

# Screening of Aqueous and Alcoholic Extracts of Some Indian Medicinal Plants for Antibacterial Activity

JIGNA PAREKH AND SUMITRA CHANDA\*

Phytochemical, Pharmacological and Microbiological Laboratory, Department of Biosciences, Saurashtra University, Rajkot-360 005, India.

The antibacterial activities of 100 extracts of 50 Indian plant species were tested against six medically important bacterial strains, viz., *B. cereus*, *S. aureus*, *S. epidermidis*, *K. pneumoniae*, *A. fecalis* and *P. aeruginosa*. The antibacterial assay was done by both agar disc diffusion method and agar well diffusion method. The antibacterial activity exhibited by alcoholic extract was better than the aqueous extract. The results evaluated as the diameter of the inhibition zone of microbial growth showed that the extracts were more active against gram-positive bacteria than gram-negative bacteria. Amongst the investigated microorganisms, the most resistant bacteria were *P. aeruginosa* and *A. fecalis*. The most susceptible bacteria were gram-positive *B. cereus* and gram-negative *K. pneumoniae*, and the maximum activity was shown by *T. chebula*, *M. indica* and *E. citriodora*.

For centuries plants have been used throughout the world as drugs and remedies for various diseases since they have great potential for producing new drugs of great benefit to mankind. There are many approaches to search for new biologically active principles in higher plants<sup>1</sup>. One such approach is systematic screening, which may result in the discovery of novel effective compounds<sup>2</sup>. Despite the existence of potent antibiotics and antifungal agents, resistant or multi-resistant strains are continuously appearing, imposing the need for a permanent search and development of new drugs<sup>3</sup>. This revival of interest in plant-derived drugs is mainly due to the current widespread belief that "green medicine" is safe and more dependable than the costly synthetic drugs, many of which have adverse side effects.

The purpose of this study was to screen some Indian medicinal plant extracts that could be useful for the development of new tools as antimicrobial agents for the control of infectious diseases.

Fresh plant/plant parts were collected randomly from the semi-arid region of Rajkot, Gujarat, India. The taxonomic identities of these plants were confirmed at the Department of Biosciences, Saurashtra University, Rajkot. The voucher specimen numbers of the plants collected and their medicinal uses are reported in Table 1. Fresh

plant material was washed thoroughly, air dried and then homogenized to fine powder.

Plant material (10 g) was extracted with 100 ml ethanol by subjecting it to agitation on rotary shaker (190-200 rpm) overnight, filtering it with muslin cloth and concentrating it to one-fifth of the volume<sup>4</sup>. Crude aqueous extract was prepared by subjecting plant material (10 g) to slow heat for 6 h and filtered through muslin cloth and concentrated to one-fifth of the total volume<sup>4</sup>.

The microbial strains investigated (Table 2) were obtained from National Chemical Laboratory (NCL), Pune, India. Microorganisms were maintained at 4°C on nutrient agar slants (for bacteria) and MGYE slants (for yeast).

*In vitro* antimicrobial activity was examined for aqueous and alcoholic extracts from 50 different traditionally used medicinal plants. Antimicrobial activity of the aqueous extracts was evaluated by agar disc diffusion method<sup>5</sup> and that of alcoholic extracts was evaluated by agar well diffusion method<sup>6</sup>.

Successful prediction of botanical compounds from plant material is largely dependent on the type of solvent used in the extraction procedure. The traditional healers use water primarily as the solvent, but in our studies we found that the plant extracts in organic solvent (methanol or ethanol) provided more consistent antimicrobial activity as compared to those extracted in water. These

\*For correspondence

E-mail: sumitrachanda@yahoo.com

observations can be rationalized in terms of the polarity of the compounds being extracted by each solvent and, in addition to their intrinsic bioactivity, by their ability to dissolve or diffuse in the different media used in the assay. The growth media also seem to play an important role in the determination of the antibacterial activity.

