Review of Analytical and Sample Processing Technologies for Detecting Antibiotic Residues in Environmental Samples

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The release of antimicrobials from the hospital, pharma industries, agriculture and animal husbandry waste into various environmental sources has considerably increased. This triggers the development of antimicrobial resistance and antimicrobial resistance genes in the ecosystem. Hence, high throughput simultaneous analytical methods are necessary to analyze multiple classes of antibiotics in aquaculture products, soil, meat, honey, milk, water resources and wastewater. This concise review describes various analytical methodologies and sample processing techniques frequently used to determine multiple classes of antibiotics in the diverse field of interest. This article also details the frequently detected antibiotics, various liquid-liquid and solid-phase extraction methods, their applications and the role of pKa values in the extraction efficiency of antibiotics.

Key words: Antibiotics, liquid chromatography with tandem mass spectrometry, extraction techniques, antimicrobial resistance, antimicrobial resistance genes, pKa values, solid-phase extraction cartridges

Worldwide, water sources are contaminated by pharmaceutically active compounds, especially antibiotics that enter the water sources via hospital waste, industrial waste, animal husbandry, and animal livestock. Based on the recent joint surveillance report on Antimicrobial Resistance (AMR) by World Health Organization (WHO) and European Center for Disease Prevention and Control (ECDC), more than 670 000 drugresistant bacterial infections occurred in the Europe/European economic area alone and approximately 33 000 people died as a direct consequence of these infections, and the health burden of AMR is comparable to that of influenza, tuberculosis, and Human immunodeficiency virus (HIV) combined. Several countries have a rising concern about increasing resistance towards thirdgeneration cephalosporins and carbapenems in Klebsiella pneumoniae and carbapenem-resistant Acinetobacter. The recommendation from WHO/ ECDC includes disseminating resistant clones in healthcare and other potential areas and creating awareness of the severe limitations in treatment options for patients with infections caused by these pathogens^[1].

Generally, pharmaceutical compounds, including antibiotics, Non-Steroidal Anti-Inflammatory Drugs (NSAIDs), steroids and analgesics are found at ng/l to μ g/l in environmental resources such as sewage water, wastewater, surface water, groundwater and soil. This may be found in either parent form or a transformed state, i.e., metabolized and called 'pseudo persistent' as it contributes continuously to the environment system^[2]. This is a concerning factor as the antibiotic load in the environment contributes to the proliferation of bacteria, which shows a resistant nature.

In most scenarios, wastewater treatment plants do not efficiently treat pharmaceutical contaminants, mainly antibiotics. Also, there are no strict regulatory guidelines to regulate wastewater treatments, especially in third-world countries. As per the Central Pollution Control Board, India, from the total of 53 998 million liters per day

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generated wastewater, only 19 827 million liters per day are treated in major metropolitan cities. It also raised concern that only 13.5% of sewage water is effectively treated out of 18.6% of the total treatment capacity^[3]. This shows the capability of water treatment plants in metropolitan cities. Apart from the above, the guidelines also recommend specific parameters like pH, biochemical oxygen demand, chemical oxygen demand, suspended solids, ammonical nitrogen, total nitrogen and fecal coliform in treated water. However, no parameters are found for the presence of active pharmaceutical ingredients.

Based on the report on pharmaceuticals in drinking water by WHO, the human health risk has been assessed by chemical risk assessment methods such as acceptable daily intake or tolerable daily intake based on a variety of calculations which includes Point of Departure (PoD). According to the 'United States Environmental Protection Agency' policy, compounds with Margin of Exposure values >100 are generally considered a low level of concern. The PoD is the concentration at which no adverse effects are detected, the No-Observed-Adverse-Effect Level (NOAEL), or, less optimally, the lowest concentration at which adverse effects are seen Lowest-Observed-Adverse-Effect Level (LOAEL), in combination with an additional uncertainty factor^[4]. The resistance in microorganisms is directly proportional to the consumption of potential antibiotics. This directly or indirectly led to an increase in the dosage or combination therapy or treatment with a new set of antibiotics. The existence of these potential components in water sources may be lethal, whether at low or high concentrations^[5].

