Therapeutic Potential of Pomegranate (*Punica granatum* Linn.) against Breast Cancer

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Breast cancer ranks as the second major cause for cancer-related female mortalities worldwide. The features such as family history, old age, late-aged menopause, early-aged menarche, estrogen-replacement treatments for longer periods and first childbirth at later-age are common breast cancer related risk factors. Studies have shown that pomegranate (*Punica granatum* Linn.) and its constituents can efficiently affect multiple signaling pathways involved in inflammation, cellular transformation, hyperproliferation, angiogenesis, initiation of tumorigenesis and eventually suppressing the final steps of tumorigenesis and metastasis. Pomegranate extracts have been reported to significantly downregulate the proliferation and expression of transcription factors or proteins which are involved in breast cancer cells. These activities may lead to the up-regulation of protective factors or suppression of the gene expression, receptors as well as regulatory factors and may be associated with the suppression of the development of new blood vessels as well. In this review we discussed the therapeutic role of pomegranate against breast cancer. In addition, we revised the bioactive constituents of pomegranate, nanoencapsulation of these compounds and its safety on health.

Key words: Breast cancer, Punica granatum, nanoencapsulation, bioactive compounds

Cancer was the second leading cause of death, after heart disease, in the United States in 2020. In 2020, there were 602 350 cancer deaths; 284 619 were among females and 317 731 among males^[1]. Breast cancer ranks as the second major reason for cancerrelated female mortalities^[2]. The features such as family history, old age, late-aged menopause, earlyaged menarche, estrogen-replacement treatments for longer period and first childbirth at later-age are common breast cancer related risk factors. Steroid hormones, especially estrogens are considered to facilitate the occurrence of breast cancer^[3,4]. Pomegranate (Punica granatum Linn.) is obtained from a deciduous tree belonging to the family Lythraceae^[5,6]. Pomegranate has been used in various medicinal systems of medicine for the treatment and therapy of a multitude of diseases and ailments^[7]. Studies have shown that pomegranate and its constituents can efficiently affect multiple signaling pathways involved in inflammation, cellular transformation, hyperproliferation, angiogenesis, initiation of tumorigenesis and eventually suppressing the final steps of tumorigenesis and metastasis^[7,8].

Pomegranate extracts have been reported to significantly decrease the Specificity protein (Sp) (Sp1, Sp3, Sp4) and micro Ribonucleic acid (miR)-27a and miR-155 in breast cancer cells. Sp1, Sp3, Sp4 transcription factors are widely overexpressed in multiple types of cancers^[9] and regulate survival, angiogenesis and proliferation-related genes. Sp1 regulates Nuclear Factor kappa B (NF- κ B) in the NF- κ B p65 subunit's promoter region through a Guanine-Cytosine (GC)-rich binding site^[10].

The downregulation of Sp transcription factors could further mediate NF- κ B downregulation by pomegranate extracts. The miR-27a based suppression of Sp-repressor Zinc Finger and BTB Domain containing 10 (ZBTB10) results in higher expression of Sp transcription factors in breast cancer cells. Several anticancer agents are known to downregulate miR-27a while up-regulating the ZBTB10^[11]. Similarly, pomegranate extracts increased the transcriptional repressor ZBTB10 expression^[10] along with the Src Homology-2 Domain-containing Inositol 5-Phosphatase 1 (SHIP1) expression, which is a target gene of miR155. This lipid phosphatase negatively regulates the Phosphatidylinositol-3,4,5-Trisphosphate (PI3K)/Protein Kinase B (Akt) signaling pathway, which is commonly activated in different tumors and promotes cell survival and proliferation^[10,11]. These findings reveal that the decreased levels of miR-27a and miR-155 could partially be responsible for the anti-cancer impact of pomegranate extract on breast cancer cells^[10].

This review provides a comprehensive review on the anti-breast cancer activities of pomegranate in the treatment and prevention of breast cancer.

POMEGRANATE AND ITS CONSTITUENTS

Pomegranate belongs to the genus *Punica* and family Lythraceae. Pomegranate is native in Asian countries including Iran to Northern India. It grows in many of the Middle Eastern countries and was originally introduced from Syria and Israel since around 1600 Before the Common Era (BCE)^[12].

