Polyelectrolyte Complexes: A Review of their Applicability in Drug Delivery Technology

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Lankalapalli and Kolapalli: Polyelectrolyte Complexes and Drug Delivery Technology

Over the past several years, great advances have been made towards novel drug delivery systems. The phenomena of interpolymer interactions and formation of polyelectrolyte complexes have been the focus of intensive fundamental and applied research. Interpolyelectrolyte complexes combine unique physicochemical properties with high biocompatibility. Studies have been carried out on many different polymer blends and types. Such combinations may possess unique properties that are different from those of individual component. The present review emphasizes on the applicability of polyelectrolyte complexes in drug delivery technology.

Key words: Polyelectrolyte complex, polymer, drug delivery, polyion, electrostatic interaction

Current state of art is witnessing a revolution in new techniques for drug delivery. These techniques are capable of controlling the rate of drug delivery, sustaining the duration of therapeutic activity and/ or targeting the drug to specific tissues. These advancements led to the development of several novel drug delivery systems, revolutionizing the medication with several advantages.

Recent decades witnessed the appearance of polymers that respond in some desired way to changes in temperature, pH, electric or magnetic field. The driving force behind these transitions include stimuli like neutralization of charged groups by either a pH shift or the addition of an oppositely charged polymer, changes in the efficiency of hydrogen bonding with an increase in temperature or ionic strength and collapse of hydrogels and interpenetration of polymer network. These types of polymers not only convert the active substances into a non-deleterious form which can be administered, but also have specific effect on the biodistribution, bioavailability or absorption of the active substances and hence increasingly gaining importance in modern pharmaceutical technology. The interaction between two oppositely charged polymers results in the formation of a complex, termed as

polyelectrolyte complex^[1]. These polyelectrolyte complexes meet the profile of requirements of biocompatible polymer systems and can be adapted to meet the various requirements like carrier substances and components for active substances.

Polyelectrolyte complexes (PECs) are the association complexes formed between oppositely charged particles (e.g. polymer-polymer, polymer-drug and polymer-drug-polymer). These are formed due to electrostatic interaction between oppositely charged polyions. This avoids the use of chemical cross linking agents, thereby reducing the possible toxicity and other undesirable effects of the reagents. The polyelectrolyte complexes formed between a poly acid and poly base are little affected by the pH variation of the dissolution medium. This concept of complexation, between DNA and chitosan^[2], has extensively been studied in the development of delivery vehicle for gene therapy and oral vaccination.

The occurrences of charge-charge interactions between ionic polymers and drugs were considered to be a negative event when the ionic polymers are used as excipients in pharmaceutical formulations. In these systems release of drugs may be strongly affected by the occurrence of charge-charge interactions. However, in recent years these negative events of polymer-drug and polymer-polymer interactions have been exploited positively for controlled drug release^[3,4].

Polyelectrolytes:

The polymers that contain a net negative or positive charge at near neutral pH are called polyelectrolytes^[5]. They are generally soluble in water. Their solubility is driven by the electrostatic interactions between water and the charged monomer. Examples of such polymers include DNA, protein, certain derivatives of cellulose polymers and carragenan.

Classification of polyelectrolytes^[6]:

The polyelectrolytes are classified into various types. Based on origin they are classified as natural polyelectrolytes, synthetic polyelectrolytes and chemically modified biopolymers. Based on composition they are homopolymers and copolymers. Based on molecular architecture linear, branched and cross linked. Based on electrochemistry they are classified as polyacids/polyanions, polybases/polycations and polyampholytes. Some of the important polyelectrolytes are exemplified in Table 1.