Antibiotics in the environment are sometimes extremely low. However, it is sufficient to create sublethal antibiotic pressure in the environment. Hence, a sensitive analytical method is necessary to detect them in the respective samples. Sample preparation is another critical factor in achieving sensitive methods. Hence, simultaneous analytical methods and sample processing techniques that can determine multi-class antibiotics with low sensitivity are critical analytical tools. This review will focus on estimating antibiotics by widely used analytical and processing technology and developing analytical methods to detect multiclass antibiotics.

CLASSIFICATION OF ANTIBIOTICS

In general, antimicrobials are active against specific microorganisms in humans and animals by suppressing their proliferation or killing their ability to survive. According to the European Medicines Agency, animal antimicrobials were categorized into four categories. Avoid (category-A), Restrict (category-B), Caution (category-C), and Prudence (category-D) which include all major classifications^[6]. Similarly, the World Organization for Animal Health has listed the specific classifications of antibiotics used in the veterinary field. The common antimicrobials used in veterinary are aminoglycosides, bicyclomycin, macrolides, cephalosporins, lincosamides, types of penicillin, phenicol, quinolones and tetracyclines^[7]. Types of antibiotics are the broad and narrow spectrum that includes aminoglycoside, beta-lactam, tetracycline, various cephalosporins, macrolides, generations of sulphonamides, quinolones and specific large molecules are also acting as antibiotics such as peptide and oligonucleotides. An oligonucleotide is an efficient therapy against bacterial resistance to antibiotic treatment. Lipid Oligonucleotides (LONs) were proved to be efficient in delivering the oligonucleotide sequences in the prokaryotic cells and decreasing the minimum inhibitory concentration of resistant bacteria to a thirdgeneration cephalosporin, the ceftriaxone^[8]. Apart from a broad and narrow spectrum, antibiotics are also classified as bactericidal and bacteriostatic based on their activity against microorganisms. describes the classification Table 1 of antibiotics^[9-11] in the simplest form. Also, the classification includes anti-herpes, antiviral anti-retrovirus, anti-influenza, etc., whereas the antifungal group consists of azole derivatives, polyene, echinocandins and allylamine.

Based on their nature, antimicrobials act on either bacterial cell wall, nucleic acid synthesis, protein synthesis, bacterial folate synthesis, and cell membrane^[12]. These key factors will decide whether the antibiotic is bactericidal or bacteriostatic against the specific microorganisms. Fig. 1 shows the functional groups of major antibiotics which dictate the mechanism of action in bacteria.

In 2019, the AWaRe^[13] Classification Database was developed by WHO based on the recommendation of an expert committee on the 'Selection and Use

of Essential Medicines'.

AWaRe stands for Access, Watch and Reserve to emphasize the importance of their optimal uses and potential for antimicrobial resistance. It includes details of 180 antibiotics and is intended to be an interactive tool for countries to support antibiotic monitoring and optimal use better. The database also says that the WHO does not recommend antibiotics due to a lack of evidence on their reliability. The 'Access' antibiotics group includes antibiotics that have activity against a wide range of commonly encountered susceptible pathogens while showing lower resistance potential than antibiotics in the other groups. The 'Watch' antibiotics group includes antibiotics with higher resistance potential and most of the highest priority agents among the critically important antimicrobials for human medicine and antibiotics at relatively high risk of selection of bacterial resistance. In contrast, the 'Reserve' group of antibiotics includes antibiotics and antibiotic classes that should be reserved to treat confirmed or suspected infections due to multi-drug-resistant organisms.