Studies have demonstrated that pomegranates may be used as natural remedy due to their capability against a wide variety of diseases. Almost all the parts of pomegranate, including its flowers, peel, juice, arils and bark has shown a good biological activity. There are wide ranges of phytochemical properties which have demonstrated antimicrobial activities in pomegranate. The most active ingredients in pomegranate are Ellagic Acid (EA) and hydrolysable tannins, like Punicalagin (PU)^[13]. Anticancer and anti-inflammatory activity has been found in the juice of the pomegranate as well as in its peel and oil resulting in antiproliferative effect, cell cycle arrest, reduction in invasion and angiogenesis. The wide variety of clinical administration of pomegranate to treat and prevent cancer and inflammation is due its phytochemistry and pharmacological properties^[14].

Published papers have revealed that pomegranate may be beneficial in the treatment and prevention of many types of cancer like breast cancer, lung cancer, prostate cancer and skin cancer. It is thought that drinking Pomegranate Juice (PJ) will provide protection from cancer because it contains high levels of antioxidants as well as other nutrients^[15].

Pomegranate reduces the proliferation of cancerous cells and induces programmed cell death, also slows down the blood supply to tumors, resulting in their starvation and shrinking^[16]. Pomegranate grows as shrub reaching in height between 1.5 to 5 m, with glossy leaves and thorny and irregular branches appearing as evergreen in frigid regions and as a deciduous shrub in temperate ones^[5].

The Punicaceae family has only one genus and two species, including: The edible *Punica granatum* and the inedible one *Punica protopunica*^[5]. The different local names given to pomegranate are presented in fig. 1^[5].



Fig. 1: Vernacular names of pomegranate tree in different countries

Pomegranate contains a variety of phenolic compounds like Ellagitannins (ETs) (punicalin, pedunculagin, PU, EA, ET, gallagic acid and gallotannins), anthocyanins (delphinidin, cyanidin and pelargonidin glycosides), flavonoids (quercetin, kaempferol and luteolin glycosides) and phenolic acids like chlorogenic acid, caffeic, o-coumaric acid, rutin, p-coumaric acid, ferulic acid, vanillic, phloridzin and syringic acid. The most abundant polyphenol with high molecular weight is PU. A huge amount of this polyphenols like gallic acid, protocatechuic acid, chlorogenic acid, caffeic acid, ferulic acid, ferulic acid, caffeic acid, ferulic acid, coumaric acids and catechin^[17].

The pomegranate fruit itself without the peel is not toxic, while its bark and roots are toxic. A hydroalcoholic extract may induce genotoxicity at various levels of expression like clastogenic, mutagenic and recombinogenic^[18] (fig. 2 and Table $1^{[19-27]}$).

EFFECTS OF POMEGRANATE COMPONENTS ON BREAST CANCER

As explained above, pomegranate comprised several biochemicals which have different biomedical activities. The effects of these constituents on breast cancer mostly investigated by the applications of extracts from different parts of pomegranate fruit or chromatographically purified juice fractions^[21]. However, recent studies have employed either as single or a combination of components targeting breast and other cancer types. In this line, one study deduced the promising anti-cancer potential of three pomegranate components (Punicic acid (P), Luteolin (L) and Ellagic acid (E)) against breast cancer and prostate cancer cells (fig. 3).



Fig. 2: Schematic illustration of the main biochemical constituents of different parts of pomegranate tree (peel, juice, flower, leave, seed)



Fig. 3: Schematic diagram showing the mechanism of action of anti-breast cancer activity of the pomegranate extract. The biochemical mainly exert effects on the process of metastasis or the proliferation of breast cancer