Theoretical aspects of PECs:

Many researchers extensively investigated the properties of the polyelectrolytes^[6,7] and the formation of PECs^[8-10]. There have been theories proposed based on the electrostatic forces and Flory-Huggins mixing

TABLE 1: SOME OF THE IMPORTANT
POLYELECTROLYTES

Name	Category	
	(based on the	
	charge type)	
Natural Polyelectrolytes		
Nucleic acids	Polyanion	
Poly (L-lysine)	Polycation	
Poly (L-glutamic acid)	Polyanion	
Carrageenan	Polyanion	
Alginates	Polyanion	
Hyaluronic acid	Polyanion	
Chemically modified biopolymers		
Pectin	Polyanion	
Chitosan (deacetylation of chitin)	Polycation	
Cellulose - based	Polyanion or polycation	
Starch - based	Polyanion or polycation	
Dextran - based	Polyanion or polycation	
Synthetic polyelectrolytes		
Poly (vinylbenzyl trialkyl ammonium)	Polycation	
Poly (4-vinyl-N-alkyl-pyridimiun)	Polycation	
Poly (acryloyl-oxyalkyl-trialkyl ammonium)	Polycation	
Poly (acryamidoalkyl-trialkyl ammonium)	Polycation	
Poly (diallydimethyl-ammonium)	Polycation	
Poly (styrenesulfonic acid)	Polyanion	
Poly (vinylsulfonic acid0	Polyanion	
Poly (acrylic or methacrylic acid)	Polyanion	
Poly (itaconic acid)	Polyanion	
Maleic acid/ diallyamine copolymer	Polyampholytic	

free energies of the polyelectrolytes to explain the mechanism of formation of PECs^[11-13]. In general the backbones of the two polymers are not compatible and repel to each other, however, the charge fraction of the polymers determines the type of interaction going to occur between the polymers. When the charge fraction is low, the polymer backbone repulsion (Flory interaction parameter) is dominant and the solution separates in to two phases each containing mostly one of the polymers. At high charge fraction, the attractive electrostatic interactions between the polymers dominate and they precipitate to form a complex. In an intermediate range of charge fraction, the equilibrium state can be a meso phase where the two polymers only separate microscopically. Depending on the stoichiometry of the mixture (the relative concentrations, the relative chain lengths and charge densities), one observes mainly two types of complex formations, a macroscopic phase separation between the solvent and the polymers or a partial aggregation of the polymer chains^[14].

Formation of PECs:

This process involves mainly 3 steps as shown in fig. 1^[15]. First step is primary complex formation and Coulomb forces are responsible for this step. Second step is formation process within intracomplexes. It involves formation of new bonds and/or the correction of the distortions of the polymer chains. Third is intercomplex aggregation process, which involves the aggregation of secondary complexes mainly through hydrophobic interactions.

Factors affecting the formation of PECs:

A number of parameters are known to influence the



Fig. 1: Schematic representation of the formation and aggregation of PECs

(a) Primary complex formation (b) Formation process within intracomplexes (c) Inter complex aggregation process

formation of PECs^[16]. These are ion site, charge density, polyelectrolyte concentration, pH, ionic strength, solvents and temperature. Several workers evaluated the factors effecting the formation of polyelectrolyte complexes with different polymeric blends^[17,18]. Precipitation is caused by effective attractions due to charge fluctuations and by short range attractions between monomers. Interesting in the context of the polyion stoichiometry are studies of layer formation from strongly asymmetric pairs of polyions. Employing polyions with a reduced charge density along the chain, i.e. consisting of charged and uncharged co-monomers, it was observed that a minimum charge density^[19] is required for polyelectrolyte adsorption. Changing the ionic strength by addition of salt^[20] can modulate the electrostatic interactions in a polyelectrolyte solution. The electrostatic interactions can be weakened by addition of inorganic salts into the solutions. Thus, an increase of the ionic strength of the solution depresses the complexation between polyions, because of the screening of opposite charges of the macromolecules by low molecular weight ions. By varying the pH environment during PEC formation, the degree of ionization of weak polyelectrolytes can be controlled^[21,22]. This was found to affect multilayer properties such as layer thickness, the degree of interpenetration between layers, surface wettability and number of unbound functional groups. Therefore, by choosing the right pH conditions, a platform may be found with properties that are advantageous for loading charged small molecules into the film via electrostatic interactions.