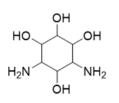
Classification	Spectrum	Nature	Synthetic/Natural
Beta-lactam*	Broad/narrow	Bactericidal	Natural/synthetic
Macrolides	Broad/narrow	Bacteriostatic	Natural/semi-synthetic
Aminoglycoside	Broad	Bactericidal	Natural/semi-synthetic
Tetracycline	Broad	Bacteriostatic	Natural/semi-synthetic
Quinolones	Broad	Bactericidal	Natural/synthetic
Sulphonamides	Broad	Bacteriostatic	Synthetic
Oxazolidinones	Broad	Bacteriostatic	Synthetic
Nitroimidazole	Broad	Bactericidal	Synthetic
Peptides	Broad	Bacteriostatic	Natural/synthetic

TABLE 1: CLASSIFICATION OF ANTIBIOTICS

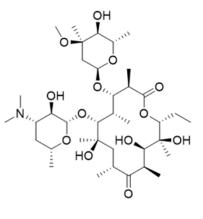
Note: *includes penicillin, carbapenem, cephalosporin, beta-lactamase inhibitors



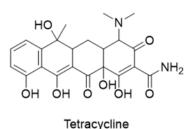
Beta lactum



Aminoglycoside



Macrolide





2-Quinolone



Sulphonamide

Fig. 1: Functional group of major antibiotics

4-Quinolone

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ANALYTICAL METHODOLOGIES

Residual antibiotics must be determined in various matrices such as food, agriculture products, animal husbandry products and portable water. Currently, several detection methods are available to identify the antibiotics from various environmental samples like wastewater, surface water, soil and other samples like milk, honey, etc., However, the right instrument must be chosen for data generation based on its application and detection limitations. The qualitative or rapid screening process and quantitative or confirmatory assays are the two types of detection methods used to determine antibiotics. Infrared detection, oscillographic polarography, and in vitro microbiological methods are generally used to determine the antibiotics qualitatively. Whereas spectrophotometry Ultraviolet methods. including and mass (UV) spectrometry, colorimetry, fluorimetry, conductometry, amperometry titration method, and chromatography methods, are applied for quantitative detection^[14], and sometime EnzymeLinked Immunosorbent Assay (ELISA) methods with fluorescent, chemiluminescent, or radiolabels are used for detection of residual antibiotics^[15]. However, ELISA-based methods suffer from poor specificity issues. Fig. 2 represents the most followed quantitative analytical techniques in the determination of antibiotics in various fields of interest.

The separation technique, such as liquid chromatography, is mostly coupled with several analyzers, such as mass spectrometry, UV detections, Nuclear Magnetic Resonance (NMR) and Fourier Transform Infrared (FTIR). Along with this coupled technology, promising methods such as capillary electrophoresis-mass spectrometry, gas chromatography-mass spectrometery, and matrix-assisted laser desorption ionization-time of flight mass spectrometry are used for quantitative analysis^[16]. Every technology has advantages and disadvantages; however, the applications are used based on the limitations concerning sample preparation.

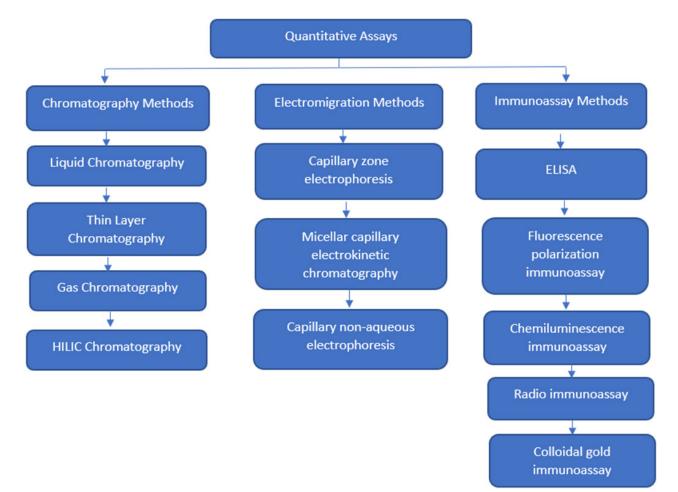


Fig. 2: Various quantitative analytical techniques

A 'nanomaterial-based' aptasensors (aptamer sensor) is used to detect ampicillin. It is a visiblelight-driven self-powered photoelectrochemical sensor with carbon dots^[17]. Aptamers are Deoxyribonucleic Acid (DNA) or Ribonucleic Acid (RNA) oligonucleotides that can bind with high affinity to specific targets. Aptasensors are developed for the detection of specific antibiotics that can be categorized into three types, i.e., electrical. fluorescent. and colorimetric^[18]. However, due to its limitations on the experimental procedure, the technique remains an average methodology from a practical utility point of view.

