-Research Paper-

Study on the Mechanism of Zhenwu Decoction in Decreasing the Fats in Non-Alcoholic Fatty Liver Disease Using System Pharmacology and Cytological Validation Test

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Purpose of this study was to screen the active compounds of Zhenwu decoction and their gene targets in the treatment of non-alcoholic fatty liver disease by network pharmacology and cytological validation test. The candidate compounds and related targets of Zhenwu decoction were obtained from traditional Chinese medicine system pharmacology database and PharmMapper. The non-alcoholic fatty liver disease-related genes were obtained from online Mendelian inheritance in man, GeneCards and DisGeNET databases. Molecular docking was carried out to simulate the binding affinities between potential core compounds and key target genes, and the chemical constituents of Zhenwu decoction were analyzed and identified by Ultra-Performance Liquid Chromatography-Quadrupole Time-of-Flight Mass Spectrometry technology. Therapeutic effect and mechanism of Zhenwu decoction on non-alcoholic fatty liver disease were validated using in vitro analysis. The protein-protein interaction network analysis identified the albumin, protein kinase B1, epidermal growth factor receptor, caspase 3 and peroxisome proliferator activated receptor gamma as the key target genes. Gene ontology annotation analysis showed that the Zhenwu decoctionnon-alcoholic fatty liver disease target genes were mainly involved in the response to steroid hormone and lipid catabolic process. The results of the Kyoto encyclopedia of genes and genomes pathway enrichment analysis were mainly related to the Forkhead box O signaling pathway and phosphatidylinositol 3-kinaseprotein kinase B signaling pathway. Paeoniflorgenone had the highest binding affinities for albumin, protein kinase B1, epidermal growth factor receptor, caspase 3 and peroxisome proliferator activated receptor gamma. The *in vitro* experiment showed that 19.5 mg/ml ($p\leq 0.05$) and 39 mg/ml ($p\leq 0.01$) of Zhenwu decoction could reduce lipid accumulation. Real-time quantitative polymerase chain reaction analyses revealed that Zhenwu decoction induced down-regulation of epidermal growth factor receptor messenger ribonucleic acid levels and up-regulation of heat shock protein 90 alpha family class a member 1, mitogen-activated protein kinase 1 and phosphatidylinositol 3-kinase messenger ribonucleic acid levels. Key words: Non-alcoholic fatty liver disease, Zhenwu decoction, network pharmacology, molecular docking,

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Non-Alcoholic Fatty Liver Disease (NAFLD) is defined as the presence of hepatic steatosis determined by imaging or histology, excluding the secondary causes of fat accumulation in the liver^[1]. NAFLD can be divided into the Non-Alcoholic Fatty Liver (NAFL) and Non-Alcoholic Steatohepatitis (NASH)^[1]. NAFLD is usually associated with metabolic complications such as obesity, diabetes and dyslipidemia. It is the major cause of liver diseases worldwide with a significantly increasing burden. At present, the incidence of NAFLD is about 200/10 000 persons per year. It is likely to be the leading cause of End-Stage Liver Disease (ESLD), which affects both adults and children^[2]. Over the past decade, chronic liver diseases, cardiovascular diseases and Type 2 Diabetes Mellitus (T2DM) have been focused on among the NAFLD-related chronic diseases. A recent meta-analysis showed that the mortality rate of NAFLD, which is mainly caused by liver-related and cardiovascular diseases, had increased by 57 %, while the risk of T2DM and NAFLD-related chronic kidney disease had nearly been tripled^[3]. It is crucial to implement novel therapeutic interventions that can prevent or reverse the damaging effects of NAFLD^[4].

The famous Chinese medicine formula, Zhenwu Decoction (ZWD), was first described in the "Treatise on Febrile Diseases" by Zhang Zhongjing. ZWD is composed of Aconiti Lateralis Radix Praeparata (Fuzi in Chinese, lateral radix of *Aconitum carmichaelii* Debx.), *Zingiber officinale* Roscoe (Shengjiang in Chinese, the rhizome of *Zingiber officinale* Rosc.), *Poria cocos* (Schw.) Wolf. (Fuling in Chinese, the sclerotium of *Poria cocos* (Schw.) Wolf.), Paeoniae Radix Alba (Bai Shao in Chinese, a radix of *Paeonia lactiflora* Pall.) and *Atractylodis macrocephalae* Rhizoma (Bazhu in Chinese, a radix of *9*:9:9:6, respectively. ZWD has a clear hypolipidemic effect, lowering serum triglycerides, total cholesterol and raising high density lipoprotein levels^[5].

Clinical studies have proved the better therapeutic potential of ZWD in combination with Sini powder on T2DM overlapping NAFLD (T2DM-NAFLD) as compared to the control group but their mechanism of action is still not clear^[6]. ZWD could regulate energy metabolism, reduce oxidative stress by lowering serum lysophospholipid levels in rats^[7].

Network pharmacology adopts the "multi-compound, multi-target and multi-pathway" method and involves system biology, bioinformatics and pharmacology. It is usually used to study the systematic effects of traditional Chinese medicine and is considered to be a novel strategy for drug discovery^[8] for significantly improving the success rate of drug development, reducing the cost and predicting the side effects of drugs^[9]. This study was the first to elucidate the mechanisms of ZWD in NAFLD treatment with a network pharmacology approach. The flowchart of the study is presented in fig. 1.



Fig. 1: Flowchart of the study design

MATERIALS AND METHODS

Extraction and screening of the active compounds of ZWD:

Chinese names of the five traditional Chinese herbs present in ZWD were used to search the active compounds of each medicine in the Traditional Chinese Medicine Systems Pharmacology Database and Analysis Platform (TCMSP) (http://tcmspw.com/ tcmsp.php). TCMSP contains Chinese medicinal herbs, chemicals, targets and drug-target networks. The active compounds in ZWD were obtained using Oral Bioavailability (OB)≥30 % and Drug-Likeness, (DL)≥0.18 as screening conditions.

Screening of the targets and core compounds of ZWD:

The screened active compounds of ZWD were submitted into the PharmMapper database (http://lilab-ecust.cn/ pharmmapper/index.html) to obtain the corresponding targets using "Norm Fit≥0.90" as screening criteria. The corresponding gene names of the target proteins were obtained from through UniProt database (https:// www.uniprot.org/). Duplicate gene names were then removed. The compound-target network of ZWD was constructed and visualized using Cytoscape 3.8.2, and the core compounds of ZWD were determined.

Collection of known therapeutic targets acting on the NAFLD and candidate genes:

The therapeutic target genes of NAFLD were obtained from three databases, including GeneCards (https:// www.genecards.org/), Online Mendelian Inheritance in Man (OMIM) (http://www.omim.org/) and DisGeNET (https://www.disgenet.org/). The search term "nonalcoholic fatty liver" was used in all three databases and the species were limited to "*Homo sapiens*". The therapeutic targets were collected and duplicate targets were removed. NAFLD-related protein targets were imported into the UniProt database. Then, the obtained genes were submitted to Venny 2.1 (https://bioinfogp. cnb.csic.es/tools/venny/) to draw a Venn diagram. The intersected genes in the two datasets were selected as candidate genes.

Construction of the Protein-Protein Interaction (PPI) network:

Search Tool for the Retrieval of Interacting Genes/ Proteins (STRING) database (https://string-db.org/) was used to study the association of the retrieved active ingredient targets of ZWD and NAFLD target genes. The confidence score of relevance was set to ≥ 0.4 as the critical value to obtain the PPI network. The PPI network was visualized using Cytoscape 3.8.2 and the key genes were identified by degree, betweenness and closeness.