Characterization of PECs:

Various methods have been used to investigate interactions between polymers^[23]. Measurements of

turbidity, pH and ionic strength^[24,25] as a function of weight ratio of polymer in the media^[26], viscosity^[27], light scattering^[28,29], infrared spectroscopy, NMR, thermal analysis, pK_a and powder X-ray diffraction^[30] were employed to evaluate interpolymer complexation.

Applications of PECs:

PECs have gained much attention in the past few years because of their potential applications. These can be used as membranes^[31-33], for coating on films and fibers^[34], for isolation and fractionation of proteins^[35,36], for isolation of nucleic acid^[37-39], for binding pharmaceutical products^[40], as supports for catalyst^[41] and for preparation of microcapsules for drug delivery^[42,43]. Many of the applications are based on the functional properties of the polyelectrolyte. The functional applications of PECs are summarized in Table 2.

The concept of PECs in the design of drug delivery systems may be useful due to the advancements made during the last two decades^[44-47]. The active components will be encapsulated in the polymer matrix at molecular level. They offer greater advantages for the drug substances through improving/ and altering physicochemical characters like stability and dissolution. The active substance can be incorporated in to PECs by four ways^[48]. In the first case the active substance will be entrapped from the solution during precipitation of the complex. The active substance will be absorbed from the solution and gets incorporated in to the already formed complex on contact in the second case. In the third case the active substance may be chemically bound to at least one complex partner and precipitates during complexation. In the last case the active compound itself may act as poly ion and form PEC. The active

TABLE 2: PRACTICAL API	PLICATION OF POLYEL	LECTROLYTE COMPLEXES
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Functional property	Application
Interaction with counter ions	Support of filtration process - removal of counter ions ^[61,62]
	Gelation process - bridging with multivalent counter ions ^[63]
	Analytical methods - counter ion exchange ^[64]
Interaction with surfactants	Insoluble polyion surfactant complex for low energy surface modification ^[65]
	Highly ordered structures (micelle) formation ^[66,67]
Interaction with charged low molar mass molecules	Polyion drug complexes - soluble or insoluble ^[68]
Interaction with charged particles	Floculation - waste water treatment ^[69]
	Dewatering - sludge, pulp and paper production ^[70]
	Flotation - mining
	Retention - paper production
Interaction with charged surfaces	Displacement chromatography - separation and concentration of biomolecules ^[38]
	Modification of surfaces and interfaces - coating (antistatic, sensors,
	multi-layer) ^[71,72]
	Additive (cosmetics, detergents)

substance from these PECs will be released either by solution equilibration or by ion exchange mechanism or by charge interaction and slow decomplexation as well as breakdown and dissolution of the complex.

There are several reports on different methods of preparation and applications of PECs in pharmacy. Kawashima *et al.*^[49] developed a novel method for the preparation of theophylline granules coated with a PEC of sodium tripolyphosphate and chitosan. The theophylline granules containing sodium tripolyphosphate were stirred in an HCl solution of chitosan. During the mixing, the dissolved sodium tripolyphosphate in the granules moved to the surface and reacted with the chitosan, resulting in the formation of a PEC film. The drug-release pattern of the coated granules followed zero-order kinetics and the release rates were significantly reduced compared with that of the original granules.

Shiraishi *et al.*^[50] studied the controlled drug release behaviour of indomethacin by chitosan-PEC. They also optimized the formulation conditions and reported its *in-vivo/in-vitro* evaluation studies. They prepared the PEC of indomethacin by using complexation of sodium tripolyphosphate and chitosan. Here the effects of the molecular weights of chitosan hydrolysates on the release and absorption rates of indomethacin from gel beads were examined. The release rates of indomethacin decreased with increasing of molecular weight and indomethacin content. A negative correlation was observed between the molecular weight and release rate constant (r=-0.983).