Another approach in the determination of certain antibiotics is Hydrophilic Interaction Liquid Chromatography (HILIC). This is an emerging yet alternative approach to separate polar compounds through the polar stationary phase^[19]. Though this technology is widely used to determine amino acids, peptides, biomarkers and oligonucleotides, it has limitations such as a high matrix effect, not being completely compatible with Liquid Chromatography-Atmospheric Pressure Chemical Ionization technology (LC-APCI) and being specific to the detection of only polar compounds. Hence the technology remains still under exploration.

Since Liquid Chromatography with Tandem Mass Spectrometry (LC-MS/MS) has a wide range of field applications, such as food, pesticides, pharmaceuticals, and biological, it plays an essential role in confirmatory assays. Tandem mass spectrometry is the most used detection technique with Selected Reaction Monitoring (SRM). However, based on the requirement, the liquid chromatography part can also be attached to other hybrid detection analyzers, such as quadrupole and time of flight and quadropule ion trap^[20], for different applications.

The antibiotic residuals in the unknown samples may be observed in pg/l. So, to determine such low concentrations in all types of matrices such as water, wastewater, aqua products and soil most sophisticated and sensitive equipment like LC-MS/MS is necessary to achieve the desired output. Many ionization techniques are available. However, electrospray ionization is a unique ionization technique where the ions are generated directly from within a solution that is sprayed from a fine needle at atmospheric pressure^[21]. A simultaneous analytical method was developed using an LC-MS/MS to successfully estimate 9 different antibiotics such as cefaclor, cefazolin, cefpodoxime, ceftazidime, cefuroxime, ciprofloxacin, ertapenem, sulfamethoxazole, and trimethoprim. The proposed method was applied for therapeutic drug monitoring in plasma^[22]. The method was sensitive to 0.05 mg/l to 1.5 mg/l for various antibiotics in biological samples. Similarly, another simultaneous method for nine compounds from four different classes of antibiotics, such as beta-lactam, fluoroquinolone, macrolide and nitro-imidazole, with a limit of detection set to 1-10, 0.8 to 10, and 0.8 to 25 ng/l in the treated ground and river water respectively, was developed using LC-MS/MS. This method was applied to determine various antibiotics in water samples^[23]. Interestingly, a simultaneous method was developed to estimate 23 types of antibiotics using LC-MS/MS to determine antibiotics in bovine milk samples^[24]. This method successfully achieved sensitivity as low as 0.17 ng/ml. At least 37 antibiotics from 6 different groups were determined from honey samples using LC-MS/MS with a 5 μ g/kg sensitivity as the lower limit^[25]. Using LC-MS/MS as a platform, simultaneous methods are being developed to estimate antibiotics in various samples and applied in diverse fields of interest. This shows that among different analytical technologies that are approached to determine antibiotics, LC-MS/MS is an exceptional and most sophisticated technology for estimating multiclass antibiotics with low sensitivity. Table 2 shows the list of antibiotics with various details in the respective matrices.