Table 1 summarizes the data of the plant species selected for the study. Fifty aqueous and alcoholic extracts of the plants belonging to 30 families were tested against three gram-positive and three gram-negative bacteria. The results of antibacterial activity of the screened plants are presented in Table 2.

**TABLE 1: SELECTED INDIAN MEDICINAL PLANTS TESTED FOR ANTIBACTERIAL ACTIVITY**

Plant species (Common name)/ Voucher no.	Part used*	Medicinal uses <sup>9,10</sup>
<i>Achyranthus aspera</i> L. (Aghedo)/ PSN635	W	Cough, rheumatism, bronchitis, piles
<i>Eranthemum</i> spp. L. (Black Croton)/ SU/BIO/486/Thakrar	L	In boils and rashes
<i>Calotropis gigantea</i> (L.) R. Br. (Ankdo)/ PSN449	L	Purgative, antihelminthic
<i>Eucalyptus citriodora</i> Hook. (Nilgiri)/ PSN1058	L	Asthma, laryngitis, sore throat, colds, fevers
<i>Fagonia cretica</i> L. (Dhamasa)/ PSN98	W	Tonic, fevers, asthma, dysentery, bronchitis
<i>Rauwolfia serpentina</i> (L.) Benth ex Kurz. (Sarpghandha )/ SU/BIO/487/Thakrar	L	In bites of poisonous insects and reptiles
<i>Musa paradisiaca</i> L. (Banana)/ PSN719	L	Scabies and inflammations
<i>Ipomea pentphylla</i> Jacq. (Gariya vel)/ PSN522	L	Ornamental
<i>Solanum khasianum</i> C. B. Clarke. (Ubhi ringani)/ SU/BIO/488/Thakrar	W	Antiinflammatory, antiarthritic
<i>Croton bonaplaniun</i> Baill. (Croton)/ PSN673	W	Ornamental
<i>Carissa congesta</i> Wt. (Karamda)/ PSN440	L	Diarrhoea, anorexia, skin diseases, scabies
<i>Tridax procumbens</i> L. (Bhangro)/ PSN414	W	Emetic and purgative
<i>Enicostema hyssopifolium</i> (Willd.) I.C. Verd. (Mamejavo)/ PSN470	W	In fever, rheumatism, sciatica,
<i>Vitex negundo</i> L. (Nagod)/ PSN611	AP	In catarrhal dengue, rheumatism, dyspepsia, diarrhoea
<i>Caesalpinia crista</i> L. (Kakach)/ PSN244	AP	Useful in amenorrhoea, dysmenorrhoea, fevers, cough
<i>Cymbopogon citratus</i> (DC.) Stapf. (LeeliCha)/ PSN808	L	Chronic rheumatism, lumbago, myalgia
<i>Ocimum canum</i> Sims. (Shyam tulsii)/ SU/BIO/243A/Thakrar	W	In skin diseases
<i>Sesamum indicum</i> L. (Tal)/ PSN569	W	In gonorrhoea, dysentery, piles, wounds, ulcers
<i>Ocimum basilicum</i> L. (Takmaria)/ PSN626	W	Cephalgia, colic worms, arthralgia, arthritis
<i>Jasminum malabaricum</i> L. (Jui)/ PSN432	A	Treatment of cataract, haemorrhoea
<i>Thevetia peruviana</i> (Pers.) Merr. (Pila Karen)/ PSN447	L	Cariotonic drug, insecticide
<i>Launaea procumbens</i> (Roxb.) Ram. and Raj. (Bhoy pathri)/ PSN397W	W	Lactagogue, rheumatism
<i>Thuja orientalis</i> L. (Vidya)/ PSN1092	L	Ornamental
<i>Cassia tora</i> L. (Kuvadiao)/ PSN258	W	Obstinate skin diseases, cardiac disorders
<i>Quisqualis indica</i> L. (Madhumalati)/ PSN289	A	Antihelminthic
