# Protective Effect of *Ipomoea staphylina* Against Cadmium-Induced Cardiotoxicity in Wistar Rats

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#### Yanchun et al.: Cardioprotective Effect of Ipomoea staphylina

The present study was aimed to evaluate the protective effect of ethanolic extract of Ipomoea staphylina leaves against cadmium-induced cardiotoxicity in rats. Rats were segregated into four groups which included normal control, cadmium treated, Cd+ Ipomoea staphylina (200 mg/kg) treated, and Cd+ Ipomoea staphylina (400 mg/kg) treated. The cadmium treatment resulted in a significant increase in the activities of lactate dehydrogenase, creatine phosphokinase, serum glutamic oxaloacetic transaminase and serum glutamic pyruvic transaminase. Also, a significant increase in the levels of cholesterol, triglycerides, and low-density lipoprotein was noticed after cadmium treatment. On the other hand, high-density lipoprotein was significantly decreased following cadmium treatment. A significant decline in the activity of antioxidant enzymes was observed in cadmium intoxicated rats. Moreover, cadmium intoxicated rats resulted in significant increase in the levels of proinflammatory cytokines such as tumor necrosis factor alpha, interleukin 1 beta, and nitric oxide in the cardiac tissue. Supplementation of *Ipomoea staphylina* leaves extract to cadmium treated rats reversed most of the indices. The extract (400 mg/kg) revealed better protective effects by decreasing the activities of lactate dehydrogenase, creatine phosphokinase, serum glutamic oxaloacetic transaminase and serum glutamic pyruvic transaminase and improving the lipid profile. Besides, the extract also significantly improved the antioxidant enzymes activity and decreased the level of tumor necrosis factor alpha, interleukin 1 beta, and nitric oxide. Further, the histopathological study of heart supported the protective effects of *Ipomoea staphylina* leaves extract. The study, therefore, concludes that *Ipomoea staphylina* leaves extract holds a strong protective effect against cadmium-induced cardiotoxicity in rats.

Key words: Diclofenac sodium, ethylhexyl palmitate, microemulsion, release kinetics, topical delivery

Cadmium (Cd), has been confirmed as a potent cardiotoxic environmental heavy metal<sup>[1,2]</sup>. It has been widely reported to induce oxidative stress and membrane disturbances in cardiac myocytes<sup>[3]</sup>. The multiple ways of Cd exposure include smoking, diet, polluted water with Cd and occupational exposures in certain industries. The prime reason behind the reported cardiotoxic effect is its capability to get absorbed in population even at low-level chronic exposures. Secondly, Cd has a long half-life, ranging from 10 to 40 y, leading to long term in vivo accumulation that in turn causes multi-organ toxicities including the heart. The heart is one of the most targeted organs by Cd exposure and results in oxidative damage. The human's myocardial autopsy reports also confirmed the presence of high levels of Cd in the myocardium<sup>[4]</sup>. Moreover, multiple reports suggested a clear association of Cd

toxicity with cardiovascular diseases (CVD) like hypertension, atherosclerosis, stroke, and cardiac arrest<sup>[5]</sup>. However, there is a paucity of information about exact mechanisms responsible for Cd mediated cardiovascular toxicities<sup>[6]</sup>.

Many researchers worldwide are focusing on the compounds with good antioxidant potential as Cd exposure ultimately leads to oxidative stress. So, this oxidative stress further results in the production of reactive oxygen species (ROS), the already established

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Accepted 23 January 2021 Revised 25 Ocotber 2020 Received 06 January 2020 Indian J Pharm Sci 2021;83(1):93-100 prime factors for cardiotoxicity. In the present study, the protective effect of the ethanolic extract of *Ipomoea staphylina* has been explored against Cd-induced cardiotoxicity. *Ipomoea staphylina* (Family: Convolvulaceae) is a perennial woody plant considered as an important medical plant in an Asian traditional medicine system.