Antibiotica	Processing method	Matrix	LOQ⁵	% Recovery ^c	Analytical system	Reference
Cephalosporins	PPT/SPE	Plasma/water	0.1 to 1 mg/l	>75	LC-MS/MS	[20,21]
Carbapenem	PPT	Plasma	0.5 mg/l	100	LC-MS/MS	[20]
Quinolones	PPT/SPE	Plasma/water/ honey	0.05 mg/l	>72	LC-MS/MS	[20,21]

TABLE 2: ANTIBIOTICS IN VARIOUS MATRICES

Trimethoprim	РРТ	Plasma	0.125 mg/l	>80	LC-MS/MS	[20]
Sulfamethoxazole	PPT	Plasma	1.5 mg/l	>70	LC-MS/MS	[20]
Macrolide	SPE	Water/honey	2 ng/l	>50	LC-MS/MS	[21,23]
nitro-imidazole	SPE	Water	10 ng/l	>45	LC-MS/MS	[21]
Tetracycline	SPE	Bovine milk/ honey	13.60 ng/ml	>80	LC-MS/MS	[22,23]

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Note: a.b.c: The list of antibiotics and their LOQ ranges in respective matrices with the percentage recovery

SAMPLE PROCESSING TECHNIQUES

The scope of sample preparation is to separate the pharmaceutical residues from the sources like water, food and biological matrices. Usually, antibiotic residues are detected at lower levels for various reasons, such as the matrix factor and impurities. To achieve a sensitive method, the sample extraction process must be selective. An appropriate extraction process will lead to a better recovery. Hence, sample processing is the critical step in multi-class analysis. Determining multi-class antibiotics in a single run is a unique strategy, as the extraction procedure has a lot of limitations due to the various physicochemical properties of the compounds^[26]. The sample processing involves three standard extraction techniques described below. However, many other hybrid technologies are also emerging to achieve better sensitivity. Protein Precipitation (PPT) is to separate protein that binds with other substances. Liquid-Liquid Extraction (LLE) is the technique makes a solute immiscible from one solvent to another. This is also known as partitioning. Solid-Phase Extraction (SPE) is the extraction technique adsorbs the analyte onto its solid stationary bed. The adsorption is based on the chemical nature of the compound towards the fixed bed.

PPT:

Though PPT is an inexpensive and cost-effective technology, the extraction method is rapid. However, achieving the low detection limit is difficult as the sample clean-up procedure is not a part of the precipitation technique. Due to this disadvantage, a sensitive method cannot be demonstrated using PPT. However, a combination of PPT followed by liquid-liquid or SPE can be followed to achieve the desired sensitivity. This was proven in the estimation of an new chemical entity molecule following a combinational processing technique^[27]. Initially, the drug was precipitated using a mixture of methanol and acetonitrile along

with formic acid. Post vortex and centrifugation, the supernatant was directly loaded into an Oasis Mixed-mode Cation Exchange (MCX) cartridge. Following this combination precipitation technique, the lower limit of quantitation was achieved as low as 0.5 ng/ml with 100 % recovery. Following the precipitation technique alone, paromomycin was analyzed in human plasma using LC-MS/MS with a limit of quantitation of 5 ng/ ml^[28]. The plasma samples were precipitated using 20 % trichloroacetic acid, and the supernatant was then diluted with water in a ratio of 1:1 for analysis. Using perchloric acid as a precipitating agent, beta-lactam and cephalosporins were determined in human plasma^[29]. Initially, when adding 60 % perchloric acid, it was observed that the compounds were degraded. However, it was overcome by reducing to 40 % perchloric acid along with acetate buffer at pH 4.5 as a medium. To analyze samples from multi-drug resistant tuberculosis patients, a method was designed to analyze at least 20 antitubercular drugs using precipitating solvent as methanol and acetonitrile. With this precipitation method, the inter and intraday precision was observed at 14.3 %, whereas the accuracy was observed between 84.8 %-113.0 **%**^[30]

LLE:

LLE is the second most prioritized technique in sample processing. Due to its better extraction recovery and improved matrix effect than PPT, LLE is considered for multi-class antibiotics analysis. However, extraction recovery cannot be achieved in some cases due to the variable physicochemical nature of the analytes towards the extracting organic solvent. Also, there are several other obstacles such as loss of analytes, low sensitivity, need for a larger volume of extraction solvent, and environmental un-friendly^[31]. Some powerful organic solvents used for this processing technology are methyl-tertiary butyl ether, diethyl ether, dichloromethane, n-hexane,

toluene, and ethyl acetate. The excessive usage of organic solvents is also against the principles of Green Analytical Chemistry (GAC)^[32], which is considered the major drawback of this technique. Other variations in the LLE technique are Liquid-Phase Microextraction (LPME), Salting-out LLE (SALLE), and Aqueous Two-Phase System (ATPS) ^[31]. These methods are followed to reduce the use of organic solvents and to reduce the processing time. However, due to limitations, these methods are less widely used.