TABLE 1: BIOCHEMICAL COMPONENTS OF POMEGRANATE AND ITS MAIN ANTI-BREAST CANCER ACTIVITIES

Category	Phytochemical	Chemical structure	Main anti-breast cancer activities	References
Anthocyanins and anthocyanidins	Gallagic acid		Induces apoptosis and ferroptosis resulting in programmed cell death	[19]
EA derivatives	EA		Direct inhibition of Alpha-Actinin 4 (ACTN4)	[20]
Anthocyanins and anthocyanidins	Punicic acid (Omega-5 fatty acid)	Он	Apoptosis induction <i>via</i> PKC pathway and based on lipid peroxidation	[21]
ETs and gallotannins	Pedunculagin		Inhibition of aromatase activity, proapoptic effect, inhibition of migration, invasion and metastasis	[22]
Anthocyanins and anthocyanidins	Delphinidin 3-O-beta-D- glucoside (Myrtillin)		Inhibition <i>via</i> Akt/ HOX Antisense Intergenic RNA (HOTAIR) signaling pathway inactivation	[23]
Anthocyanins and anthocyanidins	Cyanidin 3-glucoside		Attenuates the angiogenesis of breast cancer via inhibiting Signal Transducer and Activator of Transcription 3 (STAT3)/VEGF pathway	[24]
Anthocyanins and anthocyanidins	Pelargonidin 3-glucoside		Affect the production of selected pro and anti-inflammatory cytokines	[25]
Anthocyanins and anthocyanidins	Pelargonidin 3,5-diglucoside		Affect the production of selected pro and anti-inflammatory cytokines	[25]
Anthocyanins and anthocyanidins	Cyanidin 3,5-diglucoside	HO HO HO HO HO	Inhibition of the STAT3/ VEGF pathway and angiogenesis	[26]
Anthocyanins and anthocyanidins	Delphinidin 3,5-diglucoside	HO OH OH OH	Mediated <i>via</i> inhibition of Fyn kinase activity and the TNF-α-induced Cyclooxygenase-2 (COX-2) expression	[27]

Anti-breast cancer assessment was performed using estrogen-sensitive Michigan Cancer Foundation-7 (MCF7) cells of Estrogen Receptor positive (ER⁺), estrogen-insensitive Human Breast Cancer cell line (MDA-MB-231) cells of Estrogen Receptor negative (ER-) and a non-neoplastic cell lines like human Breast Epithelial cell line (MCF10A). The combination of L+E+P (equal amounts) not only inhibited breast cancer cell growth but also reduced cancer cell migration and enhanced cell adhesion without affecting the normal cells^[28]. L+E+P also increased the cell adhesion-related genes expressions including Intercellular Adhesion Molecule 1 (ICAM1) and Claudin 1 (CLDN1) and decreased cell migration-related genes expression including Hyaluronan-Mediated Motility Receptor (HMMR). These components also inhibited cancer cells chemotaxis to C-X-C Motif Chemokine Ligand 12 (CXCL12), which is a chemokine involved in the metastasis of breast cancer^[29]. Adhesion loss and rise in CXCL12 chemotaxis and cell migration are known to cause breast cancer metastasis. During this study, the combination of L+E+P enhanced the E-cadherin expression whereas it contrarily alleviated the two Twist-Related protein (TWIST) genes expressions, which participate in epithelialto-mesenchymal transitions. Small interfering Ribonucleic Acid (siRNA)-based knockdown of E-cadherin or overexpression of TWIST partially reversed L+E+P inhibitory effect on the cancer cell migration. Moreover, cytokine/chemokine multiplex arrays demonstrated a significant reduction in the pro-inflammatory cytokines/chemokines (Regulated upon Activation, Normal T Cell Expressed and Secreted (RANTES and Interleukin-8 (IL-8)) levels in response to L+E+P application. This phenomenon confirms their potential of decreasing inflammation to ultimately affect cancer progression^[30]. Multiple studies have confirmed the anti-cancer potential of luteolin, EA and punicic acid against breast cancer cells. Luteolin is known to induce dose-dependent suppression of MCF-7 cell proliferation and Insulinlike Growth Factor 1 (IGF-1)^[31].

