

RP-HPLC Method for Estimating Norethindrone in Plasma and Tissues Following Administration of a Controlled Release Nanoparticulate Liquid Formulation

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Al-Nimry *et al.*: RP-HPLC method for determination of Norethindrone

The purpose of this investigation was to develop and validate a simple reversed phase-high-performance liquid chromatography method coupled with UV detector estimating norethindrone in plasma and tissues. A Thermo Scientific C18 column (250×4.6 mm ID, 5 µm pore size) and a mobile phase consisting of deionized water:acetonitrile (60:40, v/v) were used. The flow was isocratic at a rate of 1.3 ml/min and the wavelength of detection was 245 nm. Estradiol was used as internal standard. Validation of linearity, accuracy and precision, limits of detection and quantification, specificity and recovery was carried out according to the International Conference of Harmonization guidelines. The method was used in estimating the bioavailability of a controlled release nanoparticulate liquid formulation, administered to dogs. It was also used estimating adhesion potential of the same formulation to the GIT of mice. The method was simple and only liquid-liquid extraction was used in case of plasma samples. However, sample preparation was more complex in the case of tissue samples. It was linear in the range studied, accurate and precise. It was specific and the excipients used in preparing the formulation did not interfere with the method. The recovery was high and consistent. The bioavailability of the drug was enhanced after the administration of the nanoparticulate liquid formulation as compared to unformulated drug. The formulation adhered to the stomach and intestines for 48 h. After 48 h the concentration reached undetectable levels. In conclusion, a simple, linear, accurate and precise reversed phase-high-performance liquid chromatography method coupled with UV detection was developed, validated, and used successfully for the determination of norethindrone in plasma and adhesion to GIT tissues. The cost of the analysis was expected to be low since the extraction procedure was simple and no radiolabeled internal standard was used.

Key words: Norethindrone, RP-HPLC, concentration in plasma, adhesion to tissue, animal models

Norethindrone is a synthetic progesterone (progestin) that belongs to a family of 19-nortestosterone derivatives^[1]. The structure of norethindrone is shown in fig. 1^[2]. Norethindrone can be used in combination with some oral contraceptive pills or progestin-only pills. Also, it is useful for treatment of certain illnesses and diseases such as premenstrual syndrome, irregular or painful menstruation, abnormal heavy bleeding, to postpone a period or menopausal syndrome (with estrogen). It is also generally used to help in prevention of uterine hemorrhage in complicated non-surgical or pre-surgical gynecologic cases and in treating non responsive cyclical mastalgia^[3].

Several methods for the determination of norethindrone alone or with other drugs/metabolites in various dosage forms have been reported. These methods

could not be applied as such for the determination of norethindrone in plasma. Norethindrone should be separated from plasma proteins. Also, several methods for the determination of norethindrone in plasma have been developed. A linear high-performance liquid chromatography (HPLC)-UV method in the range 2-20 ng/ml was developed. ³H-norethindrone radiolabeled internal standard was used and plasma samples were extracted with a mixture of hexane-methylene chloride^[4]. Another method for the

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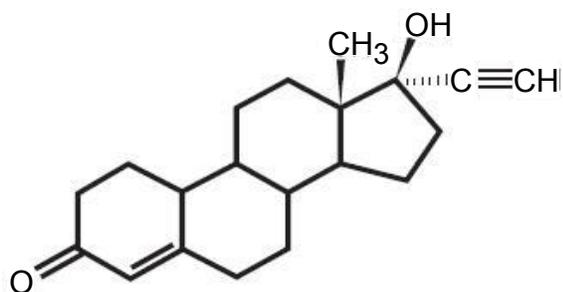


Fig. 1: Structure of norethindrone^[2]