It is commonly used for the treatments of liver diseases, purgation, stomach disorders, pain, inflammation, and rheumatism. Experimental studies on this plant revealed that it contains sitosteryl-3-O-β-D-glucoside and chiro deoxy inositol<sup>[7]</sup>. The leaves extract of this plant possesses analgesic<sup>[8]</sup>, anti-inflammatory<sup>[9]</sup>, antiulcer<sup>[10]</sup>, hepatoprotective<sup>[11,12]</sup>, antidiabetic<sup>[13]</sup>, nephroprotective<sup>[11]</sup>, and *in vitro* antioxidant activities<sup>[12]</sup>. However, there is a paucity of information about its use as cardioprotective. Therefore, the present study has been planned to evaluate its protective potential against Cd-induced cardiotoxicity in rats.

# MATERIALS AND METHODS

Cd was procured from Sigma Chemicals (United States of America (USA)). Lactate dehydrogenase (LDH), creatine phosphokinase (CPK), serum glutamic oxaloacetic transaminase (SGOT), serum glutamic pyruvic transaminase (SGPT), cholesterol, and triglycerides kits were obtained from Span Diagnostics, India. Tumor necrosis factor-alpha (TNF- $\alpha$ ), interlukin-1 beta (IL-1 $\beta$ ), and nitric oxide (NO) kits were procured from Sigma Chemicals (USA). All other chemicals were commercial products of analytical reagent grade.

# Collection and authentication of the plant:

Leaves of *Ipomoea staphylina* were collected from forest area of Karnataka near to Bangalore. The *Ipomoea staphylina* plant taxonomically identified and authenticated by Dr. K. Karthigeyan at Central National Herbarium, Botanic Garden, Howrah, where the voucher specimen is conserved under the reference number SMF-01.

# Preparation of ethanolic extract of leaves:

The leaves of *Ipomoea staphylina* were cleaned and dried under shade at room temperature for several days and powdered. The powder was defatted with petroleum ether (60-80 GR) for 72 h and then the dried powder was extracted with ethyl alcohol to get a yield of 10.20 % w/w dried extract was stored at 4° in a refrigerator<sup>[13]</sup>.

# **Experimental animals:**

Male Wistar rats in the weight range of 180-200 g were procured from the central animal house, of the institute. Animals were housed in polypropylene cages on a 12 h d/night cycle at a temperature of 22±1°. They were housed in polypropylene cages under hygienic conditions in the departmental animal house. All the experimental procedures were performed according to the ethics committee of The First Affiliated Hospital of Jilin University (Ethics No. ZN-IRB-00I-08).

# Acute oral toxicity:

Male Wistar rats were kept overnight fasting before drug administration. This was followed by administration of a single oral dose (2000 mg/kg) of ethanolic extract *Ipomoea staphylina* leaves. The diet intake of animals was withheld for 3-4 h and they were kept under observation. They were observed twice after the first 30 min after dosing, periodically during the first 24 h (with special attention during the first 4 h) and daily thereafter for 14 d<sup>[14]</sup>.

# **Experimental design:**

After 1 w acclimatization, male Wistar rats were randomly divided into four groups (n=6). The Cd solution was freshly prepared in normal saline before administration. Similarly, the suspension of the extract was freshly prepared in 0.3 % w/v carboxymethyl cellulose (CMC) before administration.

Each group received the following treatment;

Group I: Served as control and received normal saline (1 ml/kg; orally (p.o.)). Group II: Received only Cd (5 mg/kg; p.o.). Group III: Received co-treatment of ethanolic extract of *Ipomoea staphylina* (200 mg/kg; p.o.) and Cd (5 mg/kg; p.o.). Group IV: Received co-treatment of ethanolic extract of *Ipomoea staphylina* (400 mg/kg; p.o.) and Cd (5 mg/kg; p.o.).

The rats were exposed to the respective treatment as mentioned in the groups for 28 d. On the 28<sup>th</sup> d, rats fasted for 24 h and blood was drawn by cardiac puncture under ether anesthesia for various biochemical estimations. The animals were sacrificed by cervical decapitation and the heart was dissected out, immediately rinsed in ice-cold saline, and stored for further biochemical and histopathological analysis<sup>[15,16]</sup>.