Luteolin-based significant reduction of Akt phosphorylation and IGF-1-dependent IGF-1 Receptor (IGF-1R) has also been reported. ER is supposed to directly participate in the luteolinbased inhibition of IGF-1-induced cell proliferation. Luteolin also significantly reduced ER expression. ER-specific siRNA-based ER knockdown in MCF-7 cells reduced the luteolin inhibition efficacy on IGF-1-induced cell proliferation. Therefore, it can be deduced that ER is probably the molecular target of luteolin. The results further demonstrated that luteolin inhibitory effects on MCF-7 cell growth occur via IGF-1-mediated PI3K-Akt pathway inhibition. The antiangiogenesis impact of EA on breast cancer has been established^[32]. EA could significantly inhibit the Vascular Endothelial Growth Factor (VEGF)induced migration, cell proliferation and tube formation in human endothelial cells. Furthermore, the EA could also restrict the Vascular Endothelial Growth Factor Receptor 2 (VEGFR2) tyrosine kinase activity and its downstream signaling pathways (PI3K/Akt and Mitogen-Activated Protein Kinase (MAPK)) in endothelial cells. The studies have also reported a reduction in phospho VEGFR2 levels and inhibition of MDA-MB-231 xenograft tumor growth by EA. Molecular docking simulation based on in silico analysis indicated the interaction of EA with the Adenosine Triphosphate (ATP) binding region of VEGFR-2. These findings depict that EA could target the VEGFR-2 signaling pathway in breast cancer to exert anti-angiogenesis effects^[28].

Punicic acid is the most abundant seed oil ingredient, which is known to possess anti-cancer properties against breast cancer cells. Punicic acid could inhibit the proliferation of MDA-MB-231 and MDA-ER alpha (α) 7 cell lines, which are known as estrogeninsensitive and estrogen-sensitive breast cancer cell lines, respectively^[21]. Punicic acid could further induce the MDA-MB-231 and MDA-ERa7 cell apoptosis and disrupt the cellular mitochondrial membrane potential. The investigations have also revealed the importance of lipid oxidation for the apoptosis and cell proliferation effects of punicic acid. However, Protein Kinase C (PKC) inhibitors could partially block these effects of punicic acid in MDA-MB-231 and MDA-ERa7 cells. These findings establish the PKC pathway and lipid oxidation-based anticancer effects of punicic acid.

Pomegranate extract has generated cytotoxicity and anti-inflammatory properties in breast cancer cells during *in vivo* and *in vitro* investigations. The anti-breast cancer properties of pomegranate extract could partially depend upon the targeting of microRNAs^[33,34]. Pomegranate Seed Oil (PSO) has prevented different types of tumor cell proliferation^[34]. It has reduced mammary carcinogenesis in the mouse as well. Jeune *et al.*^[35] have reported MCF-7 breast cancer cell growth inhibition by pomegranate extracts through the process called apoptosis. Angiogenesis is important for the nutrient and oxygen supply during tumor metastasis and growth whereas pomegranates could suppress the development of new blood vessels which supply the cancer cells^[36].

Kim *et al.*^[37] have reported the inhibitory effects of pomegranate components on various types of breast cancers. They further suggested a potential role of pomegranate in cancer suppression and treatment. During another study, Shirode *et al.* deduced that the encapsulation of pomegranate polyphenols could enhance their bio-efficacy in inhibiting the growth of breast cancer cells (Aneuploid Mammary Epithelial cells (Hs578T) and MCF-7)^[34].

Recently, the link between breast cancer risk and dietary factors has been a major focus of investigations and several studies have revealed beneficial aspects of pomegranates in countering breast cancer^[38]. Kim et al.^[37] have reported aromatase inhibition by the polyphenols from the pericarp, PSO and fermented PJ. Aromatase is important for breast carcinogenesis as it converts androgen into estrogen^[39]. The inhibition of estrogen biosynthetic enzyme (17-beta (β)-hydroxysteroid dehydrogenase) by pericarp, PSO and fermented PJ-derived polyphenols has been established^[40]. In addition to the anti-estrogenic effects, pericarp, PSO and fermented PJ-derived polyphenols could also inhibit the growth of MB-MDA-231 and MCF-7 breast cancer cell lines. Fermented PJ-derived polyphenols have also been revealed to inhibit the formation 7,12-Dimethylbenz[a]anthracene of (DMBA)induced cancerous lesions in the mammary gland organ culture of a murine^[37]. Adams et al.^[41] have demonstrated anti-aromatase and anti-proliferative activities in breast cancer cells in response to the application of pomegranate ETs-derived compounds. During a study, pomegranate ET-derived compounds including gallagic acid, EA and urolithins A and B (sulfated, methylated and acetylated analogs) were subjected to live cell-based assay and placental microsome aromatase assay to investigate their antiaromatase activity. Methylated urolithin A, urolithin A, methylated urolithin B, urolithin B, urolithin B sulfate, acetylated urolithin B and gallagic acid caused a significant inhibition of aromatase activity in placental microsomes. Further comparison of these compounds using aromatase over-expressing cell line (MCF-7aro) presented urolithin B as the most potent inhibitor of aromatase. Aromatase activity was significantly inhibited at urolithin B doses of 2.35 μ M (p≤0.05) and 4.7 μ M (p≤0.01). Gallagic