simultaneous determination of norethisterone and 6 metabolites in human plasma by capillary gas chromatography-mass spectrometry (GC-MS) detection was developed. The compounds were determined in plasma after enzymatic hydrolysis. After addition of norgestrel as an internal standard, the compounds were extracted from plasma at pH 5 using an Extrelut column and elution with dichloromethane. After evaporation, the compounds were converted into bis trimethylsilyl derivatives, which were determined by gas chromatography using a mass-selective detector^[5]. A HPLC-MS/MS method for the simultaneous determination of norethindrone and ethinyl estradiol in plasma was reported. A derivatizing reagent was used to enhance the sensitivity. The lower limit of quantitation (LLOQ) was 50 and 2.5 pg/ml for norethindrone and ethinyl estradiol, respectively. Gradient flow of two mobile phases was used to achieve optimal separation of these 2 compounds. Norethindrone-¹³C₂ and dansylated ethinyl estradiol-d₄ were used as internal standards^[6]. Two semi-automated, relatively high throughput methods using ultra performance liquid chromatography (UPLC) coupled with MS/MS were developed and validated for the simultaneous determination of ethinyl estradiol in combination with either 19-ethindrone or levonorgestrol in human plasma. The concentration ranges were 0.01-2, and 0.1-20 ng/ml for ethinyl estradiol and 19-ethindrone or levonorgestrol, respectively. These methods involved liquid-liquid extraction (LLE) and derivatization followed by separation on a small particle size column achieving a run time of 2.7 min. Radiolabeled internal standards were used^[7]. Another UPLC-MS/MS method for determination of norethindrone alone in plasma samples over the range of 50-25000 pg/ml was developed and validated. Samples were prepared by supported liquid extraction procedure and norethindrone-¹³C₂ was used as the internal standard^[8]. No specific method was available in literature for determination of norethindrone in GI tissues.

Most of the methods documented above were complicated. In some of these methods sample preparation involved enzymatic hydrolysis followed by column extraction and derivatization. In others it involved LLE, derivatization and separation with small particle size column. Some of these methods involved the use of two mobile phases with gradient flow and some used tandem MS (MS/MS) detection. Additionally, most of these methods used radiolabeled internal standards, which increases the cost of the analysis. Usually these methods are used for the analysis of a large number of samples.

The purpose of this work was to develop and validate a simple, low cost, HPLC method coupled with a UV detector for the determination of norethindrone in plasma and tissues. Sample preparation involved only LLE in case of plasma samples. However, sample preparation from tissue samples became complex. Estradiol used was used as an internal standard. The developed method was used to study the bioavailability of norethindrone after *in vivo* administration of a previously prepared nanoparticulate liquid formulation (NLF) to dogs^[9]. It was also used to determine the adhesion potential of the formulation to stomach and intestine after *in vivo* administration to mice.

MATERIALS AND METHODS

19-Norethindrone (≥ 98 %), subtilisin, (2-hydroxypropyl)- β -cyclodextrin (HP- β -CD) and polyvinyl pyrrolidone (PVP) K30 were supplied by Sigma-Aldrich, USA. β -estradiol, tert-butyl methyl ether (TBME, 99 %) and anhydrous dichloromethane were supplied by Across Organics. High molecular weight chitosan (HMWC), 600-800 kDa was supplied by Shanghai Hanshare Industry CO., China. HPLC grade acetone and methanol were supplied by Fisher Scientific, UK. Ammonia solution was supplied by BDH Chemicals, England. Absolute ethanol was supplied by Solvochem, Holland. HPLC grade acetonitrile was supplied by Anhani Fulltime Specialized Solvents and Reagents Co., China. HPLC grade cyclohexane was supplied by Lab-Scan Analytical Science, Ireland. 0.45 μ m Sartolon polyamide cellulose membrane filters were supplied by Sartorius Stedim Biotech, Germany. Tris base was purchased from Bio Basic Inc. (Ontario, Canada). Deionized water prepared in the laboratories of Jordan University of Science and Technology was used throughout the study. Low molecular weight chitosan (LMWC) was prepared by acid hydrolysis of HMWC.

Liquid chromatographic conditions:

The analysis of norethindrone was carried out using a Merck Hitachi La Chrom HPLC system, USA. It consisted of a UV/Vis detector (L-7455), an auto sampler (L-7200), an elution pump (L-7150) and a degasser (L-7612). The chromatographic separation was done at room temperature on Thermo Scientific C18 (250×4.6 mm ID, 5 µm pore size) column. The mobile phase was composed of deionized water:acetonitrile (60:40, v/v). It was filtered through a 0.45 µm regenerated cellulose membrane filter prior to use. The flow rate was 1.3 ml/min. The injection volume of sample was 100 µl. The wavelength of the detection was at 245 nm.

Preparation of calibration standards:

A stock solution (100 µg/ml) was prepared by dissolving 10 mg of norethindrone in 100 ml of ethanol. Seven standard solutions were prepared by appropriate dilution of the stock solution with plasma, which were, 0.04, 0.08, 0.16, 0.31, 0.63, 1.25 and 2.50 µg/ml. Estradiol was used as internal standard. A stock solution (1000 µg/ml) was prepared by dissolving 10 mg of estradiol in 10 ml ethanol.