Enrichment analyses of Gene Ontology (GO) annotation and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway:

The gene targets of ZWD-NAFLD were subjected to GO annotation and KEGG pathway enrichment analysis using Metascape (https://metascape.org/). The target species was set to *Homo sapiens*. The results were presented using an R software package.

Construction of the compound-target-pathway network:

The active compound of ZWD, selected gene targets and related pathways were submitted to Cytoscape 3.8.2 in order to build a compound-target-pathway network. The graphic network represented the connections and interactions among the target genes, proteins or molecules and pathways. The value of each node represented the number of connections between the node and other nodes and the greater the degree, the more important the node.

Molecular docking between the main active components of ZWD and core proteins:

In order to further verify the reliability of this study, the top five targets in the PPI network and the top 10 core compounds of ZWD were selected for molecular docking. First, the Spatial Data File (SDF) files of the core compounds of ZWD were downloaded from the PubChem database (https://www.uniprot.org/) and then converted to mol2 format files using Open Babel 2.4.1 software. The best three Dimensional (3D) protein structures of the main protein targets were obtained from Protein Data Bank (PDB) and saved in PDB format. Then, the downloaded protein structures and active compounds of ZWD were opened in PyMOL to remove the water molecules and ligands. The active compounds and protein targets were then imported to AutoDock software for docking. The smaller the value of docking binding force, the more stable the compound binds to the protein and the more likely it is to interact. Finally, the PDB file was imported to LigPlot software to find the interaction between small molecules and large proteins.

Ultra-Performance Liquid Chromatography-Quadrupole Time-of-Flight Mass Spectrometry (UPLC-QTOF/MS) analysis of ZWD:

Standard preparation: 0.3 ml, 0.6 ml, 1.2 ml of ZWD were diluted into 1.2 ml, 0.9 ml, 0.3 ml of 50 % methanol solution, ultrasonic treatment for 30 min and then filtered with 0.22 µm microporous membrane. UPLC analysis conditions were as follows. The results were performed on a liquid chromatography-mass spectrometer (SNAPT XS) equipped with ACQUITY UPLC CSH C₁₈ (100 mm×2.1 mm, 1.7 μ m) at 40°. The mobile phase consisted of 0.1 % formic acid aqueous solution (A) and acetonitrile (B). The optimized gradient program was 0-5 min, 95 %-90 % A; 5-8 min, 85 % A; 8-15 min, 85 %-83 % A; 15-18 min, 83 %-80 % A; 18-22 min, 80 %-70 % A; 22-28 min, 70 %-5 % A; 28-31 min, 5 %-5 % A; 31-35 min, 5 %-95 % A. The flow rate was 0.2 ml/min and the injection volume was 2 µl.

MS analysis conditions were as follows. MS analysis was performed on a liquid chromatographymass spectrometer (SNAPT XS) equipped with an Electrospray Ionization (ESI) source. The capillary voltage is set to 2000 V, the source temperature is 200° , the desolvation temperature is 450° , the cone gas flow is 50 l/h and the desolvation gas flow is 600 l/h. The sample infusion flow rate was 5 µ/min. The full scan quality range of the scan is m/z 50-1200. Data is obtained in continuous mode.

Preparation of ZWD:

The dried raw herbs of ZWD were purchased from the outpatient department of Chengdu University of traditional Chinese medicine (Chengdu, China). Zingiber officinale Roscoe was homemade (fresh Chengdu-origin ginger was washed and sliced). Aconiti Lateralis Radix Praeparata, Paeoniae radix Alba, Poria cocos (Schw.) Wolf., Zingiber officinale Roscoe and Atractylodes macrocephala Koidz. (Production batch number: 21090104, 21081906, 21101061, C038210701, respectively) were mixed at the proportion of 9:9:9:9:6, that is 42 g soaked in water (500 ml) for 30 min and then boiled for 30 min. The residues were filtered and concentrated to 0.78 g/ml and after centrifugation, the supernatant was collected and five concentration gradients (9.75 mg/ml, 19.5 mg/ml, 39 mg/ml, 78 mg/ ml and 156 mg/ml) were prepared, which were then stored at 4°.

Cell cultures and treatments:

Liver hepatocellular carcinoma cell line (HepG2) cells were cultured in 90 % high glucose sugar Dulbecco's Modified Eagle Medium (DMEM, Gibco), containing 10 % Fetal Bovine Serum (FBS, Gibco), 1 % penicillin and streptomycin. The cells were maintained at 37° under 5 % Carbon dioxide (CO₂). Free Fatty Acids (FFA) are a mixture of oleic acid and palmitic acid at 2:1. The cells were treated with different concentrations of FFAs and cell viability was observed.

The HepG2 cell viability was determined using Cell Counting Kit-8 (CCK-8). For this purpose, HepG2 cells in the logarithmic phase $(2 \times 10^3$ cells per well) were inoculated into 96-well plates (100 µl/well) and treated with the different concentrations of ZWD (9.75 mg/ml, 19.5 mg/ml, 39 mg/ml, 78 mg/ml, 156 mg/ml) for 24 h. Then, the bottom solution was removed and 10 µl of CCK-8 solution was added to 100 µl complete medium. The CCK-8 solution was added to each well and the plate was incubated at 37° for 2 h. The absorbance was measured at 450 nm using a microplate reader.

Oil Red O (ORO) staining:

The cells were washed with Phosphate Buffered Saline (PBS) solution and fixed with 4 % paraformaldehyde for 30 min. The ORO storage solution (0.5 %) was prepared with 0.25 g oil red dry powder and 50 ml isopropanol and filtered through a 0.22 μ m filter. The ORO working solution was prepared by mixing the ORO storage solution and deionized water at a ratio of 3:2. The fixed HepG2 cells were stained with an ORO working solution for 30 min at 37° and then immediately washed with PBS for 3 times. The qualitative analysis of lipids and quantification of lipid accumulation under the microscope were based on the Optical Density (OD) value of the target ORO.

Ribonucleic Acid (RNA) isolation and Real-Time quantitative Polymerase Chain Reaction (RTqPCR):

RT-qPCR was performed to measure the hepatic cell expression of the predicted NAFLD-related genes. Total RNA was collected with the RNA isolation kit following the instructions, complementary Deoxyribonucleic Acid (cDNA) synthesis kit with genomic DNA (gDNA) eraser was used to carry out the reverse transcription reactions. RT-qPCR was performed to measure the relative expression of messenger RNA (mRNA), using SYBR Green RT-PCR master mix. The reaction conditions were 95°, 3 min; 95°, 10 s; 55°, 20 s and 72°, 30 s, for 40 cycles. Beta-actin was used as a control and the $2^{-\Delta\Delta Ct}$ method was conducted for the data analysis. The primer sequence of target genes was synthesized by Invitrogen Biotech Co., Ltd (Table 1). The targets Epidermal Growth Factor Receptor (EGFR), Peroxisome Proliferator Activated Receptor Gamma (PPARG), Heat Shock Protein 90 Alpha Family Class A Member 1 (HSP90AA1) and Mitogen-Activated Protein Kinase 1 (MAPK1) formed the PPI network analysis and Phosphatidylinositol 3-Kinase (PI3K) formed the PI3K-Protein Kinase B (AKT) signaling pathway.

Statistical analysis:

All the data were expressed as mean \pm Standard Error (SE). One-way Analysis of Variance (ANOVA) in GraphPad Prism v8.3.0 was used to test and analyze the data. The difference with p<0.05 was considered statistically significant.