acid also exerted significant anti-aromatase activity at a dose of 4.7 μ M (p \leq 0.01). Urolithins were further tested to assess their efficacy against testosteroneinduced cell proliferation. Urolithin B was noted to successfully inhibit testosterone-induced cell proliferation followed by gallagic acid^[41]. These data suggest that pomegranate intake could be beneficial for the chemoprevention of breast cancer.

Pomegranate Pericarp Methanolic Extract (PME) is known to exhibit Selective Estrogen Receptor Modulator (SERM) activity in in vivo estrogen deprivation models and human breast cancer cell lines^[42]. SERMs are the ligands for ER and could exhibit antagonist or agonist functions depending upon the tissue type. SERMs are commonly employed in estrogen-dependent breast cancer therapies. PME treatment caused a significant dosedependent cell growth inhibition in the ER⁺ MCF-7 cell line. However, PME was unable to affect the ER- MDA MB-231 cell proliferation. 17β-estradiolinduced MCF-7 cell proliferation was also inhibited by PME treatment and furthermore, PME downregulated the expression of estrogen-responsive genes (Progesterone Receptor (PR), pS2 and ERa) in MCF-7 cells. The lack of PME estrogenicity was finally confirmed in Ovariectomized (OVX) mice by assessing the epithelial heights and uterine wet weights as the estrogenicity markers. 17β-estradiol increased the normalized and absolute uterine wet weight by almost two times in OVX animals. However, uterus weights were not significantly different among PME-treated and vehicle-treated OVX control groups. Thus, these findings confirmed the lack of PME estrogenicity in the uterine endometrium. Similarly, uterine histology depicted that even though the 17β -estradiol induced uterine epithelium proliferation but luminal epithelial proliferation was not observed in PME-treated OVX mice^[43]. Rocha et al.^[44] further tested PJ and its components to assess their efficacy against breast cancer metastasis-related various processes. They used two breast cancer cell lines (MCF-7 (ER⁺) and MDA-MB-231 cells (ER⁻)) and a non-neoplastic cell line (MCF10A). The results demonstrated that PJ or a combination of its components (punicic acid, luteolin and EA) decreased cancer cell migration and growth, and increased the adhesion of cancer cells, without affecting normal cells. PJ and its three components restricted pro-inflammatory cytokines/ also chemokines production in cancer cells. Interestingly, PJ and its components were found to promote the

cancer cell adhesion-related gene expressions, inhibit the genes related to cell migration and prevent cancer cell chemotaxis to stromal cell-derived factor 1α .

Several chemo-preventative investigations have highlighted the antioxidant and pro-apoptotic properties of Pomegranate Fruit Extract (PFE) and its components against breast cancer^[45]. Punicic acid, which is a polyunsaturated fatty acid of PSO has been reported to initiate the apoptosis in estrogen-insensitive and sensitive cell lines (MDA-ER-7 and MDA-MB-231) along with significant inhibition of cell growth^[46]. Caruso et al.^[47] have described PFE methanolic extract-based reduction in MCF-7 cell proliferation and a dose-dependent rise in cell apoptosis. These PFE properties could be related to the enhanced pro-apoptotic gene (Bcl-2 Associated X-Protein (BAX)) expression and a reduced anti-apoptotic gene (B-cell lymphoma-2 (Bcl-2)) expression. Costantini et al.[48] found a high abundance of punicic acid and its components in the hydrophilic PSO fraction (80 % aqueous methanol extract). They further evaluated their anti-inflammatory potential by employing breast cancer lines (MDA-MB-231 and MCF-7). The results demonstrated significantly, decreased cell viability of both cell lines after hydrophilic extract treatment with an increase in G0/G1 cell cycle phase as compared to untreated cells without significantly increased apoptosis in both cell lines. The results further revealed a decrease in the levels of proinflammatory cytokines (IL-17, IL-12, IL-6, IL-2, Monocyte Chemoattractant Protein-1 (MCP-1), CXCL10, Tumor Necrosis Factor alpha (TNF-α), Macrophage Inflammatory Protein (MIP)-1a and MIP-1 β) and VEGF at the higher doses of PSO hydrophilic extracts. Shirode et al.^[49] have studied PFE anti-breast cancer potential using MCF-7 cells by investigating the changes in gene expression at the whole genome level. PFE treatment reduced the MCF-7 cell proliferation, which altered the expressions of 903 genes (up-regulation of 505 genes and down-regulation of 398 genes). Most of the upregulated genes were related to apoptosis regulation whereas the down-regulated genes were involved in the chromosomal organization, mitosis, RNA processing, Deoxyribonucleic Acid (DNA) repair and DNA damage response. Genes including RAD50 Double Strand Break Repair Protein (RAD50), MRE11 Homolog, Double Strand Break Repair Nuclease (MRE11), Nibrin 1 (NBS1), MutS Homolog 6 (MSH6), RAD51 Recombinase (RAD51), BRCA1/