Jimenez-Kairuz et al.^[51] developed and characterized swellable drug-polyelectrolyte matrices (SDPM) using carbomer and different basic drugs like atenelol, lidocaine and metoclopramide. The authors concluded that drugs can be loaded in a high proportion on to the polymer and therefore the resulting SDPM material could be diluted with other polymers to modulate delivery properties of SDPM. Matrices of atenolol and lidocaine exhibited robust delivery properties with regard to change in proportion of loading drug. Liao et al.^[52] prepared drug-loaded chitosan-alginate fibers by interfacial polyelectrolyte complexation technique. Depending on the component properties, the release time of encapsulated components from these fibers could range from hours to weeks. Dexamethasone was completely released within 2 h, whereas charged compounds such as

bovine serum albumin, PDGF-bb, and avidin showed sustained release for 3 w. In this study, interfacial polyelectrolyte complexation demonstrated to be a promising technique for producing drug-loaded fibers with high encapsulation efficiency, sustained release kinetics, and capacity to retain the bioactivity of the encapsulants.

Tapia *et al.*^[53] evaluated the possibility of using mixtures of PECs from both chitosan (CS)-alginate and CS-carrageenan as prolonged release systems. Different dissolution profiles for diltiazem clorhydrate were obtained by changing the polymer matrix system (CS-alginate or CS-carrageenan) and the method used to include these polymers into the formulation (physical mixture or PEC). Drug dissolution profiles from the matrices have been discussed by considering the swelling behavior of the polymers used. They reported that CS-alginate systems were considered to be better in prolonging the release when compared to CS-carrageenan systems.

Paloma et al.^[54] prepared polyionic complexes of CS and poly(acrylic acid) (PAA) in a wide range of copolymer composition and with two kinds of drugs. Release of amoxicillin trihydrate and amoxicillin sodium from these different complexes was studied. The swelling behavior of and solute transport in swellable hydrogels were investigated to check the effect of polymer/polymer and polymer/drugs interactions. The electrostatic polymer/polymer interactions took place between the cationic groups from CS and the anionic ones from PAA. The diffusion of amoxicillin trihydrate was controlled only by the swelling/eroding ratio of the polyionic complexes. The swelling degree of amoxicillin sodium hydrogels was more extensive when compared to the swelling degree of amoxicillin trihydrate formulations. It was concluded that the water uptake was mainly governed by the degree of ionization. Restriction of amoxicillin sodium diffusion could be achieved by polymer/ionized-drug interaction that retards the drug release. Win et al.[55] developed PEC gel beads based on phosphorylated chitosan (PCS) for controlled release of ibuprofen in oral administration. The PCS gel beads were prepared from soluble phosphorylated chitosan by using an ionotropic gelation with counter polyanion, tripolyphosphate (TPP) at pH 4. Surface morphology studies for the prepared beads were done by using SEM. The percentage release of ibuprofen from PCS gel beads was found to be increased as

the pH of the dissolution medium increased. The release rate of ibuprofen at pH 7.4 was higher than the release rate at pH 1.4 due to the ionization of phosphate group and higher solubility of ibuprofen at pH 7.4 medium. The ability of the prepared copolymer to be used as drug carrier for colon-specific drug delivery system was estimated using ketoprofen as model drug.

Albeno *et al.*^[56] obtained a patent for preparation of stable water insoluble complexes of poorly soluble compounds molecularly dispersed in water insoluble ionic polymers. The compounds were micro precipitated in the ionic polymers in amorphous form. The complexes according to the present invention significantly increased the bioavailability of poorly soluble therapeutically active compounds.