Liquid-liquid extraction of plasma samples:

Five hundred microlitres of plasma was spiked with 20 µl of estradiol (internal standard 1000 µg/ml) and vortexed. 3 ml of dichloromethane:hexane (40:60, v/v) was added to the plasma sample and vortexed. The sample was centrifuged for 4 min at 4000 rpm. Appropriate amount of the organic phase was transferred to a 5-ml test tube, evaporated in a concentrating dryer at 35° with a cold trap for about 1 h. The residue was immediately reconstituted with 300 µl of mobile phase and analyzed using the developed and validated RP-HPLC method. The RP-HPLC method of analysis for norethindrone was validated in terms of linearity, accuracy and precision, Limit of detection (LOD) and limit of quantitation (LOQ), specificity and recovery according to the ICH Guidelines^[10].

Linearity was determined by injecting the 7 prepared standard solutions and plotting peak area ratio of norethindrone to estradiol versus norethindrone concentrations to construct a calibration curve. Six calibration curves were prepared on different days. The correlation coefficient (R), y-intercept, and the slope of the regression line were computed. The results were analyzed using one-way analysis of variance (ANOVA)

at a significance level of $\alpha = 0.05$ (95 % of confidence interval) using JMP statistical analysis software.

Accuracy and precision of the method were evaluated on 3 consecutive days by preparing QC samples of norethindrone at 3 concentration levels, 0.05 (low), 0.5 (medium) and 2 µg/ml (high) from the stock solution. The results were analyzed by one-way ANOVA test at a significance level of $\alpha = 0.05$ (95 % of confidence interval) using JMP statistical analysis software.

LOD and LOQ were calculated using Eqns., $LOD = (3.3 \times \sigma) / s$ and $LOQ = (10 \times \sigma) / s$, where, σ is the standard deviation of the response and s is the slope of the calibration curve. Specificity was evaluated to ensure that the peaks of the excipients used in the preparation of the NLF (LMWC, HP- β -CD and PVP K30) were resolved from the peak of the drug. Samples were spiked with the appropriate levels of these excipients and injected to demonstrate that the assay of the drug was not affected by their presence. Recoveries of norethindrone and estradiol were evaluated by comparing peak areas in dog plasma before and after extraction. The recovery of norethindrone was determined at 3 concentration levels (0.05, 0.5 and 2 µg/ml) and the recovery of estradiol was determined at 20 µl (1000 µg/ml). Two sets of samples were prepared. In the first set (before extraction) blank plasma samples were spiked with the drug solution (0.05, 0.5 and 2 µg/ml) and 20 µg/ml of estradiol (1000 µg/ml) then extracted as mentioned above. In the second set (after extraction) the blank samples were first extracted and the residue was spiked with norethindrone and the internal standard, reconstituted with the mobile phase, and injected. NLF was prepared and the details of the preparation and optimization of the NLF were published previously^[9].

Bioavailability study in the dog:

All experiments were carried out in accordance with Institutional Guidelines Animal Use for Jordan University of Science and Technology. Oral bioavailability of the NLF was evaluated using a single dose cross over study design. Six dogs (8 mo old) weighing about 30 kg and from Kinnani family were used. The dogs were randomly divided into 2 groups, 3 dogs were orally administered 10 ml of the test formulation (NLF) and the other 3 dogs were orally administered 10 ml of the reference formula (norethindrone dissolved in ethanol:water solution (40:50, v/v) using a syringe. The norethindrone dose

for both groups was 0.1455 mg/ml. About 2 ml blood samples were collected from the suborbital vein with a heparinized syringe from both groups at predetermined time intervals (0, 0.5, 1, 2, 4, 6, 8, 12, 24, 48, 72 and 96 h). Blood samples were immediately centrifuged at 5000 rpm for 5 min. The plasma obtained was stored in Eppendorf tubes at -20° until analysis. The procedure was repeated after 2 w where the first group received the reference drug formula and the second group received the test formulation in a cross-over design. The analysis was carried out using the developed and validated RP-HPLC method. Plasma concentration-time profiles for the NLF and norethindrone were constructed

Adhesion to GIT in mice:

The adhesion potential of the NLF was evaluated in 4 mo old mice (*Mus musculus*) weighing about 28 g. Twenty mice were randomly divided into 5 groups of 4 mice each. All mice were orally administered about 1 ml of the test formulation containing 0.16 mg of norethindrone using a feeding tube. The mice were subjected to light ether anesthesia. The stomach and intestine were collected from each mouse at predetermined time intervals and stored at -20° until analysis. The time intervals after administration were, group 1 2 h, group 2 24 h, group 3 48 h, group 4 72 h and group 5 after 96 h of administration. Norethindrone that had attached to the tissue of the stomach or intestines was extracted according to method developed by Kaklamanos *et al.*^[11]. Tissue sample were subjected to enzymatic hydrolysis, extracted with tert-butyl methyl ether, defatted and subjected to solid phase extraction (SPE). The resultant residue was reconstituted with 300 μ l of methanol and analyzed using the validated HPLC-UV method.