<i>Lantana camera</i> L. (Abhagani)/ PSN605	W	antispasmodic, vulnerary, diaphoretic, carminative
<i>Tephrosia purpurea</i> (L.) Pers. (Sharpankho)/ PSN223	W	In obstruction of the liver, spleen and kidney, dropsy, boils
<i>Andrographis paniculata</i> (Burm. F.) Wall. (Lilu Karyatu)/ PSN574	L	Diarrhoea, dysentery, dyspepsia, constipation, tumor
<i>Cassia angustifolia</i> Vahl. (Mindhiaval)/ PSN248	L	Chronic constipation, abdominal disorders, leprosy
<i>Embelica ribes</i> Burm. F. (Vavding)/ SU/BIO/489/Thakrar	S	Brain tonic, skin diseases, neuro disorder, constipation
<i>Dalbergia sisoo</i> Roxb. (Moto Shisham)/ PSN179	L	Gonorrhoea, menorrhoea, excoriation, ophthalmopathy
<i>Aloe barbadensis</i> Mill. (Kuamwarpanthu)/ PSN723	L	Eye infection, piles, cough, inflammation
<i>Centella asiatica</i> (L.) Urb. (Brahmi)/ PSN340	L	Skin diseases, insomnia, asthma, bronchitis
<i>Mangifera indica</i> L. (Mango)/ PSN1132	L	Hiccough, hyperdipsia, haemoptysis, haemorrhages,
<i>Xanthium strumarium</i> L. (Gadariyu)/ PSN419	W	Malarial fever, renal complaints, leucorrhoea, menorrhagia,
<i>Ervatamia divaricata</i> (L.) Burkill. (Chandani Tagar)/ PSN446	AP	Ornamental
<i>Vitis vinifera</i> L. (Grapes)/ PSN128	L	Bilious fever, anemia, heart disease, urinary disorder
<i>Picrohiza kurroa</i> L. (Kadu)/ SU/BIO/493/Thakrar	Rh	To treat worms, constipation, low fever, scorpion sting
<i>Cicer erientinum</i> L. (Gingra)/ PSN159	W	Appetizer, tonic, skin diseases, dyspepsia, fever
<i>Spinacia oleracea</i> L. (Palak)/ SU/BIO/491/Thakrar	L	Carminative, laxative, hypoglycaemic, inflammation
<i>Coleus aromaticus</i> Benth. (Ajma pan)/ SU/BIO/492/Thakrar	L	In asthma, chronic cough, strangury, calculas
<i>Allium sativum</i> L. (Lasun)/ SU/BIO/490/Thakrar	W	Eye and heart diseases, inflammation, piles
<i>Daucus carota</i> L. (Carrot)/ PSN 343	AP	Anorexia, dyspepsia, diarrhoea, cough, asthma, ulcer
<i>Lagenaria vulgaris</i> Seringe (Dudhi)/ PSN 328	F	Antibiotic; Antidote; Diuretic; Emetic; Febrifuge
<i>Mucuna pruriens</i> (L.) DC. (Kaucha)/ PSN 209	W	Dyspepsia, neuro diseases, leucorrhoea, colic worms
<i>Glycyrrhiza glabra</i> L. (Jethimadh)/ SU/BIO/494/Thakrar	L	Sore throat, cold, cough, broncial infections, influenza
<i>Bassella rubra</i> L. (Velbondi, Poi)/ PSN654	L	Mild laxative, diuretic, used in urticaea, gonorrhoea
<i>Solanum surattense</i> Burm. F. (Bethi ringani)/ PSN543	W	Kidney disorder
<i>Aristolochia bracteolata</i> Lam. (Kidamari)/ PSN662	S	Constipation, inflammation, boils, syphilis, gonorrhoea
<i>Terminalia chebula</i> Retz. (Himej)/ PSN292	F	Asthama, urinary disorders, piles, ulceration, rheumatism