RESULTS AND DISCUSSION

According to the screening criteria of OB \geq 30 % and DL \geq 0.18, a total of 61 active compounds were obtained for the five medicinal herbs of ZWD. After removing the duplicate data, a total of 59 compounds were obtained of which 20 were from Fuzi, 12 from Bai Shao, 7 from Baizhu, 15 from Fuling and 5 from Sheng Jiang. After removing the duplicate data and screening the target genes using PharmMapper and through the standardized transformation of Uniprot, a total of 133 target genes were obtained.

The compound-target network was constructed to show the complex compounds and multiple target genes and the correlations between them. There were 303 nodes and 824 edges in the compound-target network of ZWD. It was suggested that the compounds of ZWD might play a synergistic effect on these target genes and might have pharmacological effects on NAFLD and other diseases. The highest connectivity was that of BSH5 with 55 target genes with an OB value of 53.87 %; this might be considered as the potential key active compound as shown in fig. 2.

Gene name	Amplicon length	Sequences
EGFR-F	97	5'-ACCCATATGTACCATCGATGTC-3'
EGFR-R	97	5'-GAATTCGATGATCAACTCACGG-3'
PPARG-F	120	5'-AGATCATTTACACAATGCTGGC-3'
PPARG-R	120	5'-TAAAGTCACCAAAAGGCTTTCG-3'
HSP90AA1-F	191	5'-CCAGTTCGGTGTTGGTTTTTAT-3'
HSP90AA1-R	191	5'-CAGTTTGGTCTTCTTTCAGGTG-3'
MAPK1-F	188	5'-ATGGTGTGCTCTGCTTATGATA-3'
MAPK1-R	188	5'-TCTTTCATTTGCTCGATGGTTG-3'
PI3K-F	180	5'-GAGATTGCAAGCAGTGATAGTG-3'
PI3K-R	180	5'-TAATTTTGGCAGTGATTGTGGG-3'
Beta-actin-F	285	5'-AGCGAGCATCCCCCAAAGTT-3'
Beta-actin-R	285	5'-GGGCACGAAGGCTCATCATT-3'

TABLE 1: THE NUCLEOTIDE SEQUENCES OF THE PRIMER PAIRS USED FOR QUANTITATIVE GENE EXPRESSION

Note: F: Forward primer and R: Reverse primer



Fig. 2: Compound-target network Note: The pink square represents the herbs in ZWD, the circle represents the active compounds of each herb, the A1 and B1 circles represent the active compounds common in different herbs and the blue diamond shape represents the related goals

As shown by the compound-target network, some compounds play a crucial role in the network, thereby proving to be a great potential in the treatment of NAFLD, such as paeoniflorin, dihydrocapsaicin and poricoic acid B. Zhang found that paeoniflorin could prevent NAFLD by restoring the serum Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), Serum Total Cholesterol (TC), Triglyceride (TG), High-Density Lipoprotein (HDL) and Low-Density Lipoprotein (LDL) levels. Paeoniflorin could alleviate the infiltration of liver fats induced by a high-fat diet through reducing steatosis, inflammation, balloon degeneration and necrosis. Suggested mechanisms may relate to the protection on cardiovascular system as body weight and hyperlipidemia reduced, inflammation blocked and lipid deposition inhibited^[8]. In addition, paeoniflorin could also significantly reduce serum insulin and glucagon levels, enhance insulin sensitivity, restore serum lipid spectrum and alleviate liver steatosis^[9]. The dietary dihydrocapsaicin could also reduce blood lipid levels and improve cholesterol metabolism in hyperlipidemia rats^[10]. It is worth

mentioning that the three compounds in *Poria cocos* Wolf, including poricoic acid, pachymic acid and ergosterol, could significantly reduce the TG level in HepG2 cells treated with FFAs^[11].

A total of 3181 NAFLD-related target genes were obtained from OMIM, GeneCards and DisGeNET databases. After removing the duplicate data, a total of 2608 target genes were obtained as shown in Table 2. The intersection genes among the NAFLD-related and ZWD target genes were extracted, which were 67 target genes and were considered as a candidate gene. The intersection genes are shown in fig. 3.

A ZWD-NAFLD target PPI network was obtained, which contained 64 nodes and 718 edges (fig. 4). The nodes represented proteins and edges represented protein interactions. The average node degree value was 10.7 and the average local clustering coefficient was 0.627. In the PPI network, some nodes had a high degree, such as Albumin (ALB), Protein Kinase B1 (AKT1), EGFR, Caspase 3 (CASP3) and PPARG.

TABLE 2: THE BINDING ENERGY VALUES OF CORE COMPOUNDS OF ZWD AND CORE TARGETS

Source	Company	Decree	Binding affinity/KJ/mol								
Source	Compound	Degree -	ALB	AKT1	EGFR	CASP3	PPARG				
BSH5	Paeoniflorin	56	-1.74	-1.63	-1.24	-2.36	-1.35				
BSH2	Paeoniflorgenone	30	-4.42	-3.8	-4.55	-3.83	-3.67				
BZ3	14-acetyl-12-senecioyl- 2E,8Z,10E-atractylentriol	28	-1.74	-2.31	-2.82	-2.05	-1.55				
SHJ5	Dihydrocapsaicin	22	-2.42	-2.26	-2.45	-3.22	-2.43				
FZ17	Jesaconitine	20	0.88	0.26	-0.02	-1.64	0.74				
BZ2	14-acetyl-12-senecioyl- 2E,8E,10E-atractylentriol	20	-3.34	-1.83	-2.41	-2.59	-2.56				
FUL12	Poricoic acid B	17	-1.56	-1.93	-1.84	-2.08	-1.77				
FUL11	Poricoic acid A	17	-1.93	-1.35	-1.98	-2.14	-1.75				
FUL3	7,9(11)-dehydropachymic acid	16	-2.5	-2.42	-3.03	-3.45	-2.46				
SHJ2	6-methylgingediacetate 2	15	-0.36	-0.51	-1.82	-1.78	-1.04				



Fig. 3: Venn diagram of intersection genes among the ZWD target and NAFLD-related genes. There were 70 targets for ZWD and 2541 targets for disease and 67 intersection genes



Fig. 4: PPI network for the intersection genes of ZWD target and NAFLD-related genes. Note: Each node represents a protein target and each edge represents the interaction between two nodes. The PPI network diagram was arranged according to the degree of freedom (df). The more important the node is, the closer it is to the center

These target nodes were identified as key genes and might be potential therapeutic targets for the treatment of NAFLD by ZWD. Alkaline phosphatase, ALT, AST, ALB and total serum protein were significantly correlated with the severity of NAFLD^[12]. Besides, there were significant differences in the ALB-binding function between healthy subjects and different severity groups of NAFLD^[13]. Differential regulation of AKT1 and AKT2 is consistent with upregulation of Forkhead Box Protein O1 (FOXO1), may justify the paradoxical state of insulin resistance relative to the glucoregulatory pathway and augmented insulin sensitivity of the liporegulatory pathway, typical of steatosis and the metabolic syndrome in obese patients^[14]. Liang et al. demonstrated that the EGFR was phosphorylated in the liver tissues of the highfat diet murine model of NAFLD. The inhibition of EGFR could prevent diet-induced lipid accumulation, oxidative stress, hepatic stellate cell activation and matrix deposition^[15]. Hepatocyte death is an important feature of NASH. The CASP3 activation in hepatocytes played an important role in the NASH-related apoptosis and fibrosis^[16]. Finally, a study reported PPARG was positively correlated with liver TG levels in male mice^[17]. Therefore, ALB, AKT1, EGFR, CASP3 and PPARG might be the potential therapeutic targets for the treatment of NAFLD.