RESULTS AND DISCUSSION

Several researchers used radiolabeled internal standards for analyzing contraceptive hormones, but this increased the cost of the analysis. The radiolabeled compounds could be replaced by analog hormones for contraception^[5,12]. β -estradiol was selected as the internal standard due to the structural similarity to norethindrone. The injection volume, for the size of the column used, 4.6 ID \times 250 mm, usually ranges between 0-40 microliter. However, a larger volume (100 μ l) was used in this study. This increased the sensitivity of the method without adversely affecting the peak shape, symmetry, and resolution. The peaks of norethindrone and the internal standard (estradiol)

were sharp, symmetrical, and well resolved even at the highest concentration. The peak of estradiol appeared at a retention time of 7.23 ± 0.2 min and the peak of norethindrone at 9.04 ± 0.2 min (fig. 2).

Validation of an analytical procedure is a proof that it is suitable for its intended use. The method of analysis was validated according to the ICH guidelines^[10] in term of linearity, accuracy, precision, specificity, sensitivity, LOD and LOQ. Linearity of an analytical method describes its ability to give results for conducted test that are directly proportional to the quantity of analyte in the sample^[10]. It is generally evaluated by plotting the area ratio (area under the peak of the drug to that under the peak of the internal standard) against the concentration of the drug. From the plot, correlation coefficient, y-intercept, and slope can be obtained. Linearity in this investigation was evaluated by injecting 7 standard solutions (0.04, 0.08, 0.16, 0.31, 0.63, 1.25 and 2.50 μ g/ml) into the system. Six calibration curves were injected on different days and the data were represented in Table 1. The coefficient of correlation (R) value was greater than 0.999 for all the 6 calibration curves, which is generally considered as evidence of acceptable fit of the data to the regression line^[13]. The linear fit for the obtained data is shown in fig. 3. The results of the ANOVA test for the calibration curves are shown in Table 2. The p-value >0.05 indicated that the null hypothesis is not rejected and that there was no significant difference between the calibration curves run on different days.

Accuracy of an analytical method measures the closeness of agreement between the resulted practical value obtained from the test and the expected real theoretical value of the analyte in the sample. The ICH guidelines recommend for accuracy assessment using a minimum of 9 determinations over a minimum of

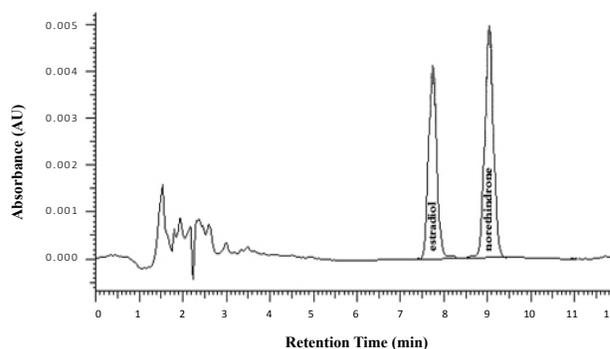


Fig. 2: Chromatogram of spiked plasma sample. Chromatogram of a plasma sample spiked with 2 μ g/ml estradiol (internal standard) and 0.63 μ g/ml norethindrone.

3 concentration levels, covering the specified range (i.e., 3 concentrations and 3 replicates of each concentration)^[10]. It is calculated as percent recovery^[14] using the Eqn., % accuracy = 100-((theoretical value-experimental value/theoretical value)×100)

The precision of an analytical procedure is the degree of agreement among individual test results when the procedure is performed repeatedly to multiple samplings of a homogeneous sample. The precision of an analytical procedure is usually analyzed by examining the standard deviation or relative standard deviation (coefficient of variation) of a series of measurement. According to the ICH, precision should be performed at three different levels; repeatability, intermediate precision (inter-day precision) and reproducibility (between laboratories precision)^[12]. Repeatability is the intra-day variation in assay obtained at different concentration levels using the same analyst with the same equipment expressed in terms of RSD calculated for each day. Intermediate precision is the inter-day

variations expresses within-laboratory variation, as on different days, or with different analysts or equipment within the same laboratory^[15]. Reproducibility is the precision between laboratories (collaborative or inter-laboratory studies)^[16]. Precision is expressed statistically as standard deviation (SD) or coefficient of variation (CV) or the relative standard deviation (RSD) which is calculated using Eqn. 4, % RSD= standard deviation/average×100.