The ethnobotanical information like vernacular name, voucher no., part used and medicinal use of the selected Indian medicinal plants. \*: W - Whole; L - Leaves; S - Stem; Rh - Rhizome; AP - Aerial parts

TABLE 2: ANTIBACTERIAL ACTIVITY OF AQUEOUS AND ALCOHOLIC EXTRACTS OF INDIAN MEDICINAL PLANTS

Sr. No.	*Inhibition zone (mm)											
	<i>B. cereus</i>		<i>S. aureus</i>		<i>S. epidermidis</i>		<i>K. pneumoniae</i>		<i>A. fecalis</i>		<i>P. aeruginosa</i>	
	A	B	A	B	A	B	A	B	A	B	A	B
1	4	2	2	0	15	0	0	7	0	2	0	0
2	4	1	2	0	11	0	0	2	0	2	0	1
3	0	2	0	0	0	0	0	6	0	2	0	1
4	4	12	0	10	7	15	2	17	0	12	0	9
5	0	3	0	0	4	0	0	7	0	7	0	1
6	0	3	0	0	4	2	2	6	0	2	0	1
7	0	1	0	0	8	0	0	0	0	0	0	1
8	6	4	0	0	5	0	0	2	0	0	0	1
9	0	3	0	0	0	0	0	3	0	0	0	1
10	0	4	0	0	0	2	0	5	0	0	0	1
11	3	10	0	4	0	4	0	6	0	0	0	1
12	0	0	0	0	4	0	0	4	0	0	0	1
13	0	3	0	0	0	0	3	2	2	2	0	1
14	0	12	0	1	0	0	0	4	3	0	0	0
15	0	4	0	0	0	0	0	5	3	0	0	0
16	0	7	0	1	0	0	0	3	1	0	0	0
17	0	6	0	1	0	0	0	9	3	0	0	0
18	0	5	0	1	0	0	0	2	2	0	0	0
19	0	10	0	0	0	0	0	0	0	0	0	0
20	0	3	0	0	0	2	0	1	0	0	0	0
21	0	12	0	0	0	1	0	0	0	0	0	0
22	0	15	0	20	0	12	0	2	0	0	0	0
23	4	2	0	0	0	1	3	3	0	0	0	0
24	0	7	0	0	0	1	4	4	0	0	0	0
25	8	4	2	0	0	1	4	0	0	0	0	0
26	0	5	0	0	0	6	0	0	0	0	0	0
27	0	3	0	0	0	3	0	0	0	0	0	0
28	0	3	0	0	0	4	0	0	0	0	0	0
29	0	6	6	0	3	5	3	0	0	0	0	0
30	2	4	3	0	3	3	2	0	0	0	0	0
31	0	2	4	0	1	2	0	10	0	0	0	0
32	0	1	4	0	0	2	0	2	0	0	0	0
33	0	5	2	4	7	0	0	6	0	0	0	0
34	6	14	2	12	3	9	4	22	0	11	3	7
35	0	7	3	4	2	0	0	9	0	0	0	0
36	0	7	3	0	0	0	0	0	0	0	0	0
37	6	12	3	6	4	3	7	6	3	1	2	2
38	0	10	0	2	0	0	0	5	0	3	0	0
39	0	7	0	1	0	0	0	7	0	2	0	0
40	0	3	0	0	0	0	0	1	0	0	0	0
41	0	3	0	0	0	0	0	6	0	0	0	0
42	0	4	0	0	0	0	0	13	0	0	0	0
43	3	4	0	0	0	0	0	6	0	0	0	0
44	0	6	0	1	0	0	0	7	0	0	0	0
45	0	12	6	7	0	0	0	12	0	0	0	0
46	0	0	0	0	0	0	0	4	0	0	0	0
47	0	0	0	0	0	0	0	5	0	0	0	0
48	0	0	0	0	0	0	0	3	0	0	0	0
49	0	15	0	20	0	0	0	15	0	0	0	0
50	11	27	15	23	8	21	9	20	5	17	5	7