The GO annotations of the candidate genes were carried out in three categories, including Biological Process (BP), Molecular Function (MF) and Cell Composition (CC). The top 20 significantly-enriched terms in BP, MF and CC categories (adjusted p<0.01) are presented in fig. 5A-fig. 5C, respectively. A total of 793 entries were displayed in the BP enrichment analysis, among which, many BPs such as response to nutrient levels, response to a steroid hormone, cellular response to lipids and lipid catabolic process, were closely associated with the pathogenesis of NAFLD. The CC enrichment results showed 45 terms, including vesicle lumen, secretory granule lumen, ficolin-1-rich granule, lytic vacuole and pigment granule. A total of 67 enrichment processes were identified related to MF, including nuclear receptor activity, steroid binding, steroid hormone receptor activity, DNA-binding, transcription factor binding and beta-catenin binding. In order to further investigate the biological process of these targets, the KEGG pathways enrichment analysis was conducted, which resulted in a total of 202 entries (p<0.01), including FOXO signal and PI3K-AKT signaling pathway, endocrine resistance and estrogen signaling pathway.

The pathogenesis of NAFLD is closely related to insulin resistance^[18]. A core regulatory mechanism in the FOXO signaling pathway is the phosphorylation of AKT which is the downstream of the PI3K for insulin response (fig. 6). A study showed that the FOXO signal pathway could lead to lipid accumulation and the denaturation of hepatocytes and promote the occurrence and development of NAFLD^[19]. PI3K-AKT signal transduction pathway in hepatocytes is a common molecular mechanism, which is involved in metabolic dysfunction, including obesity, metabolic syndrome and NAFLD^[20]. Cai *et al.* found that the insulin-PI3K/AKT-p70 S6 Kinase (p70S6K) pathway played an important role in the early activation of hepatic stellate

cells^[21]. Interestingly, a study reported that paeoniflorin could have positive influence on improving NAFLD and obesity by regulating the PI3K/AKT pathway^[22]. The results of our compound-target-pathway suggested that paeoniflorin could interact with the PI3K-AKT signaling pathway through AKT1, EGFR, Glycogen Synthase Kinase-3 Beta (GSK3B), HSP90AA1, Mouse Double Minute 2 homolog (MDM2) and Insulin-like Growth Factor 1 Receptor (IGF1R), while paeoniflorgenone could interact with this pathway through MAPK1 and Retinoid X Receptor Alpha (RXRA). In addition, there are also numerous endocrine-related pathways, such as endocrine resistance, estrogen signaling pathway and thyroid hormone signaling pathway. Zhu et al. found that the estrogen signaling pathway could inhibit fatty acid oxidation and TGs in the liver and increase peripheral fat mobilization^[23].

Cytoscape v3.8.2 software was used to construct a compound-target-pathway network, which is shown in fig. 7. There were 328 edges and 91 nodes in the network, which showed that ZWD could treat NAFLD through its multiple compounds and multiple target genes. Among them, paeoniflorin, paeoniflorgenone, 14-acetyl-12-senecioyl-2E, 8Z, 10E-atractylentriol, dihydrocapsaicin, etc., might play important roles in

the efficacy of ZWD for the treatment of NAFLD. The specific information of the compounds is listed in Table 1.

The top 10 active compounds (paeoniflorin, 14-acetyl-12-senecioyl-2E, paeoniflorgenone, 8Z, 10E-atractylentriol, dihydrocapsaicin, jesaconitine, 14-acetyl-12-senecioyl-2E, 8E, 10E-atractylentriol, poricoic acid В, poricoic acid А, 7, (11)-dehydropachymic acid and 6-methylgingediacetate 2) obtained from the compound-target network analysis were docked with the five potential core target genes (ALB, AKT1, EGFR, CASP3 and PPARG) obtained from the PPI network analysis. The details of these compounds and target genes are listed in Table 2 and presented in fig. 8. The binding energies of all the compounds except jesaconitine were less than 0, showing a good binding affinity of these compounds with the top five target genes. Among them, the binding energy of paeoniflorgenone with ALB, AKT1, CASP3 and PPARG proteins was lower than that of the other compounds and the paeoniflorgenone-EGFR showed the lowest binding energy (-4.55); ASN-842 was the active site, showing hydrogen bond interactions (fig. 9). Docking prediction provided a preliminary basis for the further study of these drug targets.



Fig. 5: Top 20 most important entries of (A) BP; (B) MF; (C) CC in GO annotations and 21 important entries of (D) KEGG pathway enrichment analysis



Fig. 6: FOXO signaling pathway

Note: The green circle indicates that this gene is ZWD-NAFLD target gene and p stands for phosphorylation



Fig. 7: Compound-target-pathway network

Note: Pink squares represent the active compounds in ZWD, blue squares represent the target genes and green squares represent the signal pathway of NAFLD







Fig. 9: Molecular docking of paeoniflorgenone with EGFR, (A) Overall diagram; (B) Magnified diagram, showing detailed docking, the dashed yellow lines represent hydrogen bonds and (C) Plane drawing

The docking of the compounds with target proteins and their binding free energy, which were less than 1, suggested that the 10 major core compounds had a good binding affinity for the five core target proteins. The binding energy of paeoniflorgenone was the lowest among the 10 compounds, indicating its good binding affinity with all the five target proteins. The paeoniflorgenone-EGFR showed the lowest binding energy (-4.55) and ASN-842 was the active site for hydrogen bonding.

In order to verify whether the 10 components predicted by molecular docking exist in ZWD, UPLC-Q-TOF-MS was performed to analyze the chemical constituents of ZWD. Full scanning was carried out in positive and negative ion mode and the analysis showed that the chromatogram of representative Base Peak Intensity (BPI) in positive and negative ion mode was as shown in fig. 10. A total of 67 compounds were detected by UniFi software (Table 3). Among them, paeoniflorin, paeoniflorgenone and poricoic acid B were proved to exist in ZWD. Therefore, it can be speculated that paeoniflorin, paeoniflorin and poricoic acid B may play a major role in ZWD in the treatment of NAFLD.

Referring to the relevant literature^[24] and the experimental results in this study, 1 mM FFA was selected as model concentration. As shown in fig. 11, the results of CCK-8 assay showed that ZWD at concentrations ranging from 9.75-156 mg/ml+1 mM FFA displayed no cytotoxicity on cells.

Treatment with FFA can induce significant lipid accumulation in cells. After the cells were treated with the different doses of ZWD for 24 h, the ORO staining showed a significant difference between the experimental group and model group, there were obvious differences when the cells were treated with 19.5 mg/ml and 39 mg/ml of ZWD. The TG contents in HepG2 cells in the experimental group were lower than those in the control group, as shown in fig. 12. These results suggested that 19.5 mg/ml (p \leq 0.05) and 39 mg/ml (p \leq 0.01) of ZWD could reduce lipid accumulation in a concentration-dependent manner.