Typically, for assay methods, RSD between the two sets of data must be ≤2.0 %^[13,17]. Acceptance criteria similar to those applied to intermediate precision also apply to reproducibility^[16]. The intra- and inter-day accuracy and precision were determined using six QC samples at three concentration levels (QC low (0.05 µg/ml), QC medium (0.5 µg/ml) and QC high (2 µg/ml)). These QC samples were prepared and injected on three consecutive days. Additionally, 6 LLOQ samples were injected on the first day of the validation. The results are shown in Table 3. The

TABLE 1: LINEARITY DATA AND PARAMETERS OF NORETHINDRONE CALIBRATION ON DIFFERENT DAYS

	Concentration (µg/ml)						Slope	Intercept	R ²
	0.04	0.08	0.16	0.31	0.63	1.25			
	Area Under Peak (AU)								
1	0.0780	0.1488	0.3665	0.6811	1.2521	2.5724	4.8677	1.949	0.9991
2	0.0795	0.1511	0.3887	0.6799	1.2611	2.5833	4.9881	1.992	0.9995
3	0.0882	0.1661	0.3555	0.6541	1.2522	2.5711	5.0254	2.007	0.9998
4	0.0755	0.1477	0.3551	0.6832	1.2384	2.4611	4.779	1.905	0.9995
5	0.0722	0.1495	0.3761	0.6689	1.2433	2.5412	4.8412	1.936	0.9993
6	0.0912	0.1578	0.3489	0.6381	1.2577	2.5551	4.8512	1.943	0.9994
Mean	0.0808	0.1535	0.3651	0.6676	1.2508	2.5474	4.8921	1.9553	0.9994
SD	0.0074	0.0071	0.0151	0.0181	0.0086	0.0448	0.0945	0.0377	0.0074
RSD	9.1738	4.6500	4.1254	2.7056	0.6853	1.7574	1.9313	1.9290	20.4994

TABLE 2: RESULTS OF ANOVA FOR THE LINEARITY DATA OF NORETHINDRONE

Source of variation	DF	SS	MS	F Ratio	Probability > F
Between groups	1 Model	0.0160	0.003	0.001	1.0000
Within group	40 Error	110.6318	3.0735		
Total	41 C. Total	110.6474			

DF-degrees of freedom; SS-sum of squares; MS-mean square; *p<0.05

TABLE 3: INTRA-DAY ACCURACY AND PRECISION AT LOQ AND INTRA- AND INTER-DAY ACCURACY AND PRECISION AT DIFFERENT CONCENTRATION LEVELS

QCs	Concentration (µg/ml)	Intra-day assay			Inter-day assay		
		Mean	Accuracy	Precision	Mean	Accuracy	Precision
LLOQ	0.04	0.04	100.10	0.14			
Low QC	0.05	0.05	100.00	1.76	0.05	100	0.38
Mid QC	0.50	0.50	100.00	1.85	0.50	100	0.2
High QC	2.00	2.02	100.01	1.62	2.02	100.02	0.36

Intra-day accuracy and precision at the lower limit of quantitation (LOQ) and intra- and inter-day accuracy and precision at different concentration levels, N=6

intra-day accuracy and precision at the LLOQ were 100.10 % and 0.14, respectively. The intra-day accuracy and precision at the low QC, mid QC, and high QC ranged between 100.00-100.01 % and 1.62-1.85, respectively. The inter-day accuracy and precision at the low QC, mid QC, and high QC ranged between 100.00-100.02 % and 0.20-0.38, respectively. This indicated that the accuracy and precision at the LLOQ and at the three concentration levels (intra-and inter-day) met the acceptance criteria^[13,18-20]. The results of the ANOVA test for the QC samples are presented in Table 4. The p-value >0.05 (0.7411 for QC high and 0.7281 for QC mid) indicated that the null hypothesis is not rejected and that there was no significant difference between the QC samples run on the same day or on different days.