The evaluation of antibacterial activity of aqueous and alcoholic extracts of the selected medicinal plants. The results are mean values of diameter of zone of inhibition in millimetres. A: Aqua extract; B: Alcoholic extract (1-36: ethanol extract, 37-50: methanol extract); 1-50: plant species in order as shown in Table 1; \*: mean values of zone diameter in millimetres (mm) are taken

Out of 100 extracts, 56 showed some degree of activity against one of the tested bacterial strains. The plants which exhibited significant antibacterial activity (defined as a perfectly clear zone with diameter >16 mm) were *Terminalia chebula*, *Aristolochia bracteolata*, *Mangifera indica*, *Launaea procumbens* and *Eucalyptus citriodora*. The gram-positive bacteria were more susceptible than

gram-negative bacteria; the most susceptible gram-positive bacteria were *B. cereus*, while the most susceptible gram-negative bacteria were *K. pneumoniae*. The most resistant bacteria amongst those investigated were *P. aeruginosa* and *A. fecalis*.

The greatest zone of inhibition, with 27 mm diameter,

was displayed by *Terminalia chebula* seed extract against *B. cereus*. In fact, this plant extract (both aqueous and ethanolic) could inhibit all the bacterial strains investigated. Out of 50 aqueous extracts, 16 extracts showed activity against any one of gram-positive bacteria, while only 10 extracts showed activity against any one of gram-negative bacteria. The plant extracts inhibited the gram-positive microorganisms better than gram negative microorganisms. This is in agreement with previous reports that plant extracts are more active against gram-positive bacteria than gram-negative bacteria<sup>7,8</sup>. From our investigation for screening of different plant species, the results obtained confirm the therapeutic potency of some plants used in traditional medicine. In addition, these results form a good basis for selection of plant species candidates for further phytochemical and pharmacological investigation.

## REFERENCES

1. Farnsworth, N.R., and Loub, W.D., In Plants: The Potentials for Extracting Protein, Medicines, and Other Useful Chemicals.

- Workshop Proceedings, Congress Office of Technology Assessment, Washington, D.C, 1983, 178.
2. Janovsk, D., Kubikov, K. and Kokosk, L., **Czech. J. Food Sci.**, 2003, 21, 107.
3. Silver, L.L., **Antimicrob. Agents. Chemother.**, 1993, 37, 377.
4. Parekh, J. and Chanda, S., **Indian J. Pharm.**, 2005, 37, 408.
5. Bauer, A.W., Kirby, W.M.M., Sherris, J.C., and Turck, M., **Amer. J. Clin. Pathol.**, 1966, 45, 493.
6. Perez, C., Paul, M. and Bazerque, P., **Acta. Bio. Med. Exp.**, 1990, 15, 113.
7. Vlietinck, A.J., van Hoof, L., Totte, J., Lasure, A., Vanden Berghe, D., Rwangobo, P.C. and Mvukiyuniwami, J., **J. Ethnopharmacol.**, 1995, 46, 31.
8. Rabe, T. and van Staden, J., **J. Ethonopharmacol.**, 1997, 56, 81.
9. Anjaria, J., Parabia, M. and Dwivedi, S., 1st Edn., Ethnovet Heritage Indian Ethnoveterinary Medicine – An Overview , Pathik Enterprise, Ahmedabad, India, 2002, 1-160.
10. Sriram, S., Patel, M.A., Patel, K.V. and Punjani, N.H., 1st Edn., Compendium on Medicinal Plants, Gujarat Agricultural University, Ahmedabad, India, 2004, 1-116.

Accepted 29 December 2006

Revised 29 March 2006

Received 18 July 2005

Indian J. Pharm. Sci., 2006, 68 (6): 835-838