BRCA2-containing Complex Subunit 3 (BRCC3), Breast Cancer gene 1 (BRCA1) and Breast Cancer gene 2 (BRCA2) related to DNA repair and damage response were noted to be down-regulated^[49]. Chen et al.^[50] conducted a complementary DNA (cDNA) microarray-based investigation to understand the underlying molecular mechanisms of EA-induced MCF-7 cell growth inhibition. The results revealed that EA involves cell cycle arrest-based inhibition of breast cancer cell growth and proliferation. The alterations in genes belonging to the Transforming Growth Factor-beta (TGF-β)/Suppressor of Mothers against Decapentaplegic (SMAD) signaling pathway were found to regulate the EA-based cell cycle arrest in MCF-7 cells. TGF- β is considered as a highly potent tumor suppressor that acts by promoting apoptosis, differentiation and cell growth inhibition^[51].

Bishayee et al.^[52] performed a study to evaluate the chemopreventive efficacy of Pomegranate Emulsion (PE) oral administration DMBA-induced mammary tumorigenesis in female Sprague-Dawley rats. Reduced tumor incidence and cumulative tumor burden were noted in PE-administered rats in comparison to control rats. PE-treated tumors presented almost normal alveolar and ductal structures having uniform epithelial cells without hyperplasia whereas the tumors in control rats were characterized by extensive histological epithelial proliferation. The chemopreventive impact against DMBA-initiated mammary tumors mainly depended upon the PE-based induction of apoptosis and reduction in cell proliferation^[52]. The mechanism of PE chemopreventive potential was further evaluated during another study^[52,53]. The results depicted reduced ER- α and ER- β expressions, nuclear translocation of β-catenin and cytoplasmic accumulation in PEtreated tumors. These data deduced a PE-induced disruption of ER and Wingless-Related Integration Site $(Wnt)/\beta$ -catenin signaling pathways as the molecular basis of its chemopreventive impact against DMBA-inflicted rat mammary tumors.

NANOENCAPSULATION OF POMEGRANATE BIOACTIVE COMPOUNDS

Despite the known beneficial aspects, the issues such as short retention time, low systemic bioavailability and poor absorption of ETs and their metabolites might undermine their full chemo-preventive potential. For example, ETs such as PU could not be absorbed by the human body in intact form. They are hydrolyzed in the human intestinal tract to EA moieties, which are further converted to urolithins by colonic microbiota before absorption^[54]. EA mainly accumulates in the epithelial cells of the intestine and their absorption into the systemic circulation is limited^[55]. Therefore, after PJ consumption only low nanomolar range concentrations of free EA and urolithins are detected in human blood^[56]. The absorbed EA and urolithins also have short half-life due to rapid liver metabolism and urinary excretion^[57].

ETs encapsulation into biodegradable and biocompatible Nanoparticles (NPs) could overcome the drawbacks such as short half-life, gastrointestinal hydrolysis, low systemic bioavailability and poor absorption. Nanotechnology approaches have been applied in cancer therapies to reduce toxicity, promote selective tumor uptake and increase bioavailability and stability^[58,59]. Recently, these approaches are being applied to prevent cancer through dietary phytochemicals^[60]. These investigations have led to the development of a promising nanoencapsulationbased nano chemoprevention approach with better efficacy of bioactive food compounds. Paliwal et al.[61] have reported better efficacy of Epigallocatechin-3-Gallate (EGCG) nano prototypes from green tea, resveratrol from table grapes and curcumin from turmeric as compared to their free counterparts.

