried drug<sup>[14,15]</sup>.

### **Qualitative phytochemical screening:**

Various extracts of A. racemosa were subjected to qualitative chemical analysis using standard procedures. Carbohydrates were detected using the Molisch's test, proteins were detected using either Biuret test or Millon's test and amino acids using Ninhydrin test. Steroids were detected by Salkowski, Libermann-Burchards and Libermann's tests, alkaloids were identified with freshly prepared Dragendroff's, Mayer's, Hager's and Wagner's reagents by the presence of turbidity or precipitation. The flavonoids were detected using four tests, Shinoda, sulphuric acid, aluminium chloride, lead acetate and sodium hydroxide. Tannins were detected with four tests that included gelatin, lead acetate, potassium dichromate and ferric chloride. The froth, emulsion, and lead acetate tests were applied for the detection of saponins. Steroids were detected using a combination of acetic anhydride with sulphuric acid or acetic chloride with sulphuric acid. Sample extracted with chloroform was treated with sulphuric acid to test for the presence of terpenoids. Ammonia solution and ferric chloride solutions were used to detect the presence of anthraquionones<sup>[14-21]</sup>.

#### Fluorescence analysis of the powdered drug:

The fluorescence examination of the powdered leaves was done by placing dry powdered leaves on a slide and observed by treating with a few drops of different chemical reagents to detect the colour changes under UV/Vis light<sup>[21,22]</sup>.

#### **RESULTS AND DISCUSSION**

The stem of *A. racemosa* has greyish white ting. The outer stem is greyish white with yellow white inside,

stem has aromatic odour and acrid in taste. The transverse section of stem of A. racemosa showed the presence of periderm, cortex, phloem and xylem. Periderm shows the presence of tangentially elongated cork (phellem), which is non-lignified in nature, followed by phellogen, which are non-living rectangular cells made up of cellulose. The inner most layer is phelloderm, which are square shaped cells composed of cellulose and living in nature. All these layers collectively called as periderm as shown in fig. 1. The cortex region is composed of parenchymatous cells, with small intercellular spaces, which is multi-layered with 10-15 layers. The phloem is sandwiched between two medullary rays as shown in fig. 2. The medullary rays are parenchymatous in nature, which are mono- to tri-serrate, but mostly biserrate. The phloem also shows the presence of phloem parenchyma and sieve tubes. Xylem is surrounded by bi-serrate lignified medullary rays as shown in fig. 3. Xylem also comprises of xylem fibres, xylem vessels and xylem parenchyma. Beneath the xylem is pith or medulla as shown in fig. 4. Powder microscopy of stem of A. racemosa revealed the presence of cork cells, parenchyma cells, lignified xylem fibres and xylem vessels as showed in fig. 5.

The total ash, acid insoluble ash and water soluble ash were found to be  $6.36\pm0.02$ ,  $0.36\pm0.03$  and  $0.83\pm0.02$ , respectively. However the extractive values of petroleum ether, chloroform, methanol and water were found to be  $1.8\pm0.23$ ,  $3.4\pm0.32$ ,  $12.8\pm0.22$  and  $16.3\pm0.32$  as showed in Table 1. Preliminary phytochemical analysis revealed the presence of saponins, flavonoids, tannins, phenols, glycosides and volatile oil as showed in Table

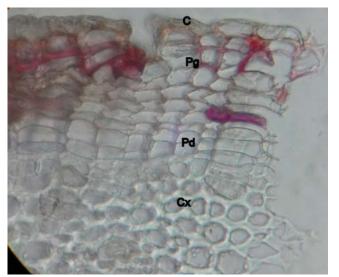


Fig. 1: Transverse section of *A. racemosa* stem Transverse section of *A. racemosa* stem showing the cork and cortex regions. C: cork; Pg: phellogen; Pd: phelloderm; Cx: cortex

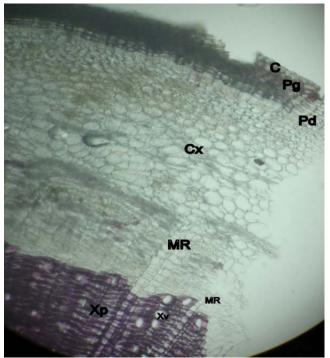


Fig. 2: Transverse section of *A. racemosa* L stem C: cork, Pg: phellogen; Pd: phelloderm; Cx: cortex; MR: medullary rays; Xv: xylem vessels; Xp: xylem parenchyma

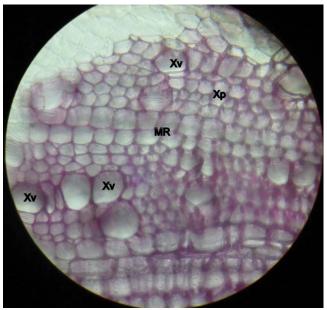


Fig. 3: Transverse section of *A. racemosa* L stem showed xylem region

Xv: xylem vessels; MR: medullary rays; Xp: xylem parenchyma

2. Fluorescence characteristics of powdered stem of *A*. *racemosa* was observed in visible, short and long UV light as showed in Table 3.