LOD of an analytical procedure is the lowest amount of analyte in a sample which can be detected but not necessarily quantitated as an exact value. The LOQ is the lowest amount of analyte in a sample which can be quantitatively determined with suitable precision and accuracy^[10]. For norethindrone method of analysis, LOD and LOQ were estimated from the standard deviation of the blank. The calculated LOD and LOQ were 0.0154 and 0.0264, respectively. Experimental value of the LLOQ was 0.04 µg/ml which is the lowest

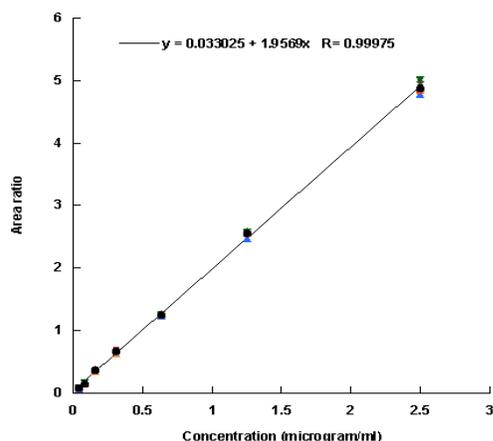


Fig. 3: Linear fit for 6 calibration curves of norethindrone on different days

point in the calibration curve. Low values of LOD and LOQ indicate the sensitivity of the method.

Specificity is the ability of the method to accurately measure the analyte in the presence of other different molecules which may be expected to be present. These molecules might include impurities, degradants, excipients^[12]. In this study norethindrone was prepared as a disperse system containing different ingredients and polymers. In order to evaluate the specificity of the method, the main three excipients, LMWC 11 kDa, HP-β-CD and PVP K30 were dissolved separately in an aliquot of plasma free of norethindrone and subjected to the same extraction method and injected into the HPLC system. No peak was observed at the retention time of norethindrone in all samples, which indicated the specificity of the method (fig. 4).

Recoveries of norethindrone and estradiol were evaluated by comparing peak areas from dogs' plasma before and after extraction. The latter represents 100 % recovery. The average recovery of norethindrone was determined at three concentrations levels (0.05, 0.5 and 2 µg/ml) and of estradiol at 20 µg/ml (Table 5). The recovery was consistent and high (>96.28 %).

The plasma concentration-time profiles of the NLF and norethindrone are represented in fig. 5. It indicated enhanced bioavailability of the drug from NLF as compared to norethindrone alone. Concerning the adhesion study, samples were obtained at different time intervals following the administration of about 1 ml of the formula containing 0.16 mg of norethindrone for the twenty mice. The chromatographic conditions were the same as in the case of plasma samples, but the sample preparation was different. It involved enzymatic hydrolysis, extraction with tert-butyl methyl ether, defatting, and SPE. This was expected due to the nature of the sample, which was more complex than plasma. The results are shown in Table 6. Results indicated the ability of the NLF to adhere to the stomach and intestines for 48 h. The concentration of norethindrone

TABLE 4: INTER- AND INTRA-DAY ACCURACY AND PRECISION AT DIFFERENT CONCENTRATION LEVELS

	Source of variation	DF	SS	MS	F Ratio	Probability > F
QC Mid	Between groups	2 Day	0.00002	0.00001	0.064	0.9382
	Within group	15 Error	0.0013	0.00009		
	Total	17 C	0.0013			
QC High	Between groups	2 Day	0.0009	0.0005	0.400	0.6775
	Within group	15 Error	0.0175	0.0012		
	Total	17 C	0.0185			