#### **Biochemical analysis:**

LDH, CPK, SGOT, SGPT, cholesterol, triglycerides, and lipoproteins were estimated using commercial kits

(Span Diagnostics, India) as per the manufacturer's manual.

# Endogenous antioxidants and pro-inflammatory cytokines content in the heart:

Tris phosphate buffer (50 mM, pH 7.4) was used to prepare heart tissue homogenates (10 % w/v). The homogenate was centrifuged at 3000 rpm for 10 min at 4°. The supernatant was used to estimate lipid peroxidation (LPO), superoxide dismutase (SOD), reduced glutathione (GSH), and catalase (CAT)<sup>[17-20]</sup>. The levels of pro-inflammatory cytokines (TNF- $\alpha$  and IL1- $\beta$ ) and NO were estimated by commercially available enzyme-linked immunosorbent assay (ELISA) kits, according to the instructions of the manufacturer.

# Histopathological studies:

For the histopathological study fresh heart tissue pieces of rats were fixed in formalin. Following an overnight fixation in formalin, the specimens were dehydrated in ascending grades of alcohol, cleared in benzene and embedded in paraffin wax. Blocks were made and 4-5  $\mu$ m thick sections were double-stained with hematoxylin and eosin (H&E) and which were observed under the light microscope.

# Statistical analysis:

The statistical significance of the data has been determined using one-way analysis of variance (ANOVA) followed a multiple post hoc least significant difference test. The results are represented as mean±standard deviation (SD).

### RESULTS AND DISCUSSION

The results obtained from various experiments conducted in this study are depicted in Table 1, Table 2, Table 3, Table 4 and fig. 1. The data from various treatment groups have been compared with the

TABLE 1: EFFECTS OF Ipomoea staphylina ON SERUM LDH, CPK, SGOT, SGPT IN Cd TREATED RATS

Groups	Treatment	LDH (U/l)	CPK (U/I)	SGOT (IU/I)	SGPT (IU/I)
I	Normal control	154.08±1.36	40.26±0.87	34.15±0.94	58.33±1.12
II	Cd control	422.02±1.13 <sup>c</sup>	161.63±1.13°	181.83±2.51 <sup>c</sup>	169.73±1.04 <sup>c</sup>
III	Cd+ Ipomoea staphylina (200 mg/kg)	261.99±1.43b,y	86.19±1.32 <sup>c,y</sup>	103.45±1.54 <sup>c,x</sup>	91.98±1.18 <sup>b,y</sup>
IV	Cd+ <i>Ipomoea staphylina</i> (400 mg/kg)	217.55±2.84 <sup>b,y</sup>	75.24±1.36 <sup>b,x</sup>	82.69±1.78c,z	70.99±1.21a,z

n=6, Data are expressed in Mean $\pm$ SD,  $^ap<0.05$ ,  $^bp<0.01$  and  $^cp<0.001$  by least significance difference test when values are compared with group I;  $^xp<0.05$ ,  $^yp<0.01$ , and  $^zp<0.001$  by least significance difference test when values of groups III and IV are compared with group II

TABLE 2: EFFECTS OF *Ipomoea staphylina* ON CHOLESTEROL, TRIGLYCERIDES AND LIPOPROTEINS LEVELS IN Cd TREATED RATS

Groups	Treatment	Cholesterol (mg/dl)	Triglycerides (mg/dl)	LDL (mg/dl)	HDL (mg/dl)
I	Normal control	54.08±1.36	48.26±0.17	9.15±0.34	37.33±0.12
II	Cd control	97.02±1.13 <sup>c</sup>	77.30±0.13 <sup>b</sup>	15.05±0.51a	24.73±0.04 <sup>b</sup>
III	Cd+ Ipomoea staphylina (200 mg/kg)	$68.99 \pm 0.43^{a,x}$	54.78± 0.32×	11.04± 0.54 <sup>a,x</sup>	$30.98 \pm 0.18^{a,x}$
IV	Cd+ Ipomoea staphylina (400 mg/kg)	57.55±0.84 <sup>y</sup>	46.41±0.36 <sup>y</sup>	$8.69\pm0.78^{y}$	33.3±0.21 <sup>y</sup>