Rolfes et al.^[57] reported a method of making a solid interpolymer complex for use as a controlled release matrix for oral administration. The process involved mixing of two oppositely charged polymers and spray-dried to evaporate the solvent and to prepare solid particles of interpolymer complex. An active agent such as drug can be preferably embedded or encapsulated in the interpolymer complex before spray drying or may be incorporated by suitable means at a later stage. Mi et al.[58] employed enzyme hydrolyzed CS to prepare CS tripolyphosphate and CS polyphosphoric acid gel beads using a polyelectrolyte complexation method for the sustained release of anticancer agent, 6-mercaptopurine. Nandini and Cherng-Ju^[59] developed drug PECs with poly(acrylamido-2-methyl-1-propansulfonate sodiumco-methyl-methacrylate. They studied and reported that the release kinetics were strongly dependent on the drug solubility rather than on the type of amine in the drug. The release of drugs from the tablets of drug-poly(acrylamide-2-methyl-1-propane sulfonate sodium-co-methyl methacrylate complex were well described by the dissociation/erosion mechanism. Petzold et al.[60] prepared different PECs from poly(diallyl-dimethyl-ammoniumchloride) and two different polyanions and characterized their application as flocculants. The results showed that the most important advantages of PEC were the high velocity of sedimentation and a very broad range of the optimum flocculation concentration.

The review summarized emerging popularity and valuable potential offered by the polyelectrolyte

complexes in the field of drug delivery. They represent an attractive class of polymer-based materials finding an irreplaceable role in many areas of the everyday life used for the preparation of biodegradable and biocompatible three-dimensional membranes, microcapsules, nano-sized formulations and various types of controlled release drug delivery systems.

CONCULSIONS

An extensive research is going on in the area of polyelectrolytes and polyelectrolyte complexes. There is a great potential in utilizing these PECs in ecology, biotechnology, medicine and pharmaceutical technology. However these techniques are not effectively applied for the development of drug delivery systems. They may efficiently modify the release; improve the stability and character of the drug substances due to their capacity to entrap the drug at molecular level. Hence the polyelectrolyte complexes have great potential in the design of novel drug delivery systems.

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REFERENCES

- Philipp B, Dautzenberg H, Linow K, Kotz J, Dawydoff W. Polyelectrolyte complexes-recent developments and open problems. Prog Polym Sci 1989;14:91-172.
- Krishnendu R, Hai-Quan M, Shau-Ku H, Kam WL. Oral gene delivery with chitosan–DNA nanoparticles generates immunologic protection in a murine model of peanut allergy. Nature Med 1999;5:387-91.
- Genta I, Perugini P, Modena T, Pavanetto F, Castelli F, Muzzarelli RA, et al. Miconazole-loaded 6-oxychitin–chitosan microcapsules. Carbohydr Polym 2003;52:11-8.
- Macleod GS, Collett JH, Fell JT. The potential use of mixed films of pectin, chitosan and HPMC for bimodal drug release. J Control Release 1999;58:303-10.
- Kubisa P. Terminology of Polymers Containing Ionizable or Ionic Groups and of Polymers Containing Ions, IUPAC Recommendations 2004, (DRAFT 23 December 2004).
- Available from: http://scgc.epfl.ch/load/cours_chim/chwanrey_part.5.pdf. [cited in 2004].
- Joanny JF, Castelnovo M. Polyelectrolyte adsorption and multiplayer formation. In: Decher G. Schlenoff JB, editors. Multilayer Thin Films. Weinheim: Wiley-VCH; 2002. p. 87-97.
- Webster L, Huglin MB, Robb ID. Complex formation between polyelectrolytes in dilute aqueous solution. Polymer 1997;38:1373-80.