DF-degrees of freedom; SS-sum of squares; MS-mean square; *p<0.05

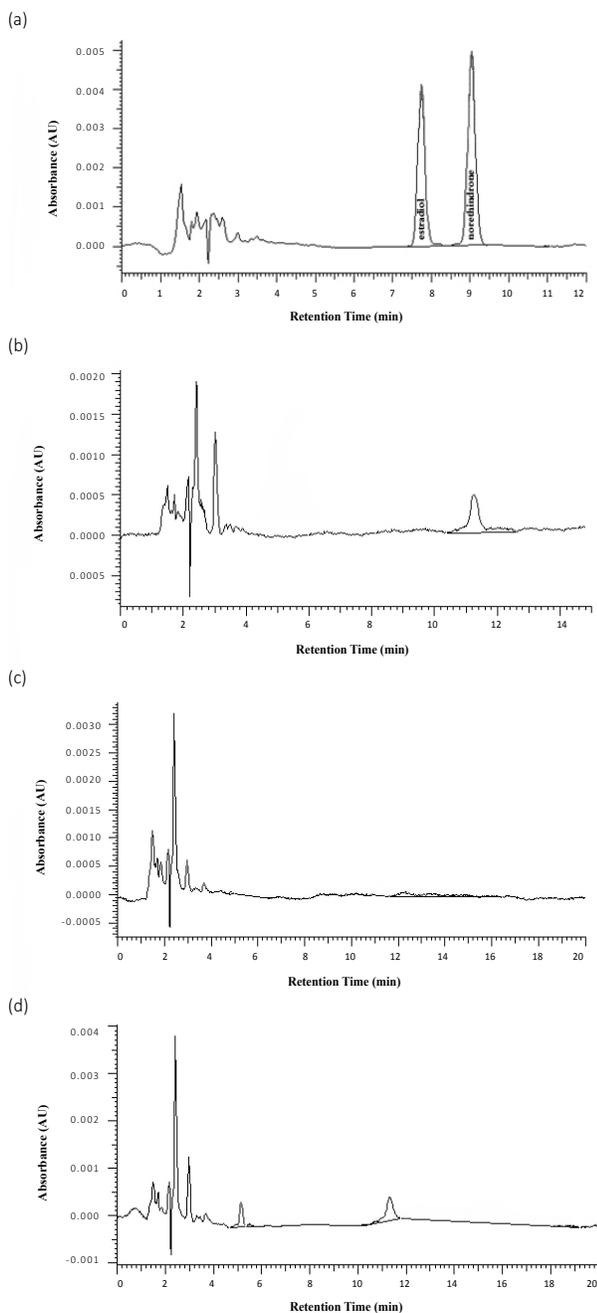


Fig. 4: Chromatograms indicating specificity of the HPLC method (a) Estradiol internal standard (2 $\mu\text{g/ml}$) and norethindrone (0.63 $\mu\text{g/ml}$) (b) Chitosan (c) HP- β -CD and (d) PVP K30 in plasma

in these organs decreased with time and almost reached undetectable level after 48 h. However, analysis of dog plasma indicated the presence of norethindrone for 96 h.

In conclusion, a new RP-HPLC method coupled with UV detection was developed and validated for the determination of norethindrone in plasma and tissue. The method was simple, the sample preparation from plasma involved only LLE, the elution was isocratic and a single detector was used. It was linear in the range studied, and the statistical analysis proved it was accurate and precise. The excipients used in the formulation did not interfere in the analysis of norethindrone. The recoveries of norethindrone and the internal standard from plasma were high and consistent. The cost was expected to be low as compared to the other reported methods specially those using complex sample preparation procedures and radiolabeled internal standards. The method was successfully applied to a bioavailability of norethindrone after administration of a NLF and the adhesion potential to GI tissue in animal model.

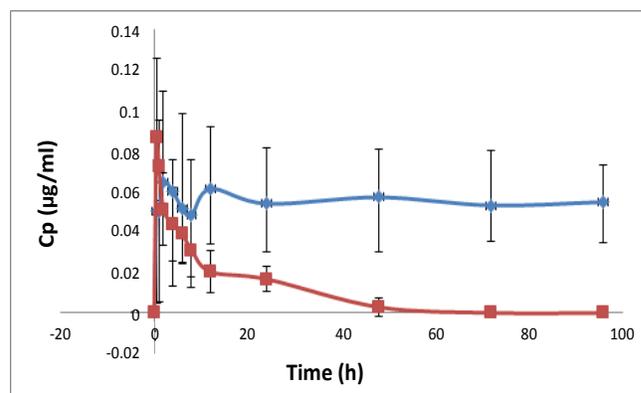


Fig. 5: Plasma concentration time profile of norethindrone given orally to dogs

Average plasma concentration time profile of norethindrone after administration of 10 ml (0.1455 mg/ml) of the pure drug or the test formulation orally to dogs (n=6), (—♦—) nanoparticle formulation of norethindrone; (—■—) raw norethindrone

TABLE 5: EXTRACTION RECOVERY OF NORETHINDRONE AT DIFFERENT CONCENTRATION LEVELS AND ESTRADIOL

Substance	Concentration ($\mu\text{g/ml}$)	Peak area		% Recovery	Coefficient of Variation
		Samples spiked before extraction)	Samples spiked after extraction)		
Norethindrone	0.05	8911	7846	113.57	5.12
	0.50	57522	59492	96.69	7.42
	2.00	201626	209423	96.28	9.11
Estradiol	2.00	87773	100699	87.16	3.22

Estradiol at 1000 $\mu\text{g/ml}$ from plasma

TABLE 6: ADHESION TO GASTROINTESTINAL TRACT AFTER TEST FORMULATION

Time (h) following administration	Weight of tissue (g)	Concentration of drug in tissue ($\mu\text{g/g}$)
2	4.01 \pm 0.37	7.74 \pm 0.75
24	3.41 \pm 0.56	1.37 \pm 0.17
48	3.38 \pm 0.41	0.08 \pm 0.01
72	2.93 \pm 0.85	<LLOQ
96	3.16 \pm 0.32	<LLOQ

Adhesion to gastrointestinal tract after administration of 1 ml of the test formulation containing 0.16 mg of norethindrone, N=4N=4

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Conflict of Interest:

The authors declare that they have no conflict of interest.