(B)

Fig. 10: Representative base peak chromatogram of ZWD in positive and negative ionization mode, respectively, (A) Negative ionization mode and (B) positive ionization mode

TABLE 3: IDENTIFICATION OF THE COMPOUNDS OF ZWD BY UPLC-Q-TOF/MS ANALYSIS

Component name	Neutral mass (Da)	Observed neutral mass (Da)	Observed m/z	Mass error (mDa)	Mass error (ppm)	Observed RT (min)	Detector counts	Response	Adducts	Total fragments found
Furfuryl alcohol	98.03678	98.0374	143.0356	0.6	4.4	0.43	430	430	HCOO⁺	1
Epinephrine	416.15299	416.1552	455.1184	2.2	4.9	0.43	1149	1149	K⁺	40
16-Oxoacetylpyrochloric acid	570.39204	570.3945	609.3576	2.4	4	0.45	388	388	K⁺	42
Paradisamide	421.28282	421.2832	444.2724	0.3	0.8	0.46	1283	400	Na⁺	50
Coryneine	195.12593	195.1268	196.1341	0.9	4.4	0.46	619	619	H⁺	12
1-Galloyl-B-D-glucose	332.07435	332.0751	331.0678	0.7	2.2	0.62	8711	7759	H.	19
Gallic acid	170.02152	170.022	169.0148	0.5	3	0.74	42699	40055	H.	2
5-Hydroxymethyl-2- furaldehyde	126.03169	126.0322	125.025	0.5	4.4	0.74	6362	6017	Η [.]	3
1-O-B-D- Glucopyranosylpaeonisuffrone	360.14203	360.1432	405.1414	1.1	2.8	0.81	17152	8794	HCOO⁺, H [.]	12
Carmicheline	377.25661	377.2566	378.2639	0	0.1	1.24	395	395	H⁺	0

Senbusine B	423.26209	423.2619	424.2692	-0.2	-0.4	1.91	772	772	H⁺	1
Atractylenolide II	232.14633	232.147	233.1542	0.6	2.7	2.01	2012	1710	H⁺	5
d-Catechin	290.07904	290.08	289.0727	0.9	3.2	2.24	26247	22781	H.	8
Nonyn	357.23039	357.231	358.2383	0.7	1.8	2.25	20376	16960	H⁺	1
Benzoic acid	122.03678	122.0372	123.0445	0.4	3.6	2.26	496	496	H⁺	1
Dictysine	347.24604	347.247	348.2542	0.9	2.6	2.27	433	433	H⁺	2
Oxypaeoniflorin	496.15808	496.1591	495.1518	1	2.1	2.28	44480	35623	H⁻, HCOO⁺	21
Paeoniflorin	480.16316	480.1649	481.1721	1.7	3.5	3.04	2749	2186	H⁺	10
Fuziline	453.27265	453.2739	454.2812	1.3	2.8	3.08	1526	1526	H⁺	3
Nyonin	437.27774	437.2789	438.2862	1.2	2.7	3.18	33673	27072	H⁺	6
Kaempferol-3,7-di-O-B-D- glucoside	610.15338	610.1551	609.1478	1.7	2.8	3.21	326	326	H-	1
B-Eudesmol	222.19837	222.1978	245.187	-0.6	-2.4	3.27	697	697	Na⁺	0
6-О-В-D- Glucopyranosyllactinolide	362.15768	362.1584	361.1511	0.7	2	3.31	1142	1142	H.	14
(Z)-(1S,5R)-B-Pinen-10-yl-B- vicanoside	446.2152	446.2168	491.215	1.6	3.3	3.62	563	563	HCOO⁺	1
Denudatine	343.25113	343.2524	344.2597	1.3	3.7	3.72	4188	3440	H⁺	4
Ethyl salicylate	166.06299	166.0637	165.0565	0.7	4.4	3.93	2153	2153	H.	14
Poricoic acid B	196.07356	196.0744	197.0817	0.8	4.2	3.94	9904	9086	H⁺	18
Benzyl alcohol	108.05751	108.058	109.0652	0.5	4.2	3.94	2463	2463	H⁺	4
Delbruline	479.2883	479.2903	480.2976	2	4.2	4.42	373	373	H⁺	3
Paeoniflorigenone	318.11034	318.1115	319.1188	1.2	3.7	5.3	1695	1695	H⁺	2
Lactiflorin	462.1526	462.1546	507.1528	2	3.9	6.27	1006	785	HCOO⁺	0
Paeonilactone C	318.11034	318.1119	319.1192	1.6	4.9	7.86	2726	2355	H⁺	7
Hokbusine A	603.30435	603.3056	604.3128	1.2	2	10.17	48550	36036	H+	5
Benzoylaconitine	603.30435	603.3057	648.3039	1.3	2.1	10.18	5395	3794	HCOO⁺	2
Benzoylhypaconine	573.29378	573.2947	574.302	0.9	1.6	11.42	72877	54681	H⁺	5
Atractylenolide IV	306.14672	306.148	305.1407	1.2	4	12.27	277	277	H.	7
Beiwutine	647.29418	647.2967	648.304	2.5	3.9	12.65	1232	838	H⁺	1
Benzoyloxypaeoniflorin	600.18429	600.1854	599.1781	1.1	1.8	13.53	602	602	H.	25
3-Deoxynivalenol	629.32	629.3219	628.3146	1.9	3	13.84	417	417	H.	5
Aconitine	645.31491	645.3173	646.3246	2.4	3.7	14.02	538	538	H⁺	4
Benzoylpaeoniflorin	584.18938	584.1922	585.1995	2.8	4.8	14.48	464	464	H⁺	2
Palbinone	358.21441	358.215	357.2077	0.6	1.7	14.49	319	319	H.	1
Poricoic acid A	512.35017	512.3483	551.3114	-1.9	-3.5	14.77	856	856	K⁺	3
Delbruine	465.27265	465.2744	466.2816	1.7	3.7	14.8	1321	901	H⁺	3