n=6, Data are expressed in Mean±SD,  $^{\rm a}$ p<0.05,  $^{\rm b}$ p<0.01, and  $^{\rm c}$ p<0.001 by least significance difference test when values are compared with normal control group;  $^{\rm x}$ p<0.05 and  $^{\rm y}$ p<0.01 by least significance difference test when values of groups III and IV are compared with group II

TABLE 3: EFFECTS OF *Ipomoea staphylina* ON ANTIOXIDANT DEFENSE SYSTEM ENZYMES AND LPO IN Cd TREATED RATS

Groups	Treatment	LPO (n mol of MDA formed/min/mg protein)	SOD (IU)	GSH (µg/mg of protein)	CAT (µ mol H <sub>2</sub> O <sub>2</sub> /mg of protein )
I	Normal control	1.08±1.36	17.96±0.17	15.75±0.34	2.33±0.12
II	Cd control	2.91±0.31 <sup>c</sup>	8.30±0.13 <sup>c</sup>	8.08±0.51 <sup>c</sup>	1.43±0.40°
III	Cd+ <i>Ipomoea staphylina</i> (200 mg/kg)	1.51±1.43 <sup>b,y</sup>	12.78±0.32 <sup>a,y</sup>	12.04± 0.54 <sup>a,y</sup>	1.70±0.18 <sup>b,x</sup>
IV	Cd+ <i>Ipomoea staphylina</i> (400 mg/kg)	1.05±0.84 <sup>a,z</sup>	14.41±0.3a,y	14.69±0.78 <sup>y</sup>	2.09±0.21a,y

n=6, Data are expressed in Mean±SD,  $^{a}p<0.05$ ,  $^{b}p<0.01$ , and  $^{c}p<0.001$  by least significance difference test when values are compared with group I;  $^{x}p<0.05$ ,  $^{y}p<0.01$ , and  $^{z}p<0.001$  by least significance difference test when values of groups III and IV are compared with group

TABLE 4: EFFECTS OF Ipomoea staphylina ON HEART TNF-A, IL-1β, AND NO IN Cd TREATED RATS

Crauna	Treatment	TNF-α	IL-1B	NO
Groups		(ρg/mg tissue)	(ρg/mg tissue)	(ρg/mg tissue)
I	Normal control	26.08±1.36	23.26±0.17	15.15±0.34
II	Cd control	117.02±1.13 <sup>c</sup>	53.30±0.11 <sup>c</sup>	46.05±0.51°
Ш	Cd+ Ipomoea staphylina (200 mg/kg)	67.99±1.43 <sup>c,y</sup>	40.78± 0.32b,x	34.04± 0.54 <sup>c,x</sup>
IV	Cd+ Ipomoea staphylina (400 mg/kg)	47.55±0.84 <sup>b,z</sup>	$32.41 \pm 0.36^{a,y}$	$22.69 \pm 0.78^{a,y}$

n=6, Data are expressed in Mean±SD,  ${}^{a}p<0.05$ ,  ${}^{b}p<0.01$ , and  ${}^{c}p<0.001$  by least significance difference test when values are compared with group I;  ${}^{x}p<0.05$ ,  ${}^{y}p<0.01$ , and  ${}^{c}p<0.001$  by least significance difference test when values of groups III and IV are compared with group II