- Webster L, Huglin MB. Observations on complex formation between polyelectrolytes in dilute aqueous solution. Eur Polym J 1997;339:1173-7.
- Burgess DJ. Practical analysis of complex coacervate systems. J Colloid Interfacial Sci 1990;140:227-38.
- Overbeek JT, Voorn MJ. Phase separation in polyelectrolyte solutions: Theory of complex coacervation. J Cell Comp Physiol 1957;49:7-26.
- Voorn MJ. Phase separation in polymer solutions. Fortschr Hochpolym-Forsc 1959;1:192-233.
- Xavier C, Jean-Francois J. Adsorption of polyelectrolyte solutions on surface: A Debye-Huckel theory. J Phys II France 1996;6:1669-86.
- Tsuchida E. Formation of polyelectrolyte complexes and their structures. J Macromol Sci Pure Appl Chem 1994;A31:1-15.
- Kokufuta E. Colloid titration behavior of poly (ethyleneimine). Macromolecules 1979;12:350-3.
- Monika S, Layered polyelectrolyte complexes: Physics of formation and molecular properties. J Phys Condens Matter 2003;15:R1781-808.
- Maarten P, Biesheuvel, Martien A, Cohen S. Cylindrical cell model for the electrostatic free energy of polyelectrolyte complexes. Langmuir 2004;20:4764-70.
- Fredheim GE, Christensen BE. Polyelectrolyte complexes: Interactions between lignosulfonate and chitosan. Biomacromol 2003;4:232-9.
- Weinbreck F, de Vries R, Schrooyen P, de Kruif CG. Complex coacervation of whey proteins and gum arabic. Biomacromol 2003;4:293-303.
- 20. Alexander K, Monica OD. Precipitation of oppositely charged polyelectrolytes in salt solutions. J Chem Phys 2004;20:404-12.
- Gamzazade AI, Nasibov SM. Formation and properties of polyelectrolyte complexes of chitosan hydrochloride and sodium dextransulfate. Carbohydr Polym 2002;50:339-43.
- 22. Shiratori SS, Rubner MF. pH-Dependent thickness behavior of sequentially adsorbed layers of weak polyelectrolytes. Macromol 2000;33:4213-9.
- Kumar V, Yang T, Yang Y. Interpolymer complexation. I. Preparation and characterization of a polyvinyl acetate phthalatepolyvinylpyrrolidone (PVAP-PVP) complex. Int J Pharm 1999;188:221-32.
- Argüelles-Monal W, Cabrera G, Peniche C, Rinaudo M. Conductimetric study of the interpolyelectrolyte reaction between chitosan and polygalacturonic acid. Polymer 2000;41:2373-8.
- Herman P, Van L, Rob FM, Cleven, Pavel V. Conductometric analysis of polyelectrolytes in solution. Pure App Chem 1991;63:1251-68.
- Takayama K, Nagai T. Application of interpolymer complexation of polyvinylpyrrolidone/carboxyvinyl polymer to control of drug release. Chem Pharm Bull 1987;35:4921-7.
- Zhang LM. Synergistic blends from aqueous solutions of two cellulose derivatives. Colloid Polym Sci 1999;277:886-90.
- Herbert D, Light scattering studies on polyelectrolyte complexes. Macromolecular Symposia 2001;162:1-22.
- Natalia VP, Nickolay VT. Structure and dynamics of the polyelectrolyte complex formation. Macromol 1997;30:4897-904.
- Anlar Ş, Çapan Y, Güven O, Göğüş A, Dalkara T, Hincal A. Formulation and in-vitro-in-vivo evaluation of buccoadhesive morphine sulfate tablets. Pharm Res 1994;11:231-6.
- Senuma M, Kuwabara S, Kaeriyama K, Hase F, Shimura Y. Polymer complex from copolymers of acrylonitrile and ionic vinyl benzyl compounds. J Appl Polym Sci 1986;31:1687-97.
- Sato H, Maeda M, Nakajima A. Mechanochemistry and permeability of polyelectrolyte complex membranes composed of poly (vinyl alcohol) derivatives. J Appl Polym Sci 1979;23:1759-67.
- Harris EL, Angal S. Protein purification methods: A practical approach. New York: Oxford University Press; 1993.
- Yamamoto H, Horita C, Senoo Y, Nishida A, Ohkawa K. Polyion complex fiber and capsule formed by self-assembly of chitosan and gellan at solution interfaces. Macromol Chem Phys 2000;201:84-92.
- Hirouki Y, Takeshi K. Adsorption of BSA on cross-linked chitosan: The equilibrium isotherm. Chem Eng Japan 1989;41:B11-5.