Paeonilactone 168.1150 168.1150 169.1229 0.6 3.5 14.87 1947 1947 H* 5 6-Gingerot 294.18311 294.184 317.1732 0.9 2.8 14.99 5834 5014 Na* 38 Hypaconitine 615.3045 615.3066 638.2958 2.3 3.6 15 393 393 Na* 14 Poricoic acid D 514.3294 514.3294 513.3229 0.7 1.3 15.06 905 642 H 2 25-hydroxy-3r epidehydrotumulosic acid 803.3017 500.3498 493.3425 0.4 1.6.8 15.23 1282 212 H 0 Hydroxy-atractylenolide 248.1412 248.1425 247.1352 1.2 5 15.26 222 222 H 0 36.160-Dhydroxy-atractylenolide 248.1425 247.1352 1.2 5 15.26 221 222 H 0 36.160-Dhydroxy-atractylenolide 248.1424				www.ijps	Unime.cc	////					
Hypac615.30435615.3066638.29582.33.615393393Na'14Poricoic acid D514.3294514.3301513.32290.71.315.06905642H225-Hydroxy-3- epidehydrotunulosic acid500.3507500.3498499.34250.40.815.231282955H3Poricoic acid B484.31887484.3196483.31230.71.415.25217217H0Hydroxy-atractylenolide248.1412248.1425247.13521.2515.26222222H038.160-Dihydroxylauroster323.1463232.1473233.15450.9416.0370816265H438,160-Dihydroxylauroster472.3552472.356471.34870.81.616.2217217H3Poricoic acid F498.33452488.3464483.34911.12.316.39452452H3Dehydrotumulosic acid486.3701486.3718485.34650.91.816.48386386H336-Hydroxy-11c_12c- epoxolean-28-138-olide486.3462485.32962.44.816.61338338H236-Hydroxy-11c_12c- epoxolean-28-138-olide204.1878205.19580.73.61.742703648H4Poricoic acid C482.3491481.33280.73.617.67353353H <td>Paeonilactone</td> <td>168.11503</td> <td>168.1156</td> <td>169.1229</td> <td>0.6</td> <td>3.5</td> <td>14.87</td> <td>1947</td> <td>1947</td> <td>H⁺</td> <td>5</td>	Paeonilactone	168.11503	168.1156	169.1229	0.6	3.5	14.87	1947	1947	H⁺	5
Poricoic cacid D514.3294514.3301513.32290.71.315.06905642H22F-Hydroxy-3- epidehydrotumulosic acid500.35017500.3498499.34250.40.815.231282955H3Poricoic acid B484.31887484.3190483.31230.71.415.25217217H0Hydroxy-atractylenolide248.1412248.1425247.13521.2515.26222222H038.160-Dihydroxylauroster- 8,24-dien-21-oic acid472.3552472.356471.34870.81.616.2217217H3Poricoic acid F498.33452498.3365497.32932416.39452452H3Poricoic acid F498.33452498.3365497.32932416.48366366H3Poricoic acid F498.33452498.3365497.32932416.33452452H3Poricoic acid F498.33452486.3718485.34911.12.316.39452452H3Poricoic acid F486.3709486.3718485.34911.12.316.61338386H3Poricoic acid C486.3704486.3709237.113870.83.416.61338338H'2Sal-Hydroxy-110_12a- epoxyolean-28-138-olide276.1724276.1725277.18050.72.516	6-Gingerol	294.18311	294.184	317.1732	0.9	2.8	14.99	5834	5014	Na⁺	38
25-Hydroxy-3- epidehydrotumulosic acid500.3501500.3498499.3425-0.40.815.231282955H3Poricoic acid B484.31887484.3196483.31230.71.415.25217217H0Hydroxy-atractylenolide248.14124248.1425247.13521.2515.26222222H0B6-hydroxy-atractylon232.1463232.1473233.15450.9416.0370816265H483,160-Dihydroxylauroster- 8,24-dien-21-0ic acid472.3552471.34670.81.616.2217217H3Poricoic acid F498.33452498.3365497.32932416.48309309H43-Epidehydrotumulosic acid484.3552844.3564483.34911.12.316.39452452H3Dehydrotumulosic acid486.3709486.3718485.36450.91.816.48386386H33-Ehydroxy-111,120- epoxylean-28-138-olide486.3345231.13870.83.416.53438438H23-Bridehydroxy-112,120- epoxylean-28-138-olide486.3452276.1732277.18050.72.516.742703648H4Arcetylenolide A203.1364481.33280.40.916.77353353H11B-Red myrcene204.1878204.1885205.19580.7 <td< td=""><td>Hypaconitine</td><td>615.30435</td><td>615.3066</td><td>638.2958</td><td>2.3</td><td>3.6</td><td>15</td><td>393</td><td>393</td><td>Na⁺</td><td>14</td></td<>	Hypaconitine	615.30435	615.3066	638.2958	2.3	3.6	15	393	393	Na⁺	14
epidehydrotumulosic acid500.3307500.3498499.3425-0.4-0.815.231282953H3Poricoic acid B484.31887484.3196483.31230.71.415.25217217H0Hydroxy-atractylenolide248.14124248.1425247.13521.2515.26222222H038-hydroxy-atractylon232.14633232.1473233.15450.9416.0370816265H'438,16d-Dihydroxylauroster- 8,24-dien-21-oic acid472.35526472.356471.34870.81.616.2217217H'3Poricoic acid F498.33452498.3365497.32932416.24309309H'43-Epidehydrotumulosic acid484.35526484.3564483.34911.12.316.39452452H'3Dehydrotumulosic acid486.37091486.3718485.34550.91.816.48386386H'3Atractylenolide A230.1306230.1315231.13870.83.416.53438438H'2Ginger brain276.17254276.1732277.18050.72.516.742703648H'4Poricoic acid C482.33961482.3401481.33280.40.916.77353353H'1B-aqua-anisidine136.1252136.1258137.13310.64.418.181	Poricoic acid D	514.32944	514.3301	513.3229	0.7	1.3	15.06	905	642	H.	2
Hydroxy-atractylenolide248.14124248.1425247.13521.2515.26222222H03B-hydroxy-atractylon232.1433232.1473233.15450.9416.0370816265H'43B-hydroxy-atractylon472.35526472.355471.34870.81.616.2217217H'3Poricoic acid F498.33452498.3365497.32932416.24309309H'43-Epidehydrotumulosic acid486.3709486.3748483.34911.12.316.39452452H'3Dehydrotumulosic acid486.3709486.3745231.13870.83.416.63386386H'3Atractylenolide A230.13068230.1315231.13870.83.416.61338338H'23B-Hydroxy-114.122* epoxylean-28-138-olide486.3349485.32962.44.816.61338338H'4Poricoic acid C482.33961482.3401481.33280.40.916.77353353H'0Pachymic acid528.3817528.3828527.37561.42.617.67485485H'2Pachymic acid136.1252136.128137.1310.64.418.1810951095H'14Poricoic acid C482.3496193.1109194.11820.73.518.5526822682H'<		500.35017	500.3498	499.3425	-0.4	-0.8	15.23	1282	955	H.	3
3B-hydroxy-atractylon232.14633232.1473233.15450.9416.0370816265H'43B,16a-Dihydroxylauroster 8,24-dien-21-oic acid472.3552472.356471.34870.81.616.2217217H'3Poricoic acid F498.33452498.3365497.32932416.24309309H'43-Epidehydrotumulosic acid484.3552484.3564483.34911.12.316.39452452H'3Dehydrotumulosic acid486.37091486.3715231.13870.83.416.53438436H'3Atractylenolide A230.1306230.1315231.13870.83.416.53438438H'23B-Hydroxy-110,12ar epoxyolean-28-138-olide486.33452486.3369485.32962.44.816.61338338H'2Ginger brain276.17254276.1732277.18050.72.516.742703648H'4Poricoic acid C482.33961482.3401481.33280.40.916.77353353H'1B-Red myrcene204.1878204.1885205.19580.73.617.67485485H'2B-aqua-anisidine136.1252136.1258137.1310.64.418.1810951095H'14Salsolinol193.1102193.1109194.11820.73.518.55 </td <td>Poricoic acid B</td> <td>484.31887</td> <td>484.3196</td> <td>483.3123</td> <td>0.7</td> <td>1.4</td> <td>15.25</td> <td>217</td> <td>217</td> <td>H.</td> <td>0</td>	Poricoic acid B	484.31887	484.3196	483.3123	0.7	1.4	15.25	217	217	H.	0
3B, 16a - Dihydroxylauroster- 8,24-dien-21-oic acid472.35526472.355471.34870.81.616.2217217H3Poricoic acid F498.33452498.3365497.32932416.24309309H43-Epidehydrotumulosic acid484.35526484.3564483.34911.12.316.39452452H3Dehydrotumulosic acid486.37091486.3718485.36450.91.816.48386386H3Atractylenolide A230.13068230.1315231.13870.83.416.53438438H'23B-Hydroxy-11a,12a- epoxyolean-28-138-olide486.33452486.3369485.32962.44.816.61338338H'2Ginger brain276.17254276.1732277.18050.72.516.742703648H'4Poricoic acid C482.33961482.3401481.33280.40.916.77353353H'1B-Red myrcene204.1878204.1885205.19580.73.617.67485485H'2B-aqua-anisidine136.1252136.1258137.1310.64.418.1810951095H'14Salsolinol193.11028193.1109194.11820.73.518.5526822682H'2Eburicoic acid470.376470.3781493.36742.14.318.6<	Hydroxy-atractylenolide	248.14124	248.1425	247.1352	1.2	5	15.26	222	222	H.	0