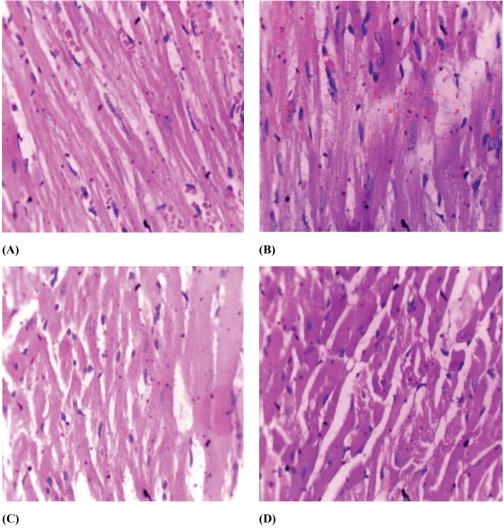


Fig. 1: Histopathological analysis of heart tissues of experimental animals (A) Normal control: showing normal myocardial fibers with normal architecture (H&E, 400x); (B) Cd control: showing degeneration and discontinuity of the hypertrophic myocardial fibers with infiltration of inflammatory cells and damaged nucleus (H&E, 400x); (C) Cd+ *Ipomoea staphylina* (200 mg/kg): showing less degeneration and discontinuity of the myocardial fibers with mild infiltration of inflammatory cells (H&E, 400x); (D) Cd+ *Ipomoea staphylina* (400 mg/kg): showing almost normal heart tissue architecture (H&E, 400x). \*=damage discontinued muscle fibres; +=inflammatory cells; #=damaged nucleus

normal control animals. Further, results obtained from group III and group IV animals were compared with that of Cd treated group II.

In the acute oral toxicity study, it was found that animals showed no changes in normal behaviour, signs of toxicity and mortality to a maximum dose of 2000 mg/kg. Thus, 200 and 400 mg/kg doses were considered in this study.

A statistically significant increase in the enzyme activity of LDH as well as CPK was observed in Cd treated rats (Table 1). Also, both SGOT and SGTP also showed a significant increase in their activities after

treatment with Cd. On the other hand, the ethanolic extract of *Ipomoea staphylina* leaves (200 and 400 mg/kg) resulted in a significant decrease in activities of all the above enzymes. Further, the modulatory potential was more prominent at a higher dose of the extract.

Cd treatment resulted in a significant increase in the levels of cholesterol, triglycerides, and low-density lipoprotein (LDL) (Table 2). On the other hand, high-density lipoprotein (HDL) showed a significant decline after Cd treatment. Co-administration of the extract of *Ipomoea staphylina* (200 and 400 mg/kg) significantly reduced the level of cholesterol, triglycerides, and LDL. Moreover, the extract significantly improved the level of HDL. The effect was more efficient at higher does (400 mg/kg).

A significant increase in the levels of LPO was observed in the cardiac tissues of Cd treated rats (Table 3). Besides, a significant decrease was observed in the levels of GSH as well as in the activities of SOD and CAT (Table 3) in the Cd treated group. Treatment with the extract of *Ipomoea staphylina* resulted in a significant decrease in LPO levels in Cd treated rats. Moreover, the extract also resulted in a significant improvement of the GSH levels and the activities of SOD and CAT.

Cd treatment resulted in a significant increase in the level of TNF- $\alpha$ , IL-1 $\beta$ , and NO (Table 4). Treatment with the extract of *Ipomoea staphylina* resulted in a significant decrease in the enzyme activities of all three indices. Further, the protective potential was more evident at a higher dose of ethanolic extract of *Ipomoea staphylina*.

Fig. 1 shows that sections of the normal heart (A) showing normal myocardial fibers with normal architecture. Rats intoxicated with Cd (B) showing degeneration and discontinuity of the hypertrophic myocardial fibers with infiltration of inflammatory cells. Section of the heart of rats treated with extract of *Ipomoea staphylina* (200 mg/kg) (C) showing less degeneration and discontinuity of the myocardial fibers with mild infiltration of inflammatory cells. Sections of the heart tissue of rats treated with the extract of *Ipomoea staphylina* (400 mg/kg) (D) showing almost normal heart tissue architecture.