Poly Lactic-co-Glycolic Acid (PLGA) NPs are biodegradable, biocompatible and stable in biological fluids. They could ensure the sustainable release of loaded compounds by preventing their early degradation^[62]. Cells uptake PLGA NPs *via* clathrinmediated endocytosis and fluid-phase pinocytosis and they could rapidly exit the endo-lysosomes to enter the cytoplasm^[63,64]. Ester linkages of PLGA undergo enzymatic and spontaneous hydrolysis to generate glycolic acid and lactic acid. Glycolic acid and lactic acid are endogenous molecules, which can be easily metabolized into carbon dioxide and water through the Krebs cycle. PLGA polymer applications in humans are considered safe^[64].

The European Medicine Agency and United States (US) Food and Drug Administration have approved PLGA NPs applications through the parenteral route and the usage of PLGA microparticles as implants. Rapid opsonization of PLGA NPs by immunoglobulins and complement proteins is the main drawback. The reticuloendothelial system may clear PLGA NPs to restrict their approach to the target tissues. The modification of PLGA NPs surfaces with biocompatible polymers (Polyethylene Glycol (PEG)) could help in reducing opsonization to prolong their blood circulation time^[64].

Shirode *et al.*^[34] have studied the nanoencapsulation of pomegranate bioactive compounds using PLGA NPs for the chemoprevention of breast cancer (fig. 4). The data demonstrated that nanotechnologyenabled pomegranate polyphenols delivery enhanced the anti-breast cancer effects.



pomegranate extract

Fig. 4: Schematic diagram showing the nano encapsulation of pomegranate extract Note: PLGA-PEG: Poly Lactic-co-Glycolic Acid-Polyethylene Glycol; PVA: Polyvinyl alcohol and CH,Cl,: Dichloromethane

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SAFETY OF POMEGRANATE

Pomegranates are being used for centuries without adverse outcomes^[65]. Animal studies have not indicated pomegranate toxicity at the commonly used concentrations in traditional medicine^[66]. PJ extracts and oil could be used without risking the health of individuals. Pomegranate administrations (1420 mg/d extract tablets) have not produced adverse effects on the renal function or liver in humans^[67].

A significant ratio of antioxidants is found in pomegranate extracts. Therefore, pomegranatebased natural supplements are a suitable alternative for bioactive polyphenols^[68]. Bassiri-Jahromi *et al.*^[69] have recently administered 3 different doses of pomegranate peel extract to Bagg and Albino (BALB)/c mice and no toxicity was observed regarding the weight gain, food intake and biochemical or behavioral factors. These administrations did not disturb the biochemical parameters such as cholesterol, glucose, Aspartate Aminotransferase (AST) and Alanine Aminotransferase (ALT). Similarly, the inflammation was not observed at the supratherapeutic dose (7.5 mg/kg) as well.

CONCLUSION

In the current setting, cancer prevention *via* dietary agents is quite a promising arena of oncology that has drawn a significant amount of attention from both scientists in basic and clinical sciences and the general masses due to dietary agents have proven ability to prevent or suppress cancers, their low cost and easy availability.

However, current challenges relate to establish the key component of these dietetic agents which are responsible for the anticancer effects and the mechanisms through which they suppress cancer. Accumulating research provides extensive evidence related to biological activities of pomegranatederived products particularly with respect to their anticancer properties. Anti-breast cancer activity of pomegranate, reduced cell migration and inhibited chemotaxis, pomegranate extract may reduce cancer cell invasion and motility, both of which are required for metastasis. One study demonstrated that PFE resulted in dose-dependent inhibition of estrogen and PR-negative human breast cancer cell proliferation, invasion and motility, and inhibited breast cancer cell growth. The anti-cancer activity of pomegranate have been enhanced by using different types of carriers like nanoparticles, however, intensive research is needed in this area for better therapeutic effects.

Conflict of interests:

The authors declared no conflict of interest.

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