8,24-dien-21-oic acid472.3352472.3352472.3356477.34870.81.616.2217217H3Poricoic acid F498.33452498.3365497.32932416.24309309H43-Epidehydrotumulosic acid484.35526484.3564483.34911.12.316.39452452H3Dehydrotumulosic acid486.37091486.3718485.36450.91.816.48386386H3Atractylenolide A230.13068230.1315231.13870.83.416.53438438H'238-Hydroxy-11a,12a- epoxyolean-28-138-olide486.33452486.3369485.32962.44.816.61338338H'2Ginger brain276.17254276.1732277.18050.72.516.742703648H'4Poricoic acid C482.33961482.3401481.33280.40.916.77353353H'1B-Red myrcene204.1878204.1885205.19580.73.617.67485485H'2B-aqua-anisidine136.1252136.1258137.13310.64.418.1810951095H'14Salsolinol193.1102193.1109194.11820.73.518.5526822682H'2Eburicoic acid470.376470.3781493.36742.14.318.664664	3B-hydroxy-atractylon	232.14633	232.1473	233.1545	0.9	4	16.03	7081	6265	H⁺	4
3-Epidehydrotumulosic acid484.35526484.3564483.34911.12.316.39452452H ⁻ 3Dehydrotumulosic acid486.37091486.3718485.36450.91.816.48386386H ⁻ 3Atractylenolide A230.1306230.1315231.13870.83.416.53438438H ⁻ 23B-Hydroxy-11a,12a- epoxyolean-28-13B-olide486.33452486.3369485.32962.44.816.61338338H ⁻ 2Ginger brain276.17254276.1732277.18050.72.516.742703648H ⁻ 4Poricoic acid C482.33961482.3401481.33280.40.916.77353353H ⁻ 1B-Red myrcene204.1878204.1885205.19580.73.617.67485485H ⁻ 2B-aqua-anisidine136.1252136.1258137.13310.64.418.1810951095H ⁻ 14Salolinol193.11028193.1109194.11820.73.518.5526822682H ⁺ 2Eburicoic acid470.376470.3781493.36742.14.318.6646646Na ⁺ 46463.613.612.622.624015.624022.55.2360.62.418.821567413670H ⁺ 7		472.35526	472.356	471.3487	0.8	1.6	16.2	217	217	Η [.]	3
Dehydrotumulosic acid486.37091486.3718485.36450.91.816.48386386H°3Atractylenolide A230.13068230.1315231.13870.83.416.53438438H°236-Hydroxy-11α,12α- epoxyolean-28-138-olide486.33452486.3369485.32962.44.816.61338338H°2Ginger brain276.17254276.1732277.18050.72.516.742703648H°4Poricoic acid C482.33961482.3401481.33280.40.916.77353353H°1B-Red myrcene204.1878204.1885205.19580.73.617.08431431H°0Pachymic acid528.38147528.3828527.37561.42.617.67485485H°2B-aqua-anisidine136.1252136.1258137.13310.64.418.1810951095H°14Salsolinol193.11028193.1109194.11820.73.518.5526822682H°2Eburicoic acid470.376470.3781493.36742.14.318.6646646Na*4Hexadecane256.24023256.2408255.23360.62.418.821567413670H°7	Poricoic acid F	498.33452	498.3365	497.3293	2	4	16.24	309	309	H.	4
Atractylenolide A230.13068230.1315231.13870.83.416.53438438H ³ 23B-Hydroxy-11α,12a- epoxyolean-28-13B-olide486.33452486.3369485.32962.44.816.61338338H ³ 2Ginger brain276.17254276.1732277.18050.72.516.742703648H ⁴ 4Poricoic acid C482.33961482.3401481.33280.40.916.77353353H ⁴ 1B-Red myrcene204.1878204.1885205.19580.73.617.08431431H ⁴ 0Pachymic acid528.38147528.3828527.37561.42.617.67485485H ⁴ 2B-aqua-anisidine136.1252136.1258137.13310.64.418.1810951095H ⁴ 14Salsolinol193.11028193.1109194.11820.73.518.5526822682H ⁴ 2Eburicoic acid470.376470.3781493.36742.14.318.6646646Na ⁴ 4Hexadecane256.24023256.2408255.23360.62.418.821567413670H ⁴ 7	3-Epidehydrotumulosic acid	484.35526	484.3564	483.3491	1.1	2.3	16.39	452	452	H.	3
3B-Hydroxy-11a,12a- epoxyolean-28-13B-olide 486.33452 486.3369 485.3296 2.4 4.8 16.61 338 338 H ⁻ 2 Ginger brain 276.17254 276.1732 277.1805 0.7 2.5 16.7 4270 3648 H ⁺ 4 Poricoic acid C 482.33961 482.3401 481.3328 0.4 0.9 16.77 353 353 H ⁺ 1 B-Red myrcene 204.1878 204.1885 205.1958 0.7 3.6 17.08 431 431 H ⁺ 0 Pachymic acid 528.38147 528.3828 527.3756 1.4 2.6 17.67 485 485 H ⁺ 2 B-aqua-anisidine 136.1252 136.1258 137.1331 0.6 4.4 18.18 1095 1095 H ⁺ 14 Salsolinol 193.11028 193.1109 194.1182 0.7 3.5 18.55 2682 2682 H ⁺ 2 Eburicoic acid 470.376 470.3781 493.3674 2.1 4.3 18.62 15674 13670	Dehydrotumulosic acid	486.37091	486.3718	485.3645	0.9	1.8	16.48	386	386	H.	3
epoxyolean-28-13B-olide486.33452486.3369485.32962.44.816.61338338H2Ginger brain276.17254276.1732277.18050.72.516.742703648H*4Poricoic acid C482.33961482.3401481.33280.40.916.77353353H*1B-Red myrcene204.1878204.1885205.19580.73.617.08431431H*0Pachymic acid528.38147528.3828527.37561.42.617.67485485H*2B-aqua-anisidine136.1252136.1258137.13310.64.418.1810951095H*14Salsolinol193.11028193.1109194.11820.73.518.5526822682H*2Eburicoic acid470.376470.3781493.36742.14.318.6646646Na*4Hexadecane256.24023256.2408255.23360.62.418.821567413670H*7	Atractylenolide A	230.13068	230.1315	231.1387	0.8	3.4	16.53	438	438	H⁺	2
Poricoic acid C482.33961482.3401481.33280.40.916.77353353H [·] 1B-Red myrcene204.1878204.1885205.19580.73.617.08431431H [·] 0Pachymic acid528.38147528.3828527.37561.42.617.67485485H [·] 2B-aqua-anisidine136.1252136.1258137.13310.64.418.1810951095H [·] 14Salsolinol193.11028193.1109194.11820.73.518.5526822682H [·] 2Eburicoic acid470.376470.3781493.36742.14.318.6646646Na ⁺ 4Hexadecane256.24023256.2408255.23360.62.418.821567413670H [·] 7		486.33452	486.3369	485.3296	2.4	4.8	16.61	338	338	Η [.]	2
B-Red myrcene204.1878204.1885205.19580.73.617.08431431H ⁺ 0Pachymic acid528.38147528.3828527.37561.42.617.67485485H ⁺ 2B-aqua-anisidine136.1252136.1258137.13310.64.418.1810951095H ⁺ 14Salsolinol193.11028193.1109194.11820.73.518.5526822682H ⁺ 2Eburicoic acid470.376470.3781493.36742.14.318.6646646Na ⁺ 4Hexadecane256.24023256.2408255.23360.62.418.821567413670H ⁺ 7	Ginger brain	276.17254	276.1732	277.1805	0.7	2.5	16.7	4270	3648	H⁺	4
Pachymic acid528.38147528.3828527.37561.42.617.67485485H°2B-aqua-anisidine136.1252136.1258137.13310.64.418.1810951095H°14Salsolinol193.11028193.1109194.11820.73.518.5526822682H°2Eburicoic acid470.376470.3781493.36742.14.318.6646646Na*4Hexadecane256.24023256.2408255.23360.62.418.821567413670H°7	Poricoic acid C	482.33961	482.3401	481.3328	0.4	0.9	16.77	353	353	H.	1
B-aqua-anisidine 136.1252 136.1258 137.1331 0.6 4.4 18.18 1095 1095 H ⁺ 14 Salsolinol 193.11028 193.1109 194.1182 0.7 3.5 18.55 2682 2682 H ⁺ 2 Eburicoic acid 470.376 470.3781 493.3674 2.1 4.3 18.6 646 646 Na ⁺ 4 Hexadecane 256.24023 256.2408 255.2336 0.6 2.4 18.82 15674 13670 H ⁺ 7	B-Red myrcene	204.1878	204.1885	205.1958	0.7	3.6	17.08	431	431	H⁺	0
Salsolinol 193.11028 193.1109 194.1182 0.7 3.5 18.55 2682 2682 H ⁺ 2 Eburicoic acid 470.376 470.3781 493.3674 2.1 4.3 18.6 646 646 Na ⁺ 4 Hexadecane 256.24023 256.2408 255.2336 0.6 2.4 18.82 15674 13670 H ⁻ 7	Pachymic acid	528.38147	528.3828	527.3756	1.4	2.6	17.67	485	485	H.	2
Eburicoic acid 470.376 470.3781 493.3674 2.1 4.3 18.6 646 646 Na ⁺ 4 Hexadecane 256.24023 256.2408 255.2336 0.6 2.4 18.82 15674 13670 H ⁻ 7	B-aqua-anisidine	136.1252	136.1258	137.1331	0.6	4.4	18.18	1095	1095	H⁺	14
Hexadecane 256.24023 256.2408 255.2336 0.6 2.4 18.82 15674 13670 H ⁻ 7	Salsolinol	193.11028	193.1109	194.1182	0.7	3.5	18.55	2682	2682	H⁺	2
	Eburicoic acid	470.376	470.3781	493.3674	2.1	4.3	18.6	646	646	Na⁺	4
γ-Sitosterol 414.38617 414.3875 453.3506 1.3 2.9 18.88 1913 1913 K ⁺ 1	Hexadecane	256.24023	256.2408	255.2336	0.6	2.4	18.82	15674	13670	Η [.]	7
	γ-Sitosterol	414.38617	414.3875	453.3506	1.3	2.9	18.88	1913	1913	K⁺	1