Indian systems of medicine utilize most of the crude drugs, which are of plant origin. It can be crucial that specifications need to be set down to control as well as check the identity of the plant and examine its

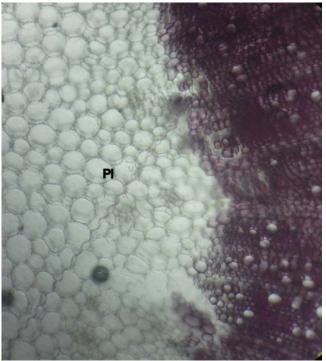


Fig. 4: Transverse section of *A. racemosa* L stems showing pith. PI: pith

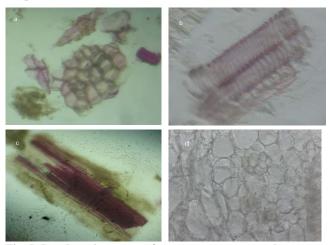


Fig. 5: Powder microscopy of *A. racemosa* stem powder (a) Cork cells (b) lignified xylem vessels (c) xylem fibres (d) parenchyma

quality prior its utilization. A detailed pharmacognostic assessment therefore is extremely an important prerequisite. In accordance with WHO, the macroscopic and microscopic description of a medicinal plant is the primitive step in the direction of establishing its identity and purity and need to be performed prior to any tests tend to be carried out<sup>[21-23]</sup>.

*A. racemosa*, traditionally utilized in conventional medicines has incredible therapeutically potential due to its multifaceted biological functions. Microscopy is one of the straight forward and modest strategies to begin with setting up the correct identity of the source

# TABLE 1: PHYSICOCHEMICAL PARAMETERS OF STEM POWDER OF *A. RACEMOSA*

Parameters	Values (% w/w)
Moisture content (loss on drying)	10±0.10
Total ash	6.36±0.02
Acid insoluble ash	0.36±0.03
Water soluble ash	0.83±0.02
Petroleum ether soluble extractive value	1.8±0.23
Chloroform soluble extractive value	3.4±0.32
Methanol soluble extractive value	12.8±0.22
Water soluble extractive value	16.3±0.32

materials. The prominent diagnostic features of stem were cork cells, lignified xylem vessels, parenchyma, medullary rays and xylem fibres. These characteristics could be utilized for standardization of drugs and also useful for preparation of plant monographs. Ash values have been widely used to establish quality and purity of crude drugs. This is indicative of the presence of various impurities such as carbonate, oxalate and silicate. The water soluble ash is employed to estimate the amount of inorganic compound found in drugs. The acid insoluble ash consisted chiefly of silica and indicated contamination with earthy substances. The moisture content of a crude drug ought to be minimized with a specific goal to avoid decomposition of crude drugs either due to chemical changes or microbial contamination. The crude drugs exert pharmacological activities principally because of active chemical constituents present. These constituents might be soluble in various polar, semi polar or nonpolar solvents. Total soluble constituents of the crude drug in a specific solvent or mixture of solvents constitute the extractive value. These extractive values are expected to give a measure of the chemical constituents present. The percent active chemical constituents in any crude drug are expressed on air-dried basis. Thus the fluorescence analysis and behaviour of powdered drug with different chemical reagents are extremely fast techniques to distinguish from suspicious specimens. In case of absence of physicochemical assessment, such techniques are critical to check the adulteration. This approach of standardization absolutely gives scientific validation of crude drugs<sup>[21-23]</sup>. Here, preliminary phytochemical examination of the stem revealed the presence of saponins, flavonoids, tannins, phenols, glycosides and volatile oils.

Since there has been no reports documenting the pharmacognostical work on this traditionally significant crude drug, the current investigation had

#### www.ijpsonline.com

#### TABLE 2: PHYTOCHEMICAL ANALYSIS OF VARIOUS EXTRACTS OF A. RACEMOSA STEM

Phytoconstituents	Method	Aqueous extract	Methanol extract	Chloroform extract
Flavonoids	Shinoda test	+	+	-
	Zinc hydrochloride test	+	+	-
	Lead acetate test	+	+	-
Volatile oil	Stain test	-	+	-
Alkaloids	Wagner test	-	-	-
	Hager's test	-	-	-
Tannins and phenols	FeCl <sub>3</sub> test	+	+	+
	Potassium dichromate test	+	+	+
Saponins	Foaming test	+	+	-
Steroids	Salkowski test	+	+	-
Carbohydrates	Molish test	-	-	-
Acid compounds	Litmus test	-	-	-
Glycoside	Keller-Killani test	+	+	-
Amino acids	Ninhydrin test	-	-	-
Proteins	Biuret test	-	-	-

+ Present; - absent

#### TABLE 3: FLUORESCENCE ANALYSIS OF A. RACEMOSA STEM POWDER

Solvent used	Visible light -	UV light		
Solvent used		At 254 nm	At 366 nm	
Distilled water	Greyish white	Black	Blue	
1 N NaOH 1 N methanol	Greyish white	Black	Dark blue	
1 N HCl	Greyish white	Green	Blue	
50% HNO <sub>3</sub>	Greyish white	Black	Brown	
50% H <sub>2</sub> SO <sub>4</sub>	Greyish white	Black	Brown	

been taken up with a view to lay down standards, that could be helpful for identification of sample quality, purity, authentication and also compendium of quality control of crude drugs<sup>[21,23]</sup>.