A hybrid technique of the Solid-Liquid Extraction (SLE) followed by the SPE method was used to extract certain antibiotics and their transformation products from anthropogenically altered solid matrices^[33]. While environmental the SLE technique was followed to separate analytes from soil sources, the SPE method was followed for the clean-up procedure. However, this combined technique's recovery ranged from 45 %-85 %. A unique extraction technique, SALLE was followed to estimate ciprofloxacin using high-performance liquid chromatography-diode array detector^[34]. Magnesium sulphate was used as a salting-out agent for this processing technology. However, this method is applied only to determine one compound from the soil samples. The main advantage of using the SALLE technique is improved extraction efficiency of polar compounds using polar solvents. The combination of LLE and dispersive-LLME showed an efficient extraction using acetonitrile as an initial extracting solvent. Followed by this, a newly synthesized water-immiscible deep eutectic solvent was used for subsequent extraction steps. Using the ion mobility spectrometry instrument, this method was applied to estimate oxytetracycline, penicillin G, and tilmicosin from sausage samples^[35]. This instrumental technique characterizes molecules based on the gaseous phase mobility of their ions formed at ambient pressure and under an electric field. With this new combination technique, the limit of quantification was achieved at around 5.1 ng/g. An integrated PLE and SPE extraction technique was followed to extract antibiotics from soil samples^[36]. In this method, the PLE was followed for extraction and diol SPE cartridges for the clean-up process. The method successfully achieved as low as 0.2 μ g/kg with a recovery rate ranging from 38 %-94 % for macrolides, 76 %±24 % for ionophore salinomycin, and 118 %±21 % for pleuromutilin tiamulin using LC-APCI-MS/MS.

SPE:

SPE is the most widely used processing technology. Due to its reliability, reproducibility, a wide range of applications and use of compound-target stationary phase cartridges, the SPE method is the most familiar technique among scientists. SPE techniques include solid-phase microextraction, dispersive micro-SPE, magnetic SPE and matrix solid-phase dispersion^[5,23,37,38].

Before selecting sorbents, it is important to check the pKa values of the selected antibiotics. pKa is a method used to identify the strength of the acid. It is a negative value of the acid dissociation constant. A lower pKa value indicates a stronger acid. pKa values for some critical antibiotics are provided in Table 3. Many antibiotics have acidic, and some have basic functional groups. Hence, pH plays a vital role in the extraction recovery of antibiotics. While the pH can affect the stability and interaction of analytes with sorbents, the same can also influence the speciation of antibiotics due to their acidic and basic functional groups in their structures^[39]. Since antibiotics are acidic in general, these are in a 'soluble' form at a lower pH. Hence, the pH should be maintained lower than the pKa value to achieve a better extraction recovery^[40].

Classification ^a	Compound	Molecular weight	Molecular formula	pKa value⁵
Beta-lactam	Amoxicillin	365.4	$C_{16}H_{19}N_{3}O_{5}S$	3.32
Cephalosporin	Cephalexin	347.39	$C_{16}H_{17}N_{3}O_{4}S$	4.5
Fluoroquinolones	Ciprofloxacin	331.35	C ₁₇ H ₁₈ FN ₃ O ₃	5.76
Macrolides	Azithromycin	748.98	$C_{38}H_{72}N_{2}O_{12}$	8.74
Nitroimidazole	Metronidazole	171.15	$C_6H_9N_3O_3$	15.44
Aminoglycoside	Streptomycin	581.58	C ₂₁ H ₃₉ N ₇ O ₁₂	11.09