Fig. 11: Cell viability after treatment with the different concentrations of ZWD and 1 mM FFA, **p<0.01 and ****p<0.0001 vs. control group



Fig. 12: Lipid concentration of HepG2 cells treated with the different concentrations of ZWD and 1 mM FFA, *p<0.05 and **p<0.01 vs. model group; ##p<0.01 vs. control group. All experiments were repeated at least two times

The results of cell-based assays showed that 1 mM FFA could significantly induce lipid accumulation in the HepG2 cells. After the intervention of ZWD, cell viability enhanced, which might be due to the effect of Fuzi because it significantly promote cell viability even in low concentration^[25]. The pathogenesis of NAFLD is a 'three-hit' process, including steatosis, lipotoxicity and inflammation, which impair the hepatocyte viability, eventually leading to death and exacerbation of NAFLD^[26]. Therefore, it was speculated that the improvement of cell vitality by ZWD might be the mechanism of its therapeutic effect. The intracellular lipid contents were decreased by ZWD in a concentration-dependent manner, where 19.5 mg/ml (p \leq 0.05) and 39 mg/ml (p \leq 0.01) of ZWD could reduce lipid accumulation. For validation purposes, RTqPCR was performed to test the expression of EGFR, PPARG, HSP90AA1, MAPK1 and PI3K. RT-qPCR experiments proved ZWD induced down-regulation of EGFR mRNA levels and up-regulation of HSP90AA1, MAPK1 and PI3K mRNA levels.

The network pharmacology results identified potential key targets and pathways of ZWD active against NAFLD. In order to verify the reliability of network pharmacological prediction results, we detected the mRNA level of these key genes by qRT-PCR. As shown in fig. 13, RT-qPCR results showed that mRNA expression of EGFR and PPARG was increased in the model group compared with that in the control group, and after three concentration gradient of ZWD treated for 24 h, the expression of EGFR mRNA was decreased, the expression of PPARG mRNA showed that there was an inconsistent change trend between high concentration and low concentration. The mRNA expression of HSP90AA1, MAPK1 and PI3K was decreased in the model group compared with that in the control group, and after treatment, the mRNA expression of HSP90AA1, MAPK1 and PI3K was increased.



Fig. 13: The expression of NAFLD-related genes after treatment with ZWD, *p<0.05, **p<0.01 and ***p<0.001 vs. model group and #p<0.05, ##p<0.01 vs. control group

The results of our compound-target-pathway suggested that the active ingredients in ZWD could interact with the PI3K-AKT signaling pathway through EGFR, HSP90AA1 and MAPK1. The activated PI3K/AKT signaling pathway can exert anti-inflammatory, anti-oxidative stress, anti-apoptotic and autophagic regulatory effects through downstream pathways and related proteins^[27], these processes are closely related to the pathogenesis of NAFLD. MAPK1 has been shown to be involved in cellular processes of autophagy, lipid metabolism, proliferation, migration^[28] and the activation of MAPK 1/3 ameliorates liver steatosis in leptin receptor-deficient mice^[29]. The function of HSP90AA1 is to promote the maturation, structural maintenance and proper regulation of specific target proteins involved in cell cycle control and signal transduction^[30]. Thus, our findings show that ZWD can positively regulate NAFLD in multiple ways through active ingredients and targets.

In summary, this study demonstrated a strategy to optimize conventional network pharmacology and explained the molecular mechanism of ZWD in NAFLD treatment was closely associated with 5 core genes (including AKT1, EGFR, CASP3, PPARG and IGF1R), which were involved in important signal pathways (including FOXO signaling pathway and PI3K-AKT signaling pathway). RT-qPCR analyses revealed that the therapeutic effect of ZWD might be achieved *via* down-regulation of EGFR mRNA levels and up-regulation of HSP90AA1, MAPK1 and PI3K mRNA levels.

Author's contributions:

Chun-Jiang Zhang and Zhi-Yan Fang contributed equally to this work. Tian-e Zhang, Yan-Qiu Wang and Chun-Jiang Zhang: Conceptualization; Chun-Jiang Zhang, Zhi-Yan Fang, Zheng Luo, Hai-Yan Zhu and Lili Huang: Methodology and visualization; Wen-Ying Huai and Yan-Qiu Wang: Software; Lili Huang and Yan-Qiu Wang: *In vitro* cells-based experiment; Chun-Jiang Zhang and Tian-e Zhang: Writing-original draft; Tian-e Zhang and Yu-Qin Tang: Writing-review and editing; Tian-e Zhang and Qiao-Zhi Yin: Supervision and project administration. All authors have read and agreed to the published version of the manuscript.

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Conflict of interests:

The authors declared that there are no conflicts of interest regarding the publication of this paper.

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