The present study evaluated the protective effect of ethanolic extract of *Ipomoea staphylina* leaves against Cd-induced cardiotoxicity in Wistar rats. The study indicated that the administration of ethanolic extract of *Ipomoea staphylina* to the Cd treated animals

appreciably modulated most of the biochemical indices along with histoarchitecture changes, which were altered due to toxic effects of Cd on cardiac tissues. All these improvements collectively contributed towards the protective effect of ethanolic extract of *Ipomoea staphylina* against Cd-induced cardiotoxicity.

LDH is a marker of cardiac tissue injury including heart failure<sup>[21]</sup> and it is recorded to be significantly elevated in the present study in Cd treated rats. Therefore, the above rise in LDH activity upon Cd treatment confirmed cardiac toxicity. Moreover, earlier reports on LDH are in corroboration with our results<sup>[22]</sup>. CPK is an enzyme responsible for the phosphorylation of creatine. High levels of CPK have been related to muscle injuries including heart muscles. In the present study, CPK showed a significant rise in Cd treated rats confirming injuries to heart muscles due to Cd toxicity. Also, a significant rise in SGOT and SGPT established Cd toxicity in the heart as these enzymes have been reported earlier in cardiac toxic states<sup>[23]</sup>. On the other hand, treatment with the extract of Ipomoea staphylina significantly reversed the effects of Cd and confirmed its protective effect. Moreover, the protective effects were more prominent at higher dose.

Alterations in the cholesterol levels, triglycerides and lipoproteins are considered as direct evidence of ill effects on heart health<sup>[24]</sup>. The present study showed significant changes, noticed in cholesterol, triglycerides, and lipoproteins. High LDL levels can cause the build-up of plaque leading to narrowing of the arteries that ultimately block the blood flow. Treatment with the extract of *Ipomoea staphylina* reversed the altered levels of cholesterol along with triglycerides. Naturally occurring phytosterols are well known for lipid-lowering activity<sup>[25]</sup>. *Ipomoea staphylina* has been reported to contain phytosterols like sitosteryl-3-O-β-D-glucoside<sup>[7]</sup>. Thus, the above lipid lowering effects could be a result of the presence of phytosterols in *Ipomoea staphylina*.

We observed a significant increase in LPO in the heart following Cd treatment. This is based on the appraisal of malondialdehyde (MDA) levels formed during the oxidative degradation of membranous polyunsaturated fatty acids. Interestingly, simultaneous treatment with ethanolic extract of *Ipomoea staphylina* to Cd treated animals showed moderation in the MDA levels that could be owed to their antioxidative nature and ability to scavenge ROS<sup>[26]</sup>. The observed increase in LPO is associated with a decrease in GSH levels as reduced

glutathione is consumed by the glutathione related enzymes to detoxify peroxides formed by LPO<sup>[27,28]</sup>. Also, the sequestration of antioxidants like GSH is reported as an essential demand during toxic states. Besides, CAT is a free radical scavenging enzyme involves in detoxification of hydrogen peroxide<sup>[29]</sup>. It has been reported that an increase in hydrogen peroxide level might induce peroxidation of polyunsaturated fatty acids and increase the formation of MDA. The antioxidant enzyme, SOD is considered to be the primary enzyme as it is involved in the direct elimination of ROS. This enzyme works in coordination with other antioxidant enzymes to eliminate ROS. Also, it is a chain breaking antioxidant and plays an important role in protection against deleterious effects of LPO<sup>[30]</sup>. In the present study, SOD activity was found to be significantly decreased following Cd treatment which in turn promoted cardiac toxic effects by stimulating oxidative stress[31]. Treatment with *Ipomoea staphylina* extract decreased the levels of MDA and improved the activity of CAT and SOD. It is quite probable that Ipomoea staphylina supplementation made zinc available for the optimum functioning of the SOD against oxidative stress as phytochemicals have been reported to maintain normal zinc levels during oxidative stress<sup>[32]</sup>.