TABLE 3: pKa VALUES FOR KEY ANTIBIOTICS

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Oxazolidinone	Linezolid	337.35	$C_{16}H_{20}FN_{3}O_{4}$	1.8
Carbapenem	Meropenem	383.5	$C_{17}H_{25}N_{3}O_{5}S$	2.90 for Carboxylic acid and 7.4 for Pyrrolidinyl amino
Azole derivatives	Voriconazole	349.31	$C_{16}H_{14}F_{3}N_{5}O$	1.76
Sulphonamides	Trimethoprim	290.32	$C_{14}H_{18}N_4O_3$	7.12
Tetracycline	Tetracycline	444.4	$C_{22}H_{24}N_2O_8$	3.3

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Note: a,b : Commonly used antibiotics with their molecular weight and the respective pKa values

Various types of SPE sorbents are being used in the extraction technique. Also, each cartridge has a distinctive feature relating to the molecule. The various SPE sorbents are categorized into three prominent SPE techniques: polar, non-polar, and ion exchange SPE. The different types of cartridges and their applications are tabulated in Table 4^[41]. Among every other cartridge C18, HLB, MCX, and Mixed-mode Anion exchange (MAX) are the most used cartridges in all applications. However, the most widely used cartridges are HLB specific to antibiotics determination. This is because the HLB cartridge shows both hydrophilic and lipophilic retention features. And hence, both polar and nonpolar compounds can be retained in the sorbent.

Hybrid SPE-Phospholipid columns were used to determine 32 antibiotics in fish and shrimp muscles^[42]. The recovery for these antibiotics ranges between 70 %-120 %, with the lower limit of quantitation at 0.062 μ g/kg. This method was successfully applied to detect various antibiotics in aquaculture products. Antibiotics and some antivirals were determined in water using HLB, MCX, and MAX cartridges. With the LOQ set at 5 ng/l, the recovery was observed between 41 %-116 %. Following this method to detect antibiotics in wastewater, almost all target compounds were observed in the range between 10 and 570 ng/l^[43]. However, a comparison study between Oasis HLB and Strata-X^[44] for the determination of 18 different classes of antibiotics was performed with an optimized pH. It was observed that the best extraction efficiency was achieved in Oasis HLB in acidic pH. The LOQ for the antibiotics ranged between 0.8 ng/l-245.1 ng/l. This method determined all proposed compounds in wastewater samples up to 6.2 µg/l concentration. An optimized Solid-Phase Microextraction (SPME) technique was used to determine sulphonamides, macrolides, and trimethoprim with the LOQ set at 16 and 35

ng/l for influent and effluent samples. To compare the results, the SPE method's LOQ was set at 4.7 and 0.86 ng/l for the same samples^[45]. The main advantages of the SPME method are decreased sample volume, less processing time, costeffectiveness, reduced matrix effect and repeated usage. A similar comparison study between the SPME and SPE techniques to determine 10 sulphonamide antibiotics^[46] in wastewater also confirms the advantages of the SPME method with the added advantage of good extraction efficiency. A study specifically compared the extraction efficiency of HLB and MCX cartridges used in the determination of certain antibiotics and their metabolites in water sources using linear-ion trap tandem mass spectrometry^[47]. This study further proved that HLB cartridges showed better recovery and lesser matrix effect compared with MCX cartridges. This was succeeded as the HLB cartridges contain the composition of lipophilic divinylbenzene units and the hydrophilic N-vinylpyrrolidone units^[48], allowing the efficient extraction of organic contaminants in a wide range of pH (from pH 1 to 14). Before extraction, a chelating agent such as sodium ethylenediaminetetraacetic acid must be used to complex the metals or multivalent cations (residual metal ions) soluble in water on SPE cartridges and glassware. The key advantages and disadvantages of the above-mentioned extraction techniques are provided in Table 5.