REFERENCES

- Schindler AE, Campagnoli C, Druckmann R, Huber J, Pasqualini JR, Schweppe KW, Thijssen JH. Classification and pharmacology of progestins. *Maturitas* 2003;46:S7-S16.
- Norethindrone Tablets. [cited 25 March 2017] Available from: <https://www.drugs.com/pro/norethindrone-tablets.html/>
- Santos M, Hendry D, Sangi-Haghpeykar H, Dietrich JE. Retrospective review of norethindrone use in adolescents. *J Pediatr Adolesc Gynecol* 2014;27:41-44.
- Loo JC, Brien R. Analysis of norethindrone in plasma by high-performance liquid chromatography. *J Liq Chromatogr* 1981;4:871-7.
- Pommier F, Sioufi A, Godbillon J. Simultaneous determination of norethisterone and six metabolites in human plasma by capillary gas chromatography with mass-selective detection. *J Chromatogr B* 1995;674:155-165.
- Li W, Li YH, Li AC, Zhou S, Naidong W. Simultaneous determination of norethindrone and ethinyl estradiol in human plasma by high performance liquid chromatography with tandem mass spectrometry--experiences on developing a highly selective method using derivatization reagent for enhancing sensitivity. *J Chromatogr B Analyt Technol* 2005;825:223-32.
- Licea-Perez H, Wang S, Bowen CL, Yang E. A semi-automated 96-well plate method for the simultaneous determination of oral contraceptives concentrations in human plasma using ultra performance liquid chromatography coupled with tandem mass spectrometry. *J Chromatogr B* 2007;852:69-76.
- Gong Z, Chandler K, Webster S, Kerley R, Buist S, McCort-Tipton M. Simple and rapid determination of norethindrone in human plasma by supported liquid extraction and ultra-performance liquid chromatography with tandem mass spectrometry. *Talanta* 2012;91:77-82.
- Altaani BM, Al-Nimry SS, Haddad RH, Abu-Dahab R. Preparation and characterization of an oral norethindrone sustained release/controlled release nanoparticles formulation based on chitosan. *AAPS PharmSciTech* 2019;20:54-67.
- ICH, Q2R1, Harmonized Tripartite Guideline, Validation of analytical procedures: text and methodology. Proceedings of the International Conference on Harmonization, Geneva, March, 1994.
- Kaklamanos G, Theodoridis G, Papadoyannis IN, Dabalís T. Determination of anabolic steroids in muscle tissue by liquid chromatography-tandem mass spectrometry. *J Agric Food Chem* 2007;55(21):8325-30.
- Harahap Y, Devina D, Harmita H. Determination of Ethinyl Estradiol and Levonorgestrel in Human Plasma with Prednisone as Internal Standard Using Ultra-performance Liquid Chromatography-Tandem Mass Spectrometry. *J Pharm Bioallied Sci.* 2019;11:254-261.
- Suresh R, Anarthanan SVJ, Manavalan R, Valliappan K. Aspects of validation in HPLC method development for pharmaceutical analysis - Comparison of validation requirements by FDA, USA and ICH. *Int J Pharm Sci Res* 2010;12:123-33.
- Validation of Compendial Procedures. USP30-NF25, United States Pharmacopoeia/National Formulary. 30th ed. Rockville, MD: Pharmacopoeial Convention; 2008.
- United States Pharmacopoeia/National Formulary. 30th ed. Rockville, MD: Pharmacopoeial Convention; 2008.
- Rao R, Talluri MVK, Raju AN, Shinde DD, Ramanjaneyulu GS. Development of a validated RP-LC/ESI-MS-MS method for separation, identification and determination of related substances of tamsulosin in bulk drugs and formulations. *J Pharm Biomed* 2008;46:94-103.
- Reddy LS, Reddy SLNP, Reddy GS. Development and validation of a stability indicating liquid chromatographic method for simultaneous estimation of Dutasteride and Tamsulosin in combined dosage form. *Orient J Chem* 2014;29:1665-73.
- Tiwari G, Tiwari R. Bioanalytical method validation: an updated review. *Pharm Methods* 2010; 1(1):25-38.
- Food, Drug Administration Centre for Drug Evaluation and Research (FDA). Guidance for Industry-Bioanalytical Method Validation, Center for Drug Evaluation and Research, US Department for Health and Human Services, Silver Spring, MD, 2013 (May 2001).
- Lindholm J. Development and validation of HPLC methods for analytical and preparative purposes. [Dissertation] Uppsala: Acta Universitatis Upsaliensis; 2004. p. 87.