#### **Acknowledgements:**

The authors thank Dr. K. Madhavachetty, plant taxonomist, Department of Botany, Sri Venkateswara University, Tirupati, Andhra Pradesh, for identifying and authenticating the plant material and also thank to Dr. A. Lakshmana Rao, Principal and Management of V. V. Institute of Pharmaceutical Sciences, Gudlavalleru, Andhra Pradesh, for providing the research facilities.

#### **Conflict of interests:**

Authors declare no conflict of interests.

#### Financial support and sponsorship:

Nil.

#### REFERENCES

 Mukhi S, Bose A, Panda P, Rao MM. Pharmacognostic, physicochemical and chromatographic characterization of Samasharkara Churna. J Ayurveda Integr Med 2016;7:88-99.

- Sreedhar S, Prakash Kumar U, Girija TP, Remashree AB. Pharmacognostic standardisation of *Combretum albidum* G. Don leaf; a medicinally important liana. Phcog J 2013;5:247-55.
- 3. Clement JA, Clement ES. The Medicinal Chemistry of Genus *Aralia*. Curr Top Med Chem 2014;14:2783-801.
- Wen J. Systematics and Biogeography of Aralia L. (Araliaceae): Revision of Aralia Sects. Aralia, Humiles, Nanae, and Sciadodendron. United States National Herbarium 2011;57:1-172.
- Frederick MC. American Medicinal Plants: An illustrated and descriptive guide to plants indigenous to and naturalized in the United States which are used in medicine. New York: Dover Publications; 1887. p. 272-75.
- 6. Umberto Q. CRC World Dictionary of Medicinal and Poisonous Plants: Common names, scientific names, eponyms, synonyms and etymology. London (UK): CRC Press; 2012. p. 353.
- Duke JA, Bogenschutz-Godwin MJ, duCellier J, Duke PK. Handbook of Medicinal Herbs. Boca Raton (US): CRC press; 2002. p. 691.
- McCune LM, Johns T. Antioxidant activity in medicinal plants associated with the symptoms of diabetes mellitus used by the indigenous peoples of the North American boreal forest. J Ethnopharmacol 2002;82:197-205.
- 9. McCune LM, Johns T. Symptom-specific antioxidant activity of boreal diabetes treatments. Pharm Biol 2003;41:362-70.
- 10. Grange JM, Davey RW. Detection of antituberculous activity in plant extracts. J Appl Bacteriol 1990;68:587-91.
- 11. Manpreet K, Harinder K. Evaluation of Antibacterial

Activity of roots of *Aralia racemosa*. Res J Pharm Technol 2013;6:1357-58.

- 12. h t t p : / / a p p s . w h o . i n t / i r i s / bitstream/10665/41986/1/9241545100.pdf.
- 13. Khandelwal KR. Practical pharmacognosy techniques and experiments. 19th ed. New Delhi: Nirali Prakashan; 2002.
- 14. Alam F, Najum us Saqib Q. Pharmacognostic standardization and preliminary phytochemical studies of *Gaultheria trichophylla*. Pharm Biol 2015;53:1711-8.
- Singh P, Khosa RL, Mishra G, Jha KK. Pharmacognostical evaluation of aerial parts of *Graptophyllum pictum* (L.) Griff. (Syn: *Justicia picta* Linn.): A well-known folklore medicinal plant. Anc Sci Life 2015;34:223-9.
- 16. Roy S, Acharya RN, Harisha CR, Shukla VJ. Macro, microscopic and preliminary analytical evaluation of root and leaf of *Globba marantina* Linn. an extrapharmacopoeial drug of Ayurveda. Indian J Pharm Sci 2016;78:469-78.
- 17. Raman N. Phytochemical Techniques. New Delhi: New India Publishing Agency; 2006.

- 18. Khandelwal KR. Practical pharmacognosy techniques and experiments. 19th ed. New Delhi: Nirali Prakashan; 2002.
- 19. Kokate CK. Practical Pharmacognosy. 1st ed. New Delhi: Vallabh Prakashan; 2005.
- 20. Harborne JB. Phytochemical methods a guide to modern techniques of plant analysis. 2nd ed. USA: Springer; 1984.
- 21. Patil PS, Venkatanarayanan R, Argade PD, Shinde PR. Assessment of pharmacognostic and phytochemical standards of *Thespesia populnea* (L.) root. Asian Pac J Trop Biomed 2012;2:S1212-S16.
- Abhimanyu KK, Ravindra CS, Avanapu RS. Application of Digital (Motic) and Scanning electron microscope in histological study of leaf of *Tecoma gaudichaudi* DC (Bignoniaceae). J Basic Appl Sci 2016;5:97-101.
- 23. Kalpna Rakholiya, Sumitra Chanda. Pharmacognostic, physicochemical and phytochemical investigation of *Mangifera indica* L. var. kesar leaf. Asian Pac J Trop Biomed 2012;2:S680-S4.