Irrespective of the applications, the SPE technique is being followed widely for sample clean-up making the method free from the matrix effect and achieving the expected recovery. Researchers applied the developed analytical method for determining multi-class antibiotic residuals in various samples such as aquaculture products, meat, raw milk, wastewater and water resources, honey and soil.

TABLE 4: TYPES OF SPE CARTRIDGES

Types of Sorbent ^a	Structure ^b	Functional group ^c	Mechanism ^d	Applications
Anionic exchange (mixed mode-SAX)		C8	Reverse phase or anion exchange selectivity	Suitable for an alkaline aqueous solution or neutral compounds in urine, and blood
Cationic exchange (mixed mode-SCX)		C8	Reverse phase or cation exchange selectivity	Suitable for an alkaline aqueous solution or neutral compounds in urine, and blood
Amino Cartridges	NH2 Sime	NH2	Strong polar adsorption in non-polar organic solvents	Suitable to analyze peptides, drugs, mono and polysaccharides, steroids, and cholesterol. Widely used in food, environmental, pharmaceutical, and medical applications
Cyanopropyl Cartridges		CN	Weak hydrophobic silica matrix used as normal or reverse phase adsorbent	Suitable to analyze drugs, pesticides, veterinary drug residues on food and dairy products, oil pollution, and pesticides in environmental samples
C18 Cartridges	¥	C18	Retain non-polar compounds by strong hydrophobic interaction	Suitable to analyze drugs, poisons, pollutants, mycotoxins in food, preservatives in cosmetics
HLB Cartridges	Hydrophilic monomer	Monomers of N-vinylpyrollidone and divinylbenzene	Universal sorbent for acidic, basic, and neutral compounds. Enhanced reverse- phase capability with retention of polar analytes	Suitable to analyze drugs, environmental samples, agrochemical, and food samples

Note: ^{a,b,c,d}: Widely used SPE cartridges and their specifications, along with structures, functional groups, etc

TABLE 5: ADVANTAGES AND DISADVANTAGES OF VARIOUS PROCESSING TECHNIQUES

Advantages	Disadvantages	
1. Crude processing technology	1. High matrix effect	
2. Mostly used in drug discovery for faster processing	2. The matrix effect may impact recovery	
3. Cost-effective	 High chances of blockage due to precipitated particles in the supernatant 	
4. The extraction solvent is either methanol or acetonitrile	4. Affects HPLC column performance	
1. Higher recovery than the PPT technique	1. Longer processing time	
2. Lesser matrix effect	2. More chances of inhaling organic solvents	
	 Crude processing technology Mostly used in drug discovery for faster processing Cost-effective The extraction solvent is either methanol or acetonitrile Higher recovery than the PPT technique 	

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	3. Better sensitivity due to concentrating the sample	3. Not following green analytical chemistry	
	1. Alternative for PPT and LLE	1. Expensive	
SPE	2. Sample clean-up	2. Single-use cartridges	
	3. Lesser matrix effect and higher recovery	3. Many steps in the extraction process	
	4. Analyte target sorbent can be used	4. Complex methodology	

CONCLUSION

This review concludes that various analytical methodologies and sample processing techniques are key factors in determining sub-lethal levels of antimicrobials in environmental sources. The methods must be sensitive enough to detect pharmaceutical ingredients in the respective samples. SPE techniques play a major role in determining antimicrobials in diverse samples compared to low-sensitive methods. The key challenges include simultaneous estimation, varied polarities of the compounds, acid dissociation constant (pKa) of varied classes of antimicrobials and other factors such as insoluble matters in the environmental samples.

Since the concentration of residual compounds is usually very low in various environmental samples, achieving a sensitive method is necessary. LC-MS/MS, or any other high-resolution mass spectrometry, is the most preferred analytical instrument to achieve the required sensitivity. The developed and validated analytical method must provide the concentration of the antimicrobials present in the environmental samples. This will help detect and take necessary actions to eliminate trace contaminations from required sources such as wastewater, surface water, soil, honey, milk products, poultry products, and aquaculture products.

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Conflict of interests:

The authors declared no conflict of interests.

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