- Dubin PL, Gao J, Mattison K. Protein purification by selective phase separation with polyelectrolytes. Sep Purif Methods 1994;23:1–16.
- Cordes RM, Sima WB, Glatz CE. Precipitation of nucleic acids with poly (ethyleneimine). Biotechnol Prog 1990;6:283-5.
- Atkinson JG. Precipitation of nucleic acids with polyethyleneimine and the chromatography of nucleic acids on immobilized polyethyleneimine. Biochim Biophys Acta 1973;308:41-52.
- Jendrisak J. In: Burgerss R, editors Protein purification: Micro to macro. New York: Alan R Liss Inc; 1987. p. 75-97.
- Chen J, Jo S, Park K. Polysaccharide hydrogels for protein drug delivery. Carbohydr Polym 1995;28:69-76.
- Dautzenberg H, Kotz J, Linow KJ, Philipp B, Rother G. Static light scattering of polyelectrolyte complex solutions. In: Dubin P, Bock J, Davis R, Schulz DN, Thies C, editors. Macromolecular complexes in chemistry and biology. Berlin: Springer Verlag; 1994. p. 119-33.
- Artur B, David H. Carrageenan-oligochitosan microcapsules: Optimization of the formation process. Colloids Surf B: Biointerfaces 2001;21:285-98.
- Murakami R, Takashima R. Mechanical properties of the capsules of chitosan-soy globulin polyelectrolyte complex. Food Hydrocolloids 2003;17:885-8.
- Kabanov VA. In: Dubin P, Bock J, Davis R, Schulz DN, Thies C, editors. Macromolecular Complexes in Chemistry and Biology. New York: Springer; 1994.
- 45. Leclercq L, Boustta M, Vert M. A physico-chemical approach of polyanion-polycation interactions aimed at better understanding the *in vivo* behaviour of polyelectrolyte-based drug delivery and gene transfection. J Drug Targeting 2003;11:129-38.
- Kekkonen J, Lattu H, Stenius P. Adsorption kinetics of complexes formed by oppositely charged polyelectrolytes. J Colloid Interface Sci 2001;234:384-92.
- Shojaei AH. Buccal mucosa as route for systemic drug delivery: A review. J Pharm Pharmaceut Sci 1998;1:15-30.
- Krone V, Magerstadt M, Walch A, Groner A, Hoffmann D. Pharmacological composition containing polyelectrolyte complexes in microparticulate form and at least on active agent. United States patent 5,700,459, Dec 23, 1997.
- 49. Kawashima Y, Handa T, Kasai A, Takenaka H, Lin SY, Ando Y. Novel method for the preparation of controlled-release theophylline granules coated with a polyelectrolyte complex of sodium polyphosphatechitosan. J Pharm Sci 1985;74:264-8.
- Shiraishi S, Imai T, Otagiri M. Controlled release of Indomethacin by chitosan polyelectrolyte complex: Optimization and *in vivo/in vitro* evaluation. J Control Release 1993;25:217-25.
- Jimenez-Kairuz AF, Llabot JM, Allemandi DA, Manzo RH. Swellable drug-polyelectrolyte matrices (SDPM): Characterization and delivery properties. Int J Pharm 2005;288:87-99.
- Liao I-C, Wan ACA, Yim EK, Leong KW. Controlled release from fibers of polyelectrolyte complexes. J Control Release 2005;104:347-58.
- 53. Tapia C, Escobar Z, Costa E, Sapag-Hagar J, Valenzuela F, Basualto C, et al. Comparative studies on polyelectrolyte complexes and mixtures of chitosan-alginate and chitosan-carrageenan as prolonged diltiazem clorhydrate release systems. Eur J Pharm Biopharm 2004;57:65-75.
- 54. Paloma M, de la T, Yewande E, Guillermo T, Susana T. Release of amoxicillin from polyionic complexes of chitosan and poly(acrylic acid): Study of polymer/polymer and polymer/drug interactions within the network structure. Biomaterials 2003;24:1499-506.
- 55. Win PP, Shin-ya Y, Hong KJ, Kajiuchi T. Formulation and characterization of pH sensitive drug carrier based on phosphorylated chitosan (PCS). Carbohydr Polym 2003;53:305-10.
- Albano AA, Phuapradit W, Sandhu HK, Shah NH. Stable complexes of poorly soluble compounds in ionic polymers. United States Patent 6,350,786, 2002.
- Rolfes H, Van Der Merve TL, Truter PA. Method of making controlled release particles of complexed polymers. United States Patent 6,221,399, 2001.
- 58. Mi FL, Shyu SS, Kuan CY, Lee ST, Lu KT, Jang SF. Chitosan-