TNF-α is an inflammatory cytokine that usually elevates in the cases of cardiotoxicity, has been demonstrated in experimental models<sup>[33]</sup>. In this study, a significant increase was noticed in TNF-α confirming the deleterious toxic effects of Cd. The above rise in the TNF- $\alpha$  level may also involve in the inflammation of cardiac tissue. Moreover, IL-1β levels also showed a significant rise in the Cd treated rats. This IL-1β is also a direct indicator of inflammation. Heart failure patients have been demonstrated a marked elevation in a variety of inflammatory cytokines including IL-1β according to the degree of severity of the disease independent of etiology<sup>[34]</sup>. Hence, it is one of the crucial inflammatory markers. Further, NO levels also showed an elevation in the Cd treated rats. The observed rise in IL-1β could be responsible for this increase in NO levels as it has been reported earlier to cause an increase in the expression of Nitric Oxide Synthase (NOS)[35]. Besides, NO is a crucial regulator of heart contractility and rate. So, any alternation in NO levels significantly affects cardiac health. So, all the above changes proved cardiac toxicity induced by Cd. Treatment with the extract of Ipomoea staphylina modulated above changes proving its anti-inflammatory ability.

Histological changes of Cd intoxicated heart tissue supported biochemical findings. Control animal slides showed normal architechture of cardiac muscle fibers. On the other hand, Cd treatment revealed significant signs of toxicity in the form of enlarged dark nuclei in the degenerated, discontinued and hypertrophic myocardial fibers with infiltration of inflammatory cells. Also, dilated ventricles and atria were visible confirming edema and fibrosis in the cardiac tissue<sup>[36]</sup>. Treatment with *Ipomoea staphylina* extract showed a marked improvement in the histoarchitecture of cardiac tissues. The signs of dilated ventricles and atria disappeared.

From this study, we came to know that Cd intoxication leads to an increase in oxidative stress and decreased the antioxidant defense specially GSH. It is known that GSH protects from oxidative assaults and it prevents the LPO and oxidized glutathione formation in the heart. It has been demonstrated that glucose-6phosphate dehydrogenase (G6PD) depletes cytosolic GSH levels and subsequently results in cardiomyocyte contractile dysfunction through dysregulation of calcium homeostasis which may be a reason for Cdinduced cardiac dysfunction<sup>[37]</sup>. In our study, we found that the extract notably increased the level of GSH which could probably via inhibition of G6PD by the extract. Moreover, a literature survey reveals that oxidative stress and inflammatory cytokines are the major culprits to bring out the artifacts in cardiac tissue due to Cd exposure and thus the effect of the extract on oxidative stress and inflammatory markers in Cd intoxicated rats was studied. Inflammation resulting from oxidative stress is the major cause of cardiac disease. An increase in the oxidative stress causes activation of Nuclear factor kappa B (NF-kappa-B), one of the key signalling molecules, which is a transcription factor that up-regulates the production of downstream inflammatory mediators, including TNF-α, IL-1β, and NO<sup>[34,35]</sup>. However, we have evaluated the effect of the extract on endogenous antioxidants especially SOD, CAT, and GSH and inflammatory markers (Heart TNF- $\alpha$ , IL-1 $\beta$ , and NO). We found a noteworthy increase in antioxidative defense and a decrease in pro-inflammatory cytokines in the heart of Cd intoxicated animals due to the supplementation of the extract. Besides, the ability of *Ipomoea staphylina* treatment to restore the histological changes induced by Cd indicated its protective efficacy against Cd. The present study, therefore, concludes that *Ipomoea* staphylina extract has got great protective potential against Cd-induced cardiac toxicity.

Our results demonstrated that ethanolic extract of *Ipomoea staphylina* showed cardioprotective activity against Cd-induced cardiotoxicity in rats. The observed beneficial findings of this study may be credited to the antioxidant effect and suppression of TNF- $\alpha$ , IL1- $\beta$  and NO in the heart tissue.

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None

#### **Conflict of Interests:**

The authors declared no conflict of interest.

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