Polyelectrolyte complexation for the preparation of gel beads and controlled release of anticancer drug: I: Effect of phosphorous polyelectrolyte complex and enzymatic hydrolysis of polymer. J Appl Polym Sci 1999;74:1868-79.

- Nandini K, Cherng-ju K. Drug release from drug-polyanion complex tablets: Poly(acrylamido-2-methyl-1-propanesulfonate sodium-co-methyl methacrylate). J Control Release 1999;57:141-50.
- Petzold G, Nebel A, Buchhammer H-M, Lunkwitz K. Preparation and characterization of different polyelectrolyte complexes and their application as flocculants. Colloid Polym Sci 1998;276:125-30.
- Dainiak MB, Izumrudov VA, Muronetz VI, Galaev IY, Mattiasson B. Affinity precipitation of monoclonal antibodies by nonstoichiometric polyelectrolyte complexes. Bioseparation 1998;7:231-40.
- 62. Izumrudov VA, Galaev IY, Mattiasson B. Polycomplexes--potential for bioseparation. Bioseparation 1998;7:207-20.
- Kim B, Peppas NA. Analysis of molecular interactions in poly (methacrylic acid-g-ethylene glycol) hydrogels. Polym 2003;44:3701-7.
- Pirogov AV, Shpak AV, Shpigun OA. Application of polyelectrolyte complexes as novel pseudo-stationary phases in MEKC. Anal Bioanal Chem 2003;375:1199-203.
- Barreiro-Iglesias R, Alvarez-Lorenzo C, Concheiro A. Controlled release of estradiol solubilized in carbopol/surfactant aggregates. J Control Release 2003;93:319-30.
- 66. Lee J, Cho EC, Cho K. Incorporation and release behavior of

hydrophobic drug in functionalized poly (lactide)-block-poly (ethylene oxide) micelles. J Control Release 2004;94:323-35.

- Swanson VM, Dubin PL, Almgren M, Li Y. Cryo-TEM of polyelectrolyte-micelle complexes. J Colloid Interface Sci 1997;186:414-9.
- Cherng-Ju K, Yvonne NN. Drug release from an erodible drugpolyelectrolyte complex. Eur Polym J 1995;31:937-40.
- Anders L, Charlotte W, Staffan W. Flocculation of cationic polymers and nanosized particles. Colloids Surfaces A: Physicochem Eng Aspects 1999;159:65–76.
- Janne L, Lindström T. Topo chemical modification of cellulosic fibers with bipolar activators: An overview of some technical applications. Sci Technol 2001;1:40-5.
- Mendelsohn JD, Barrett CJ, Chan VV, Pal AJ, Mayes AM, Rubner MF. Fabrication of microporous thin films from polyelectrolyte multilayers. Langmuir 2000;16:5017-23.
- 72. Yan X, Khor E, Lim LY. PEC films prepared from Chitosan-Alginate coacervates. Chem Pharm Bull 2000;48:941-6.

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